

Larvicidal Activity of Ethyl Acetate Extract of *Derris elliptica* Root

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Original Article

Larvicidal Activity of Ethyl Acetate Extract of *Derris elliptica* Root against the Third-instar Larvae of Cypermethrin-Resistant *Aedes aegypti* Offspring

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Abstract

Background: *Derris elliptica* extracts have a high larvicidal potential against the laboratory strain of *Aedes aegypti* larvae, but the effect on offspring larvae of pyrethroid-resistant strains of the species is lack understood. This study aimed to determine the larvicidal activity of the ethyl acetate extract of tuba root against the third-instar larvae of the Cypermethrin-resistant *Ae. aegypti* offspring.

Methods: The experimental study occupied four levels of ethyl acetate extract of *D. elliptica* namely 10, 25, 50, and 100 ppm, and each level was four times replicated. As many as twenty of healthy third-instar larvae, offspring of Cypermethrin-resistant *Ae. aegypti* were subjected to each experiment group. Larval mortality rate and lethal concentration 50% sublethal (LC₅₀) were calculated after 24 and 48 hours of exposure time.

Results: Mortality of larvae increased directly proportional to the increase of extract concentration. Larval mortality rates after 24 and 48 hours of exposure were 40–67.5% and 62.5–97.5%, and LC₅₀ were 34.945 and 6.461ppm, respectively.

Conclusion: The ethyl acetate extract of *D. elliptica* has the high effectiveness larvicidal potential against the third-instar larvae, offspring of the Cypermethrin-resistant *Ae. aegypti*. Isolation of the specific compound is necessarily done to obtain the active ingredient for larvicide formulation.

Keywords: Larvicidal activity; Ethyl acetate extract; *Derris elliptica* root; Cypermethrin resistant; *Aedes aegypti*

Introduction

The resistance of *Ae. aegypti* to several pyrethroid and organophosphate insecticide compounds such as deltamethrin, lambda cyhalothrin, cypermethrin, malathion, and temephos (1, 2) inhibits the public health action in eradicating the dengue vector, and intrigues researchers to find the other active ingredients as the alternatives. Natural chemical compounds (3), including *D. elliptica* roots (4), are interesting to study for several reasons including but not limited to readily degraded and there is no bioaccumulation in the environment (5). Researchers have proven that *D. elliptica* extracts have high larvicidal potential against the laboratory strain larvae of *Ae. aegypti* (3, 4, 6, 7). However, when methanol extract of *D. elliptica* was

exposed to the field-caught larvae of *Ae. aegypti* showed a lower larvicidal potential (8). This fact showed that the different extract types of the tubal root have different effects against the different strains of *Ae. aegypti* larvae where the field-caught larvae were more resistant to the phytochemical compound.

The results of monitoring of dengue vector susceptibility in Central Java, Indonesia showed a wide spread of resistance to cypermethrin 0.05% (1), as occurs in various Dengue endemic areas in other countries (4, 9, 10). Cypermethrin is one of the active ingredients of pyrethroid class insecticide which has caused knockdown resistance (kdr) (11). This resistance mechanism was indicated with the target site insensitivity

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in the voltage-gated sodium channel (VGSC) gene (1, 12). The action mechanism of cypermethrin is different from the temephos. This compound is an active ingredient of organophosphate insecticide class which inhibited the acetylcholinesterase enzyme (13). This different mechanism of action is interesting to be studied in understanding the larvicidal activity spectrum of *D. elliptica* root extracts.

