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Antibacterial activity of seed kernel extracts of seven mangoes (Mangifera indica) cultivars native to Indonesia against MDR-Pseudomonas aeruginosa isolated from wounds

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Abstract. Prastiyanto ME, Darmawati S, Mukaromah AH. 2022. In vitro evaluation of the antibacterial activity of seed kernel extracts of seven mangoes (Mangifera indica) cultivars native to Indonesia against MDR-Pseudomonas aeruginosa isolated from wounds. Biodiversitas 23: 5629-5637. Pseudomonas aeroginusa 41 the most common bacterium causing wound infections, with the most common solution being antibiotics. However, excessive and inappropriate use of antibiotics will lead to the emergence of multi-drug resistant (MDR) bacterial strains. Therefore, natural ingredients are needed as alternative antibacterial agents. This study aimed to determine the antibacterial activity of seed kernel extracts from seven cultivars of mango (Mangifera indica) from Indonesia, i.e., Cengkir, Kopyor, Golek, Kweni, Avocado, Arumanis, and Manalagi, against MDR-P35 roginusa bacteria isolated from wounds. The agar well diffusion method was carried out to determine the inhibition zone, and the microdilution method was used to determine the MIC and MBC values. The results showed that the seed kernel extracts of seven cultivars of mangoes had antibacterial activity against 59 R-P. aeruginosa. Of the seven mango cultivars, Kweni cultivar seed kernel extracts demonstrated the lowest MIC and MBC values of ≥0.75 mg/mL and ≥12.5 mg/mL. This study concludes that Kweni cultivar seed kernel extracts have the potential to be developed as agents of anti-MDR-P. aeruginosa causes wound infection.

Keywords: Mangifera indica, MDR, Pseudomonas aeruginosa, seeds, wounds

INTRODUCTION

A wound is a condition in which the extracellular matrix (ECM) is damaged, resulting in the loss of the skin's protective function, with or without damage to underlying tissues (muscle, bone, and blood vessels). Wounds are caused by trauma to the skin from sharp objects or collisions. Wounds are also instigated by lacerations of membranes, most commonly the skin. Microorganisms can invade and multiply through an open wound (Nasser et al. 2018). In chronic wounds, bacteria are associated with multiple species, causing virulent tissue damage. Pseudomonas, Staphylococcus, Enterobacter, Peptoniphilus, Stenotrophomonas, Serratia, and Finegoldia are commonly found in wounds (Rahim et al. 2016). A previous study by Puca et al. (2021) reported that the wound samples examined contained Gram-negative bacteria (57%) and Gram-positive bacteria (36.6%). The Gram-positive bacteria include Staphylococcus aureus (79.4%), while the Gram-negative bacteria comprise Escherichia coli (20.7%), Pseudomonas aeruginosa (40.2%), Acinetobacter baumanii/haemolyticus (9.5%), and Proteus mirabilis (11.2%). P. aeroginusa is the most commonly found among the Gram-negative bacteria that

infect wounds. Pseudomonas aeruginosa is an opportunistic pathogenic bacteria widely distributed in nature (Moradali et al. 2017). These bacteria can form biofilms on wounds, a major public health concern. Biofilms are communities or colonies of bacteria that grow together on the extracellular matrix (ECM), a fundamental structural component of the bacterial community that serves as a protective barrier (Ma et al. 2009). Pseudomonas aeruginosa colonization can result in acute skin or wound infections (Bassetti et al. 2018). The bacteria are known to infect wounds (Smolle et al. 2018). The mortality rate of patients with wounds infected with P. aeruginosa, particularly MDR bacteria, is higher than that of patients with uninfected wounds (Branski et al. 2009).

Antibiotics are commonly used to treat wounds. Common antibiotics used in wound care include macrolides, β lactams, tetracycline, fluoroquinolones, and aminoglycosides (Tzaneva et al. 2016). Antibiotics play a role in suppressing bacterial growth, but overuse of antibiotics causes systemic damage (Everts 2017). In addition, excessive and uncontrolled antibiotic use resulted MDR bacterial strains. Therefore, alternative antibacterial sources from natural ingredients are needed. antibacterial sources comprise

microorganisms (Prastiyanto et al. 2022), lactic acid bacteria (Lestari et al. 2019), mushrooms (Prastiyanto et al. 2020a), fruit (Prasnto et al. 2020c), seeds (Prastiyanto 2021), and latex (Prastiyanto et al. 2020b). Plants are the most common and potential antibacterial agents from natural sources of antibacterial agents (Prastiyanto et al. 2021), one of which is mango.