The main biochemical compounds of *D. elliptica* are alkaloids, flavonoids, sterols, tannins, and triterpenoids (14, 15), and rotenone is the most important of a specific compound of flavonoid (16). These compounds have a toxic effect that kills insect larvae through disrupting mechanisms of the endocrine and hormonal systems (14) and reducing the esterase and monooxygenase enzymes (16). Initial studies showed that ethyl acetate, methanol, and n-hexane extracts of *D. elliptica* that have different polarity effectively killed *Ae. aegypti* larvae which were susceptible to temephos 0.02ppm (17), but on the other hand, the ethanol extract type has a lower effect against the temephos-resistant strains (18). The lethal effects of different specific phytochemicals contained in *D. elliptica* root extracts against the offspring larvae of the cypermethrin-resistant strain *Ae. aegypti* is still lack understood and is interesting to be studied. It is important to evaluate the larvicidal effect of the semi-polar extract, ethyl acetate against this strain. This study aimed to determine the larvicidal activity of ethyl acetate extract of *D. elliptica* root against the third-instar larvae of the cypermethrin-resistant *Ae. aegypti* offspring.

Materials and Methods

This experiment is the early part of an ongoing study 'Isolation of specific compounds of *Derris elliptica* as the larvicidal ingredients against *Aedes aegypti* mosquito in the dengue control'. Ethyl acetate extract is the last step of the sequential extraction process (19, 20). In the summary modification, the extraction pro-

cess started by maceration of the tuba root powder in methanol solvent for 3 x 24 hours, and then filtered. The clean part of the liquid is evaporated and produced the methanol extract (the crude extract). Furthermore, the crude extract was partitioned liquid-liquid with n-hexane solvent to bind the nonpolar lead compounds and result in water fraction and n-hexane fraction. The water fraction obtained was partitioned with ethyl acetate to bind the semi-polar lead compounds and produced the water fraction and ethyl acetate fraction. All fractions produced were evaporated by using a rotary evaporator to produce four types of extracts, including the ethyl acetate extract which was first completed.

The subjects of this study were the offspring filial 2 (F2) larvae of the cypermethrin 0.05% resistant strain of *Ae. aegypti*. The parental mosquitoes were the F1 larvae of the *Ae. aegypti* that is reared from F0 larvae obtaining from a household survey in the Dengue endemic areas in the Community Health Center of Kedung Mundu, Tembalang District, Semarang City, and subjected to bioassay test using the Cypermethrin-0.05% compound. The result of the bioassay test showed a mortality rate of 85%, which indicated that the mosquito population was resistant to the pyrethroid compound (21). Larvae were maintained in the Epidemiology and Tropical Diseases laboratory of Public Health Faculty of Universitas Muhammadiyah Semarang, Indonesia. The larvae were placed on a plastic tray containing tap water. Conditions of temperature and humidity were maintained in the range of 28 ± 2 °C and $75 \pm 10\%$. The larvae were fed dog food. Bioassay tests apply the WHO standard procedures for larvicide testing (21). There are three important parts at this stage, namely preparation of extract concentration, selection of research subjects, and exposure of research subjects with various *D. elliptica* extracts. The concentration of the bioassay test used a range of 10, 25, 50, and 100ppm, the effective concentration in another study using *Ae. aegypti* larvae of laboratory

strains (17). The subject of the research was the third instar larvae offspring of the Cypermethrin-resistant *Ae. aegypti*, in intact condition, and actively moving. As many as twenty larvae were subjected to each experiment group.

Two control groups, negative (tap water) and positive (temephos 0.02ppm) control were followed. The effectiveness of the larvicidal activity of the ethyl acetate extract of *D. elliptica* root was determined by the LC₅₀ that was obtained from probit analysis. This LC₅₀ will be compared with the LC₅₀ of previous experiment results of the same extract type against the laboratory (susceptible) strain of *Ae. aegypti* larvae (17). Analysis data was performed descriptively and analytically by using SPSS statistical software version 15.0. The research protocol obtained ethical approval from the Ethics Committee of Health Research of Public Health Faculty of Universitas Muhammadiyah Semarang with registration number 231/KEPK-FKM/UNIMUS/2019.