Mango (Mangifera indica L.) belongs to the Anacardiaceae family of the order Sapindales, an economically important tropical fruit plant, a source of traditional medicine (Kumar et al. 2021; Ghosh et al. 2022). In 2018, the global mango production was 55.4 million tonnes, with India, China, Thailand, and Indonesia leading the production (Kumar et al. 2021). There are over 1000 mango cultivars available worldwide, but only a few are traded.

Mango cultivars from various countries have been reported to have antibacterial properties. For example, a previous study showed that the Magaysian Chokanan mango seed methanol extract had antibacterial activity against Methicillin-Resistant Staphylococcus aureus (MRSA) and Escherichia coli (Kaur et al. 2010). Another study showed that ethanol and methanol extracts of Sudanese mango seed cultivars had good inhibitory activity against almost all tested strains, i.e., Bacillus cereus, Citrobacter freundii, Escherichia coli, Listeria monocytogens, Mycobacterium senegalense, Salmonella typhi, Shigella flexnerri, Staphylococcus aureus, Yersinia enterocolitica (El-Gied et al. 2012). However, there has been limited research on the antibacterial properties of Indonesian mango cultivars. This present study attempted to fill the research gap by evaluating the antibacterial activity of seven mango cultivars (Cengkir, Kopyor, Golek, Kweni, Avocado, Arumanis, and Manalagi) originated from Indonesia against MDR P. aeruginosa isolated from wounds.

MATERIAL AND METHODS

Collection and extraction of mango seeds

Seeds of mango cultivars of Čengkir, Kopyor, Golek, Kweni, Avocado, Arumanis, and Manalagi, were collected from local markets (Semara 11 Central Java, Indonesia) (Figure 1). First, the mango seeds were washed and sundried for seven days. Then, the dried seeds were soaked in a 9655 ethanol solution at a 1:3 ratio at room temperature for 24 hours. The ethanol solvent was changed every 24 hours until the solution became clear, indicating that the bioactive components had been extracted in ethanol. The supernatant was filtered using Whatman paper No. 1, and the filtrate was concentrated with a rotary evaporator at 40°C.

Isolation of Pseudomonas aeroginusa from wounds

Pseudomonas aeroginusa isolates were obtained from wound samples from patients at Tugurejo Hospital in Semarang, Indonesia. T 57 bacteria were cultured using MacKonkey agar media and incubated at 35 ± 2°C for 24 hours. The colonies were non-lactose fermenters selected using positive catalase and oxidase testing. Vitek®MS was used for bacterial identification and susceptibility testing (Prastiyanto 2021). The fol 45 ing antibiotics used as positive controls were AMP (Ampicillin), TZP (Piperacillin-tazobactam), SAM (Ampicillin-sulbactam), CAZ (Ceftazidime), CFZ (Cefazolin), FEP (Cefepime), CRO (Ceftriaxone), ATM (Aztreonam), (Tigecycline), MCA (Amikacin), CIP (Ciproflox), MEM (Meropenem), and GEN (Gentamicin). MDR-P. aeruginosa determination was based on the Clinical Laboratory Standard Institute (CLSI) of M100-S25 (CLSI

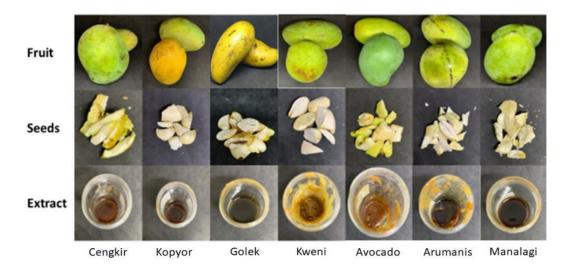


Figure 1. Fruit, seeds, and extracts of seven mango cultivars

The anti-MDR activity against *Pseudomonas aeroginusa* from mango seed extracts

Agar well diffusion

Agar well diffusion assay was used to evaluate the antibacterial activity of mango seed extracts against MDR-P. aeruginosa (Prastivanto 2021). MDR-P aeroginusa isolates from patient wounds were cultured on a blood agar plate and then incubated for 24 hours at $35 \pm 2^{\circ}$ C. All bacterial isolates were standardized with 0.5 McFarland (1.5x108 CFU/mL). Each isolate was inoculated on Muller Hilton agar (MHA) media using a sterile cotton swab, 10 minutes incubation. The MHA media was perforated using a cork borer (0.5 cm in 7 iameter). Four holes were made for each concentration (0.1 mg/mL, 1 mg/mL, 10 mg/mL, and 100 mg/mL). The extract was dissolved in Dimethyl sulfoxide (DMSO). Then, 100 µL of the extract was added to each well, followed by incubation at a temperature of 35 28 2°C for 16-20 hours. The antibiotics used were Ampicillin (AMP) (10 μ g), ceftriaxone (CTR) (30 μ g), Tigecycline (TGC) (15 μ g), Nitrofurantoin (NIT) (300 μ g), Trimethoprim (TMP) (25 μ g), Gentamicin (GEN) (10 μ g), Meropenem (MEM) (10 µg), and Ciprofloxacin (CIP) (5

Minimum Inhibitory Concentration (MIC)

The microdilution method used Mueller Hilton Broth (MHB) media added with 0.05% 2,3,5-Triphenyltetrazolium chloride on a microwell plate to determine the MIC (Prastiyanto 2021). First, MHB (100 $\mu\text{L})$ was added to each well, and 100 μL of the extract was supplemented to the first well, then serial dilutions were made up to the 12^{th} well. After that, 10 μL of MDR P. aeruginosa was added to each well, then incubated for 18-20 hours at 35 \pm 2°C. The MIC value was observed by the presence of the color change on the microwell plate. The lowest MIC value of the extract was indicated as the best antibacterial activity.

Minimum Bactericidal Concentration (MBC)

Wells on MIC $_{53}$ e subcultured on Blood Agar Plate (BAP) media and then incubated at $35 \pm 2^{\circ}$ C for 16-20 hours. The MBC value was determined by observing the growth of bacteria on BAP media. The MBC value was determined as the lowest concentration at which MDR-P. aeruginosa is unable to grow (Prastiyanto 2021).

Phytochemical screening of the extract

Phytochemical analysis (triterpenoids, steroids, flavonoids, alkaloids, phenolates, tannins, and saponins) of the crude extracts of mango seeds was carried out using previously described methods (Eve et al. 2020).

RESULTS AND DISCUSSION

MDR-Pseudomonas aeruginosa from wound

MDR-P. aeruginosa bacteria were isolated from the wounds and tested for their susceptibility to several antibiotics. The susceptibility test showed that the isolated bacteria were MDR P. aeruginosa (Figure 2) because it

was resistant to at least three or more classes of antibiotics. MDR-P. aeruginosa strain #1 showed resistance to penicillins (Ampicillin, Piperacillin-tazobactam, and Ampicillin-sulbactam), cephalosporins (Ceftazidime, Ceftriaxone, and Cefazolin), Monobactam (Aztreonam), carbapenems (Meropenem), aminoglycosides (Amikacin), fluoroquinosides (Amikacin), (ciprofloxacin) glycylcycline (Tigecycline). The MDR-P. aeruginosa strain #2 was resistant to the following classes of antibiotics: penicillins (Piperacillin-tazobactam and Ampicillin-sulbactam), cephalosporins (Ceftriaxone and Ceftazidime), Monobactam (Aztreonam), carbapenems (Meropenem), aminoglycosides (Amiconemikacin, fluoroquintacocin). MDR-P. aeruginosa strain #3 was resistant to the following antibiotics: penicillins (Ampicillin-sulbactam, Ampicillin, and Piperacillintazobactam), cephalosporins (Ceftriaxone, Ceftazidime, and Cefazolin), Monobactam (Aztreonam), carbapenems (Meropenem). aminoglycosides (Amikacinosine), (ciprofloxacin) and glycylcycline fluoroquinolones (Tigecycline). MDR-P. aeruginosa strain #4 was resistant cephalosporins (Ceftriaxone, Ceftazidime, and Cefazolin), Monobactam (Aztreonam), carbapenems (Meropenem), aminoglycosides (Gentamicin, Amikacin), fluoroquinolones (ciprofloxacin) and glycylcycline (Tigecycline). Pseudomonas aeruginosa is a bacterium commonly found in wounds (Nagoba et al. 2013).