Results

14 The ethyl acetic extracts of *D. elliptica* root showed the larvicidal activity against the *Ae. aegypti* larvae of offspring from the resistant parental to cypermethrin 32 multicide. There was an increase in the larval mortality rate of *Ae. aegypti* larvae after 24 and 48 hours of exposure to the ethyl acetic extract from 40–67.5% to 62.5–97.5% (Table 1), with LC₅₀ of 34.945 and 6.461ppm, respectively (Table 2). The larval mortality rate increased directly with the extract concentration. There were no larvae died in the negative control, and 100% of larvae died in the positive control. The trend of the knockdown larvae showed that the slow larvicidal activity of the ethyl acetate extract of *D. elliptica* root to the third-instar larvae, offspring F2 of cypermethrin-resistant *Ae. aegypti* (Fig. 1). Statistical analysis showed the differences in larval mortality rate based on the dosage and exposure time (Fig. 2).

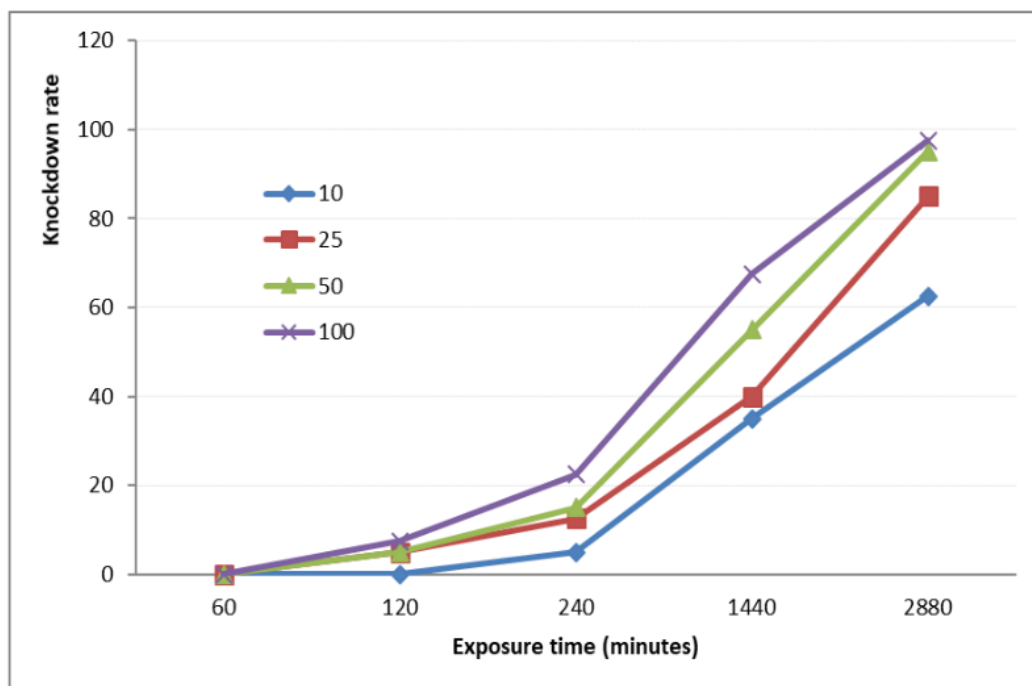


Fig. 1. The trend of larval knockdown rate in each extracts concentration based on the exposure time. The colored-line represents the concentrations of the extract

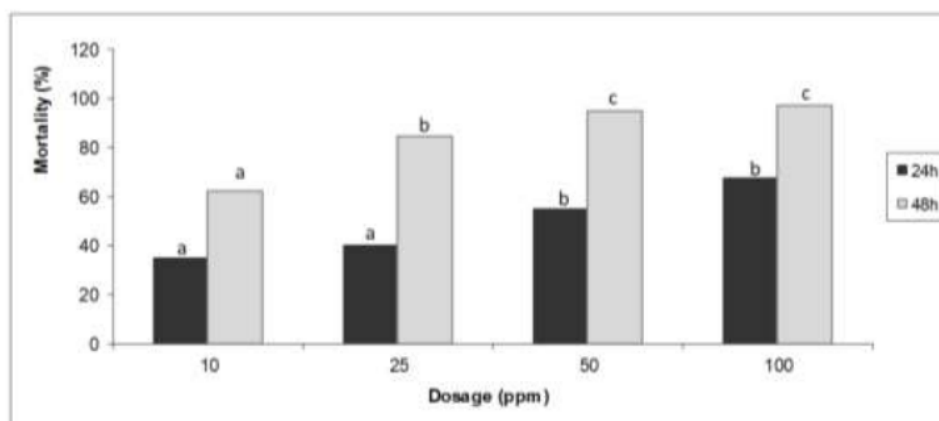
3 **Table 1.** Larvicidal activity of the ethyl acetate extract type of *Derris elliptica* against the third-instar larvae of cypermethrin-resistant *Aedes aegypti* offspring