The results of this study revealed that four bacterial isolates from the wound samples containing *P. aeruginosa* were resistant to penicillin, cephalosporin, monobactam, and carbapenem; therefore, *P. aeruginosa* produced extended-spectrum beta-lactamase (ESBL). In addition, the four isolated strains were carbapenem-resistant *P. aeruginosa* (CRPA) strains. ESBL-*P. aeruginosa* is the main cause of wound infection (Ullah et al. 2009).

Extract yields

The yield of ethanol extract from seven cultivars of mangoes (Cengkir, Kopyor, Golek, Kweni, Avocado, Arumanis, and Manalagi) was presented in Table 1. The extract yield of the Kweni cultivar was higher than the other six cultivars, indicating that the Kweni cultivar contained phytochemicals (with relatively polar constituents) that are more soluble in ethanol. In addition, ethanol solvent produced extracts with better antibacterial potential than other solvents (Prastiyanto et al. 2020c).

Table 1. The extract yields of seed kernel of seven cultivars of mango (Mangifera indica)

Cultivar	Yield (%)
Cengkir	11.86
Kopyor	25.35
Golek	28.31
Kweni	33.87
Alpukat	31.33
Arumanis	23.51
Manalagi	22.22

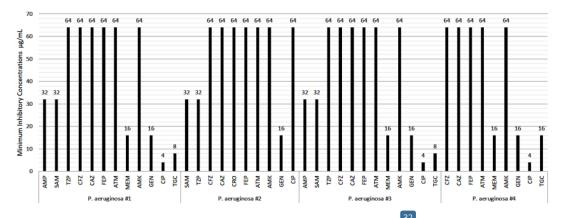


Figure 2. Susceptibility test of several antibiotics against *P. aeruginosa* isolated from wounds. AMP: Ampicillin; SAM: Ampicillin-sulbactam; TZP: 165 racillin-tazobactam; CFZ: Cefazolin; CAZ: Ceftazidime; CRO: Ceftriaxone; FEP: Cefepime; ATM: Aztreonam; MCA: Amikacin; MEM: Meropenem; CIP: ciprofloxacin, GEN: Gentamicin; TGC: Tigecycline

47e antibacterial activities assay

Agar well diffusion assay 31

The antibacterial ad 47 ty of the extract of mango seed kernels was evaluated using the agar well diffusion assay method against MDR-P. aeruginosa from wounds. The results showed that the extracts of seven mango cultivars had inhibitory zones (Figure 3), indicating the presence of antibacterial activity against MDR-P. aeruginosa. The inh 7 tory zones of mango seed extracts at concentrations of 0.1 mg/mL, 1 mg/mL, 10 mg/mL, and 100 mg/mL against four MDR-P. aeruginosa isolates from wound samples and P. aeruginosa ATCC 9027 were presented in Figure 4. This figure showed that seven mango seed extracts had antibacterial activity against MDR-P. aeruginosa wound isolates and the antibacterial activity from the extract of seven mango cultivars against MDR-P. aeruginosa was directly proportional to the ex67ct concentrations. The inhibition zone of extracts at a concentration of 0.1 mg/mL ranging from 6.0 ± 0.0 mm to 11.90 ± 0.16 mm. The inhibition zone of extracts at a concentration of 1 mg/mL ranging from 11.00 ± 0.0 mm to 17.30 ± 0.19 mm, while a concentration of 10 mg/mLhad the diameter of the inhibition ranging from 18.00 ± 0.11 mm to 28.20 ± 0.16 mm. Extract concentration of 100 mg/mL had the diameter of the inhibition zone ranging from 19.75 ± 0.43 mm to 30.63 ± 0.39 mm. Pseudomonas aeruginosa isolates had no inhibitory zones to the following antibiotics: ceftriaxone (CTR) (30 µg), Ampicillin (AMP) (10 µg), Tigecycline (TGC) (15 µg), Nitrofurantoin (NIT) (300 µg), Trimethoprim (TMP) (25 µg), Gentamicin (GEN) (10 µg), Meropenem (MEM) (10 µg), and ciprofloxacin (CIP) (5 µg). Extracts of seven mango cultivars had wider inhibitory zones than the control antibiotics.