Concentration (ppm)	24-hours mortality rate (%)			48-hours mortality rate (%)		
	Min	Max	Mean	Min	Max	Mean
0 (dw)	0	0	0	0	0	0
10	30	40	35	60	65	62.5
25	40	40	40	80	90	85
50	40	70	55	90	100	95
100	65	70	67.5	95	100	97.5
0.02 (tem)	100	100	100	100	100	100

dw= distilled water; tem= temephos

Table 2. The LC₅₀ of larvicidal activity of the *Derris elliptica* ethyl acetate extract against offspring larvae of the cypermethrin-resistant *Aedes aegypti*

Exposure time (hours)	Regression equation	LC ₅₀ (95% confidence limits)
24	Y= -1.331+0.862X	34.945 (18.179–70.780)
48	Y= -1.419+1.751X	6.461 (2.206–10.359)



2 **Fig. 2.** Mortality rate comparison of *Aedes aegypti* larvae after 24h and 48h exposure of ethyl acetate extract of *Derris elliptica* root. The differences of larval mortality rate are indicated by the letters a, b and c

Discussion

3 Results of the experiment showed that the ethyl acetic extract of *D. elliptica* has a high larvicidal activity against the third-instar larvae of the cypermethrin-resistant *Ae. aegypti* F2 offspring, although at the concentration of 100ppm for 24 hours, the larvicidal effect of the ethyl acetate extract type is still lower than the larvicidal activity of temephos 0.02ppm. Although the ethyl acetate extract type of tuba root

shows lower larvicidal activity than temephos, this extract still indicates high larvicidal potential because its LC₅₀ is lower than 50ppm. A previous study reported that the effective larvicidal activity of plant extracts was categorized into three levels, namely high (LC₅₀< 50ppm), moderate (LC₅₀< 100ppm), and low LC₅₀< 750 ppm) (4). The results of this experiment indicate that the potency of the ethyl acetate extract

of *D. elliptica* root against the third-instar larvae of cypermethrin-resistant *Ae. aegypti* F2 offspring is 84.4% lower than the same extract potency against the susceptible (laboratory) strain (17) indicating by the LC₅₀ bioassay test for 24 hours, respectively 34,945ppm and 21,063 ppm. This extract also had a higher larvicidal potential against F2 offspring larvae of Cypermethrin-resistant *Ae. aegypti* rather than the temephos-resistant offspring (18). The larvicidal activity of this extract was also better than the methanolic extract of *M. glaziovii* peel (22), *A. pinata* (23), *T. patula* (24), *H. forskalii* (25), *O. campechiana* (23) and *O. quixos* (26), and *A. occidentale* (27) against the third-instar larvae of *Ae. aegypti*, although lower than the specific isolate compound of *F. vulgare* (28) and *P. aduncum* (26) essential oils, and *P. foetida* ethyl acetate extract (29). These preliminary data and information have given new hope that this extract can be an alternative ingredient of larvicide to inhibit the growth of *Ae. aegypti* larvae, even to the strains that are already resistant to cypermethrin adulticide.

These results also indicate that temephos is still effective in killing the third-instar larvae offspring F2 of the cypermethrin-resistant *Ae. aegypti*. This condition showed that there is still a way to eradicate the Dengue vector, even from strains that have been resistant to other insecticides because each insecticide compound has a different mode of action (30, 31). Cypermethrin is a compound of the pyrethroid class, an insecticide group that disrupts the function of sodium channels in insect nerves (32). Under normal conditions, the voltage-gated sodium channel (VGSC) gene works 'open' and 'close' to regulate electrical impulses into the cell. The linkage of the pyrethroid insecticide molecule to the gene disrupts the nerve regulation and impulse of the nerve flowing continuously so that the insects become convulsion and died (33). However, if the point mutations occur in this gene, the linkage of the pyrethroid insecticide molecule does not affect the life of the insect, and this condition caused the *kdr* (34).

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The high larvicidal activity of the ethyl acetate extract of *D. elliptica* root against the third instar larvae, the offspring of the cypermethrin-resistant *Ae. aegypti* indicates that this extract has a different mechanism of action than adulticide cypermethrin. The *D. elliptica* extract contains several lead compounds such as tannins, phlobatannins, terpenoids, cardiac glycosides, and flavonoids (35) mainly rotenone and rotenoids (36). Mode of action of rotenone is inhibition the cellular respiration, while pyrethrins the active compound of the pyrethroid insecticide has a mode of action in disruption of the sodium and potassium ions exchange (37). It means that the exploration of specific isolates of *D. elliptica* extract has the opportunity to be developed into larvicidal bioactive compounds with different modes of action. On the other hand, the effectiveness of temephos larvicide in killing the offspring larvae of the cypermethrin resistant strain of *Ae. aegypti* proved that rotational insecticide with different modes of action and target sites is necessary done. Temephos is a compound that plays a role in protein carbonylation so that it causes the general oxidative damage in larval cellular of insects (38).

The maximum effect of the ethyl acetate extract of *D. elliptica* was achieved at 48 hours of exposure time. This condition indicated that the mode of action of this extract is slower than temephos. It can be understood that temephos is a pure chemical compound, while the plant extract still contains many chemical compounds, which may have antagonistic effects (39). Extraction with ethyl acetate solvent has selected chemical compounds that are semi-polar, according to the nature of the solvent. However, the extraction results still allow the dissolution of many plant chemical compounds with various modes of action although flavonoid was the dominant compound (40). Therefore, the pure compounds from these extracts that have the best larvicidal activity are necessarily understood.

Conclusion

The ethyl acetate ¹⁴ude extract of *D. elliptica* root has a high larvicidal activity against the third-instar larvae, offspring of the cypermethrin-resistant strain of *Ae. aegypti*, although the effect is lower and slower than temephos. Isolation of the pure compounds of the extract is needed to find the specific active compounds for larvicide formulation.