The results of the antibacterial activity by the agar well diffusion assay showed that seed extracts of the seven mango cultivars (Cengkir, Kopyor, Golek, Kweni, Avocado, Arumanis, and Manalagi) had the potential as antibacterial agents against 4 MDR-*P aeruginosa* bacteria isolated from wounds. The inhibitory zone of these extracts

ranges from 6.00 ± 0.00 mm to 30.63 ± 0.39 mm, which is larger than previous studies. The inhibition zone diameter of mango seed kernel extract of Apple and Sabine cultivars from Kenya against *Escherichia coli* at a concentration of 100% ranges from 19.3 and 17.3 mm (Mutua et al. 2017). The study on Julie mango and John mango cultivars from Makurdi, Nigeria, against *P. aeruginosa* showed that the hexane extract of Julie mango cultivar seeds at a concentration of 100 mg/mL showed a diameter of John mango cultivars at a concentration of 100 mg/mL showed a diameter of 7.76 \pm 01.00 mm against *P. aeruginosa* (Mutua et al. 2017).

Another study reported that mango seed methanol extract from Sudan at a concentration of 5 mg/mL had an inhibition zone of 6 mm against *P. aeruginosa* 62 TCC 27853 (El-gied et al. 2012). In addition, the MDR strain of P. aeruginosa was used in this study, while the previous study used the standard strain. The ethanol extract was more likely to be chosen as a solvent in the further study because methanol is risky due to its high toxicity.

Determination of Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC)

The microdilution method using a microwell plate was used to determine the MIC and MBC values (Prastiyanto 2021). The pathogenic bacteria can be in direct contact with the extract in the dilution assay, so it does not depend on the diffusion properties of the material and media be 61 tested (Estrela et al. 2001). In the serial micro dilutions, the MIC value was determined as the extract's lowest concentration, which had antibacterial properties. In this study, the MIC and MBC values of seed kernel extract from seven mango cultivars ranged from ≥0.75 to ≥25 mg/mL dan ≥12.5 to ≥50 mg/mL, respectively (Table 2). It is the first study that evaluates the antibacterial activity of seed kernel extracts of mango cultivars native to Indonesia against MDR-*P. aeruginosa* isolated from wound, as well as qualitative screening for phytochemical content.

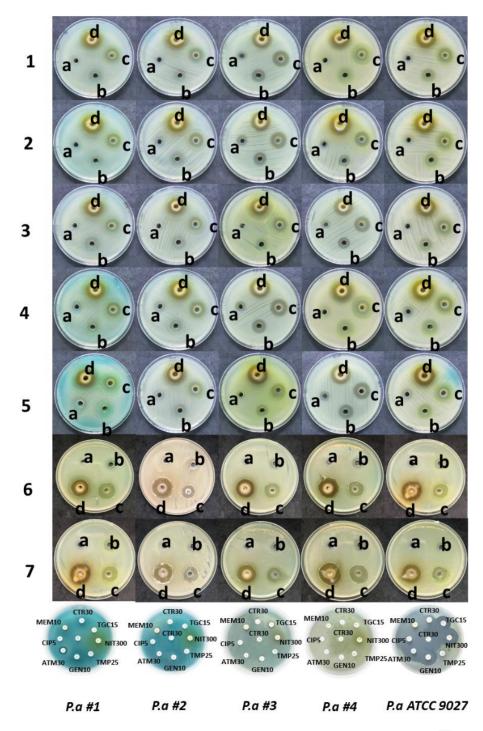


Figure 3. The inhibition zones of the extracts of seed kernels of seven mangoes (Mangifera indica L) cultivars; 1: (33-kir, 2: Kopyor, 3: Golek, 4: Kweni, 5 Avocado, 8: A 23 anis, 7: Manalagi (mg/mL) against MDR Pseudomonas aeruginosa (P.a); a: 0.1 mg/mL, b: 1 mg/mL, c: 10 mg/mL, d: 100 mg/mL; Ampicillin (AMP) (10 μg), c 45 axone (CTR) (30 μg), Tigecyclin 45 GC) (15 μg), Nitrofurantoin (NIT) (300 μg), Trimethoprim (TMP) (25 μg), Gentamicin (GEN) (10 μg), Meropenem (MEM) (10 μg), and Ciprofloxacin (CIP) (5 μg)