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References

1. Sayono S, Hidayati APN, Fahri S, Suman- to D, Dharmana E, Hadisaputro S, Asih PBS, Syafruddin D (2016) Distribution of Voltage-Gated Sodium Channel (*Nav*) alleles among the *Aedes aegypti* populations in Central Java Province and its association with resistance to pyrethroid insecticides. PLoS One. 11(3): e0150577.
2. Araújo AP, Paiva MHS, Cabral AM, Cavalcanti AEHD, Pessoa LFF, Diniz DFA, Helvecio E, Silva EVG, Silva NM, Anastácio DB, Pontes C, Nunes V, Souza MFM, Magalhães FJR, Santos MAVM, Ayres CFJ (2019) Screening *Aedes aegypti* (Diptera: Culicidae) populations from Pernambuco, Brazil for resistance to Temephos, Diflubenzuron, and Cypermethrin and characterization of potential resistance mechanisms. J Insec Sci. 19 (3): 1–15.
3. Dohutia C, Bhattacharyya DR, Sharma SK, Mohapatra PK, Bhattacharjee K, Gogoi K, Gogoi P, Mahanta J, Prakash A (2015) Larvicidal activity of few select indigenous plants of North East India against disease vector mosquitoes (Diptera: Culicidae). Trop Biomed. 32(1): 17–23.
4. Komalamisra N, Trongtokit Y, Rongsriyam R, Apiwathnasorn C (2005) Screening for larvicidal activity in some Thai plants against four mosquito vector species. Southeast Asian J Trop Med Public Health. 36(6): 1412–1422.
5. Arnason JT, Sims SR, Scott IM (2012) Natural products from plants as insecticides. Encyclopedia of Life Support System (EoLSS). Phytochemistry and pharmacognosy. Available at: <http://www.eolss.net/sample-chapters/c06/e6-151-13.pdf>
6. Zubairi SI, Sarmidi MR, Aziz RA (2015) A preliminary study on mosquito larvicidal efficacy of rotenone extracted from Malaysia *Derris* sp. J Teknologi. 76(1): 275–279.
7. Komansilan A, Suriani NW, Lawalata H (2017) Test toxic tuba root extract as a natural insecticide on larvae of *Aedes aegypti* mosquito vector of dengue fever. Int J Chemtech Res. 10(4): 522–528.
8. Sayono S, Nurullita U, Suryani M (2010) Pengaruh konsentrasi flavonoid dalam ekstrak akar tuba (*Derris elliptica*) terhadap kematian larva *Aedes aegypti*. [In Indonesian]. J Kesehat Masy Indones. 6 (1): 38–47.
9. Sayono S, Nurullita U (2016) Situasi terkini vektor dengue (*Aedes aegypti*) di Jawa Tengah, Indonesian. Kemas. Jurnal Kesehatan Masyarakat. 11(2): 96–105.
10. Moyes C, Vontas J, Martins AJ, Ng LC, Koou SY, Dusfour I, Raghavendra K, Pinto J, Corbel V, David JP, Weetman D (2017) Contemporary status of insecticides resistance in the major *Aedes* vectors of arboviruses infecting humans.

- PLoS Negl Trop Dis. 11(7): e0005625.
11. Singh AK, Tiwari MN, Prakash O, Sing MP (2012) A current review of Cypermethrin-induced neurotoxicity and nigrostriatal dopaminergic neurodegeneration. *Neuropharmacology* 10: 64–71.
 12. Kuswah RBS, Kaur T, Dykes CL, Kumar HR, Kapoor N, Sing OP (2020) A new knockdown resistance (kdr) mutation, F1534L, in the Voltage-gated Sodium Channel of *Aedes aegypti*, co-occurring with F1534C, S989P, and V1016G. *Parasit Vectors*. 13: 327.
 13. Insecticide Resistance Action Committee (IRAC) (2019) IRAC mode of action classification scheme. IRAC International Working Group. pp. 5–7.
 14. Ge Y, Liu P, Yang R, Zhang L, Chen H, Camara I, Liu Y, Shi W (2015) Insecticidal constituents and activity of alkaloids from *Cynanchum mongolicum*. *Molecules*. 20: 17483–17492.
 15. Khan MR, Omoloso AD, Barewai Y (2006) Antimicrobial activity of the *Derris elliptica*, *Derris indica* and *Derris trifoliata* extractives. *Fitoterapia*. 77(4): 327–330.
 16. Visetson S, Milne M (2001) Effect of root extract from *Derris* (*Derris elliptica* Benth) on mortality and detoxification enzyme levels in the *Demodback* Moth larvae (*Plutella xylostella* Linn.). *Kaset-sart J (Nat. Sci.)*. 35: 157–163.
 17. Sayono S, Anwar R, Sumanto D (2020) Evaluation of toxicity in four extract types of Tuba root against Dengue vector, *Aedes aegypti* (Diptera: Culicidae) larvae. *Pak J Biol Sci*. 23(12): 1530–1538.
 18. Sayono S, Permatasari A, Sumanto D (2019) The effectiveness of *Derris elliptica* (Wall.) Benth root extract against Temephos-resistant *Aedes aegypti* larvae. IOP Conference Series: Earth and Environmental Science. 292. The 1st International Conference on Food Science and Technology 28–29 November 2018, Universitas Muhammadiyah Semarang, Semarang, Indonesia, p. 102052.
 19. Liu Z (2008) Preparation of botanical samples for biomedical research. *Endocr Metab Immune Disord Drug Targets*. 8 (2): 112–121.
 20. Li XJ, Hareyama T, Tezuka Y, Zhang Y, Miyahara T, Kadota S (2005) Five new oleanolic acid glycosides from *Achyranthes bidentata* with inhibitory activity on osteoclast formation. *Planta Med*. 71(7): 673–679.
 21. World Health Organization (2016) Monitoring and managing insecticide resistance in *Aedes* mosquito populations. Interim guidance for entomologists. The WHO Department of Control of Neglected Tropical Diseases and Global Malaria Programme, Geneva. Available at: https://apps.who.int/iris/bitstream/handle/10665/204588/WHO_ZIKV_VC_16.1_eng.pdf;jsessionid=A18C3776D514404CE7F5CC90F1981029?sequence=2
 22. Sayono S, Safira FA, Anwar R (2019) In vitro study on the larvicidal activity of *Manihot glaziovii* peel extract against *Aedes aegypti* larvae. *Ann Parasitol*. 65 (4): 403–410.
 23. Ravi R, Zulkarnin NSH, Rozhan NN, Niksoff NR, Mat Rasat MS, Ahmad MI, Ishak IH, Mohd Amin MF (2018) Chemical composition and larvicidal activities of *Azolla pinnata* extracts against *Aedes* (Diptera: Culicidae). *PLoS One*. 13(11): e0206982.
 24. Krzyzaniak LM, Antonelli-Ushirobira TM, Panizzon G, Luiza Sereia AL, Souza JRP, Zequi JAC, Novello CR, Lopes GC, Medeiros DC, Silva DB, Leite-Mello EVS, Jello JCP (2017) Larvicidal activity against *Aedes aegypti* and chemical characterization of the inflorescences of *Tagetes patula*. *Evid Based Comple*

- ment Alternat Med. 2017: 9602368.
25. Sillo AJ, Makirita WE, Swai H, Chacha (2019) Larvicidal activity of *Hypoestes forskaolii* (Vahl) R.Br root extracts against *Anopheles gambiae* Giles.s., *Aedes aegypti* L., and *Culex quinquefasciatus* Say. J Exp Pharmacol. 11: 23–27.
 26. S (2019) Larvicidal activity of *Ocimum campechianum*, *Ocotea quixos* and *Piper aduncum* essential oils against *Aedes aegypti*. Parasit. 26(23): 1–8.
 27. Amado CJRR, Souto CRNP, Magalhães MS, Arranz CJCE, Carvalho CJCT (2017) Chemical composition and larvicidal activity of cashew nutshell ethanolic extract against mosquito larvae. Rev Cub Quim. 29(3): 330–340.
 28. Rocha DK, Matos O, Novo MT, Figueire- (2015) Larvicidal activity against *Aedes aegypti* of *Foeniculum vulgare* essential oils from Portugal and Cape Verde. Nat Prod Commun. 10 (4): 677–682.
 29. M (2016) Aktivitas larvasida ekstrak etanol, fraksi n-heksan, etil asetat, dan metanol daun sembukan terhadap larva nyamuk *Aedes aegypti* dan *Anopheles* instar III. Trad Med J. 21(3): 137–142.
 30. Das SK (2013) Mode of action of pesticides and the novel trends-A critical review. Int Res J Agric Sci Soil. 3(11): 393–401.
 31. Spark TC, Nauen R (2015) IRAC: Mode of action classification and insecticide resistance management. Pestic Biochem Physiol. 121: 122–128.
 32. Soderlund DM (2012) Molecular mechanisms of pyrethroid insecticide neurotoxicity: recent advances. Arch Toxicol. 86(2): 165–181.
 33. Silver KS, Du Y, Nomura Y, Olivera EE, Salgado VL, Zhoro BS, Dong K (2014) Voltage-Gated Sodium Channels as insecticide targets. Adv In Insect Phys. 46: 389–433.
 34. Dong K, Du Y, Rinkevich F, Nomura Y, Xu P, Wang L, Silver K, Zhoro BS (2014) Molecular biology of insect sodium channels and pyrethroid resistance. Insect Biochem Mol Biol. 50: 1–17.
 35. Lagu (2015) Evaluation of the phytochemical constituents of the leaves of *Ficus minahassae* Tesym and De Vr., *Casuarina equisetifolia* Linn., *Leucosyke capitellata* (Pior) Wedd., *Cassia sophera* Linn., *Derris elliptica* Benth., *Cyperus brevifolius* (Rottb.) Hassk., *Piper abbreviatum* Opiz., *Ixora chinensis* Lam., *Leea aculeata* Blume, and *Drymoglossum piloselloides* Linn. AAB Bioflux. 7 (1): 51–58.
 36. Zubairi SI, Ramli KA, Majid FAA, Sarmidi MR, Aziz RA (2005) Biological screening on the extract of *Derris elliptica*. Proceeding of Kustem 4th Annual Seminar on Sustainability Science and Management, 2005 May 2–3, Primula Beach Resort, Kuala Terengganu. Available at: <http://eprints.utm.my/id/eprint/5263/>
 37. Ghosh A, Chowdhury N, Chandra G (2012) Plant extracts as potential mosquito larvicides. Indian J Med Res. 135: 581–598.
 38. Rodríguez-Cavallo E, Guarnizo-Méndez J, Yépez-Terrill A, Cárdenas-Rivero A, Díaz-Castillo F, Méndez-Cuadro D (2018) Protein carbonylation is a mediator in larvicidal mechanisms of *Tabernaemontana cymosa* ethanolic extract. J King Saud Univ Sci. 31(4): 464–471.
 39. Millugo TK, Osoma LK, Ochanda JO, (2013) Antagonistic effect of alkaloids and saponins on bioactivity in the quinine tree (*Rauvolfia caffra-sond.*): further evidence to support biotechnology in traditional medicinal plants. BMC Compl Alternative Med. 13: 285.