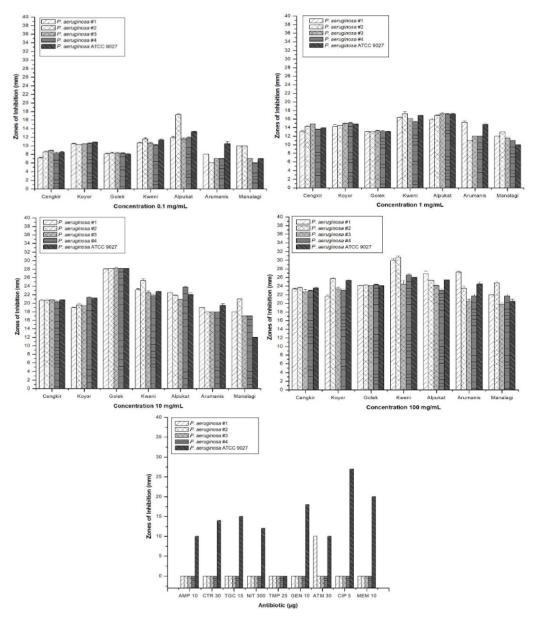


Figure 4. The diameters of the inhibition zones of seed kernels extracts of seven mangoes (Mangifera indica) cultivars from Indonesian

Table 2. The MIC and MBC values of seed kernel extracts of seven mangoes (Mangifera indica) cultivars against MDR-P. aeruginosa

MDR-P.			S	eed kern	el extrac	ts of sev	en mang	goes (Ma	ngifera i	ndica. L) cultiva	rs		
aeruginosa	40Cen	gkir	Kop	oyor	Go	lek	Kv	veni	Avo	cado	Arui	manis	Man	alagi
aeruginosa	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC
1	≥1.56	≥50	≥12.5	≥25	≥1.56	≥25	≥0.75	≥12.5	≥1.56	≥25	≥25	≥50	≥1.56	≥50
2	≥1.56	≥50	≥12.5	≥25	≥1.56	≥25	≥0.75	≥12.5	≥1.56	≥25	≥25	≥50	≥1.56	≥50
3	≥1.56	≥50	≥12.5	≥25	≥1.56	≥25	≥0.75	≥12.5	≥1.56	≥25	≥25	≥50	≥1.56	≥50
4	≥1.56	≥50	≥12.5	≥25	≥1.56	≥25	≥0.75	≥12.5	≥1.56	≥25	≥25	≥50	≥1.56	≥50
ATCC	≥1.56	≥50	≥12.5	≥25	≥1.56	≥25	≥0.75	≥12.5	≥1.56	≥25	≥25	≥50	≥1.56	≥50

The seed kernel extracts of the Kweni mango cultivar demonstrated better antibacterial activity than the other six mango cultivars, as indicated by lower MIC value against MDR-*P. aeruginosa* strains (Table 2). A previous study by Dzotam and Kuete (2017) reported that ethanol mango leaves extract had Antibacterial activities against *P. aeruginosa* PA01 and *P. aeruginosa* PA124 with a MIC value of 1.024 mg/mL. In this study, it was shown that the extracts of seed kernels of the Kweni mango cultivar had better antibacterial activity against MDR-*P. aeruginosa* with lower MIC and MBC values (≥0.75 mg/mL and ≥12.5 mg/mL).

Another study reported the Antibacterial potential of seed extracts of the Fahlun mango cultivar from Thailand against MRSA had an inhibitory zone of 10.61 ± 1.25 mm to 16.85 ± 1.94 mm at a concentration of 0.625 mg/disc to 5.00 mg/disc, with a MIC value of 0.47 \pm 0.00 mg/disc and MBC value of 1.83 ± 0.79 mg/mL (Jiamboonsri et al. 2011). These results revealed that the antibacterial activity of mango seed extract was more effective against Grampositive (MRSA) than Gram-negative (MDR-P. aeruginosa). These results align with studies by Mutua et al. (2017) that mango kernel extract has better growth inhibition against S. aureus than Escherichia coli. It relates to tannins an 48 lavonoid content in the extract. The differences in the ability to inhibit the growth of Gramnegative and Gram-positive bacteria are due to the differences in their cell walls. According to Huang et al. (2018), the low susceptibility of Gram-negative toward antibacterial agents is due to the presence of lipopolysaccharide in the membrane, which makes it more resistant.

The qualitative phytochemical screening of the seed extract of seven mango cultivars

The phytochemical screening of the ethanol extract of seven mango cultivars includes the contents of triterpenoids, steroids, flavonoids, alkaloids, phenolics, tannins, and saponins (Table 3). The results revealed that all seven mango cultivars' seed extracts contained flavonoids, alkaloids, phenolics, tannins, and saponins, but no steroids were detected in the extracts. Furthermore, triterpenoid compounds were only found in Kweni cultivars. A study by Somkuwar and Kamble (2013) showed the ethanol extract of mango seed kernels contained alkaloids, saponins, tannins, and flavonoids. Meanwhile, another study by El-gied et al. 2012) showed that the ethanol extracts of mango seeds contained saponins, triterpenes, and steroids but did not contain tannins or alkaloids. The geographic location of growth strongly influences differences in the phytochemical content of mango seed extracts. Yadav et al. (2022) highlighted that the geographical location of growth affects its phytochemical content.

This study did not evaluate the antibacterial activity of any phytochemical content of the extract against the test bacteria. However, based on previous studies, several phytochemicals have Antibacterial activity. For example, flavonoid and phenolic compounds have potent antibacterial properties. The flavonoid compounds inhibit the formation of bacterial cell walls and cause cell lysis (Royani et al. 2022). Mango seeds have been reported to have a large number of phenolic families. Polyphenols found in mango seeds include cyanidin, mangiferin gallate, homomangiferin, isomangiferin, isomangiferin gallate, and rhamnetin 3-0 galactoside/glucoside (Tirado-kulieva et al. 2021).

This study revealed that mango seed extract contains tannins; the most abundant tannin derivatives in mango are gallotanins (Kim et al. 2021). Galotannins extracted from mango seed kernels have antibacterial activity against S. aureus and E. coli (Engels et al. 2009). The compounds trap proteins on the bacterial surface, causing cell dysfunction (Luís et al. 2014). Alkaloids are one of the largest and most diverse phytochemical groups with antibacterial properties. Plants with high alkaloid content exhibit effective antibacterial properties. Alkaloids extracted from Callistemon citrinus leaves showed antibacterial activity against P. aeruginosa (ATCC 27853). The mode of action of alkaloids as antibacterial was by inhibiting ATP-dependent transport of compounds across bacterial cell membranes (Mabhiza et al. 2016). Saponins were also found in all mango cultivars. The mechanism of saponins as antibacterials is to lower the surface tension of bacterial cells, which results in increased permeability and cell leakage, which cause the release of intracellular compounds (Khan et al. 2018).

Terpenoids belong to the class of lipid compounds. Mangoes have been reported to contain several terpenoids, including terpinolene, ocimene, careen, myrcene, or limonene (Hernández-Sánchez et al. 2001). Of the seven cultivars of mangoes, Kweni showed positive results of terpenoid compounds. According to Lalel et al. (2003), terpenoid compounds are responsible for the aroma of mangoes. Therefore, it was identified that the Kweni cultivar had a strong aroma among the tested mango cultivars. The study did not identify the inhibition mechanism of terpenoid derivatives against pathogenic bacteria. However, previous studies reported that terpenoids could inhibit two important processes for bacterial survival and hinder oxygen uptake and oxidative phosphorylation of bacteria (Griffin et al. 1999). Because P. aeruginosa is an obligately aerobic bacterium, oxygen absorption is critical for producing energy for growth. Furthermore, oxidative phosphorylation is the process that causes cellular respiration in the cytoplasmic membrane. Thus, the interaction of terpenoids alters cellular respiration, resulting in the release of oxidative phosphorylation in bacteria (Zengin and Baysal 2014).

Table 3. Phytochemical content of seven mango cultivars from Indonesia

Seed Extract	Phytochemical test						
Seed Extract	Triterpenoids	Steroids	Flavonoids	Alkaloids	Phenolics	Tannins	Saponins
Cengkir	-	-	+	+	+	+	+
Kopyor	-	-	+	+	+	+	+
Golek	-	-	+	+	+	+	+
Kweni	+	-	+	+	+	+	+
Avocado	-	-	+	+	+	+	+
Arumanis	-	-	+	+	+	+	+
Manalagi	-	-	+	+	+	+	+

In conclusion, the results of the susceptibility test of P. aeruginosa bacteria from wound samples to antibiotics have ascertained that all of the bacteria are MDR-P. aeruginosa strains. The antibacterial activity in-vitro results against MDR-P. aeruginosa isolated from wounds showed evidence that all seven mango cultivars' seed kernel extracts have Antibacterial activity against MDR-P. aeruginosa. The seed extracts of the Kweni mango cultivar demonstrated greater antibacterial activity than the other six cultivars tested. Thus, seed kernel extracts of Kweni mango cultivars might be the most effective sources of antibacterials.

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