- 8
40. Thavamoney N, Sivanadian L, Tee LH, Khoo HE, Prasad KN, Kong KW (2018) Extraction and recovery of phytochemical components and antioxidative properties in fruit parts of *Dacryodes rostrata* influenced by different solvents. J Food Sci Technol. 55(7): 2523–2532.

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Neil E. Coughlan. "Aquatic plant extracts and coverage mediate larval mosquito survivorship and development", *Biological Control*, 2020

Publication

13

Tássio Rômulo Silva Araújo Luz, Ludmilla Santos Silva de Mesquita, Flavia Maria Mendonça do Amaral, Denise Fernandes Coutinho et al. "Essential oils and their chemical constituents against *Aedes aegypti* L. (Diptera: Culicidae) larvae", *Acta Tropica*, 2020

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"Green Synthesized Silver Nanoparticles: A Potential New Insecticide for Mosquito Control", *Parasitology Research Monographs*, 2016.

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Kourosh Arzamani, Yavar Rassi, Hassan Vatandoost, Amir Ahmad Akhavan et al. "Comparative Performance of Different Traps for Collection of Phlebotominae Sand Flies and Estimation of Biodiversity Indices in Three Endemic Leishmaniasis Foci in North Khorasan Province, Northeast of Iran", Journal of Arthropod-Borne Diseases, 2020

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K. Dass, P. Mariappan. "Larvicidal Activity of Colocasia esculenta, Eclipta prostrata and Wrightia tinctoria Leaf Extract Against Culex quinquefasciatus", Proceedings of the National Academy of Sciences, India Section B: Biological Sciences, 2014

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Channel (vgsc) Gene of the Main Dengue Vector, *Aedes aegypti* (Diptera: Culicidae) and the Discovery of Novel Regional Specific Point Mutation A1007G in Malaysia.", Research Square, 2020

Publication

25

Mayura Soonwera. "Efficacy of essential oil from *Cananga odorata* (Lamk.) Hook.f. & Thomson (Annonaceae) against three mosquito species *Aedes aegypti* (L.), *Anopheles dirus* (Peyton and Harrison), and *Culex quinquefasciatus* (Say)", Parasitology Research, 2015

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