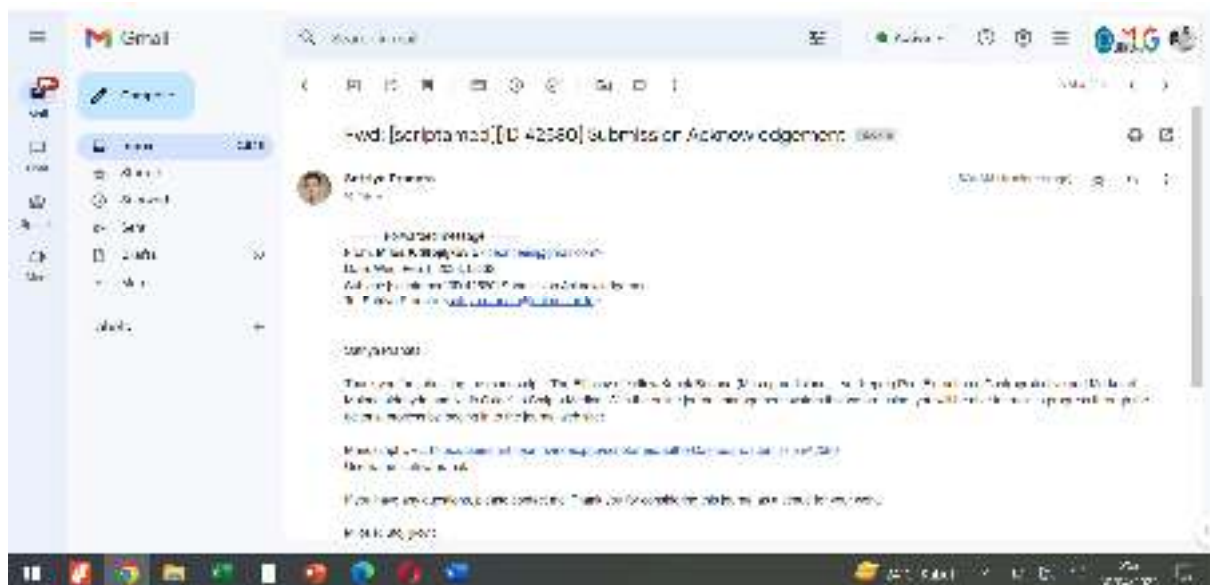
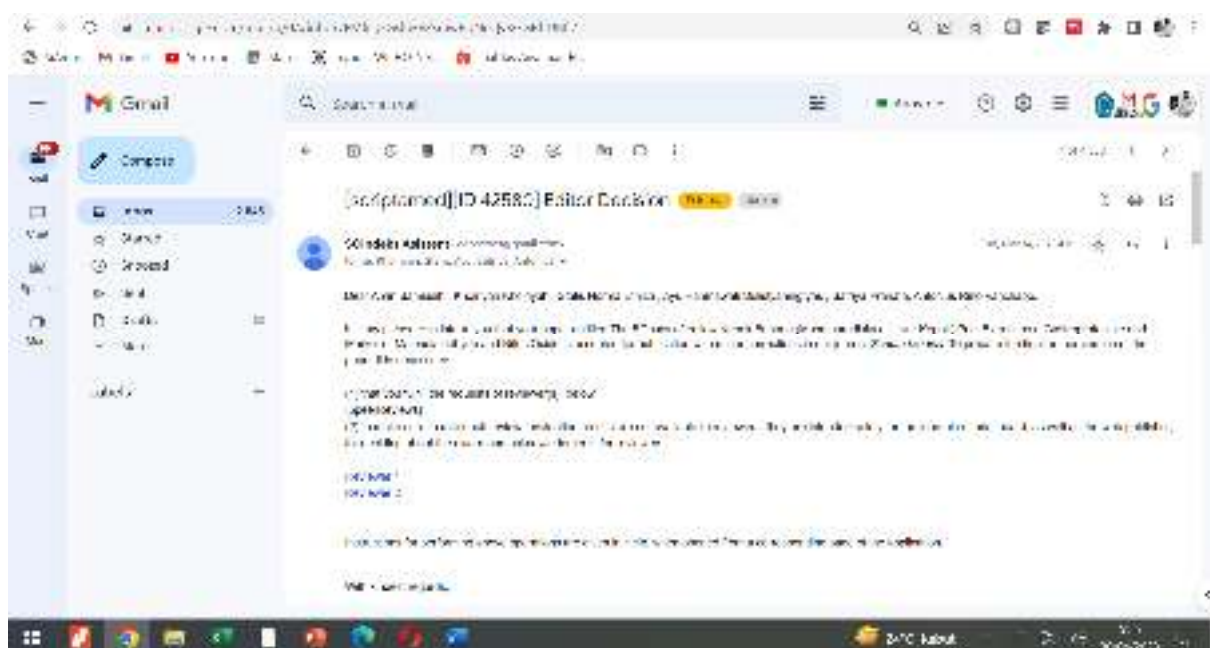


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1. submission



2. Review and revision



The Efficacy of Yellow Kepok Banana (*Musa paradisiaca* L. var Kepok) Peel Extract as a Gastroprotective and Marker of Malondialdehyde and Nitric Oxide

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Abstract

Background: Flavonoids, tannins, saponins, and polyphenols in Yellow Kepok banana (*Musa paradisiaca* L. var Kepok) peel potentially be a solution for peptic ulcer prevention. This study aims to prove the efficacy of Kepok banana peel extract on the gastroprotective effect of markers Malondialdehyde (MDA), Nitric Oxide (NO), and the number of gastric ulcers.

Methods: The research method was a pre-clinical experiment, and a post-test design. The sample was 33 female Wistar rats aged 3-4 months; body weighed 100-250 grams. Those rats were divided into 3 groups; Musa Paradisiaca Var Kepok 1 (MPVK1) treatment group, Musa Paradisiaca Var Kepok 2 (MPVK2), and control group (K). MPVK1 as the treatment group was given a dose of Kepok banana peel extract at a dose of 80mg/200gBW and the MPVK 2 group was 160mg/200gBW dose. Meanwhile, the gastritis induction using 5% acetosal at a dose of 1500 mg/kg BW. MDA

examination by HPLC method, NO examination by ELISA method, and macroscopic examination by counting the number of ulcers on the gastric mucosa.

Results: The results showed that the lowest average MDA level was in the MPVK2 group which was 3.27, whereas the highest average NO level was in the MPVK2 group 286.17. The highest mean number of ulcers was in the control group at 3.55. The conclusion is that there is a significant difference in the average levels of MDA (p-value 0.013), NO (p-value 0.000), and the number of ulcers (p-value 0.000) of the three groups.

Conclusion: Banana peel extract was proven to be used as a gastroprotective through markers of MDA, NO, and the number of ulcers.

Keywords: Banana peel, malondialdehyde, nitric oxide

Introduction

According to WHO (World Health Organization), the occurrence of gastritis in Indonesia is 40.8% with the incidence rate in several regions in Indonesia was quite high with a prevalence of 274,396 cases out of 238,452,952 people. Gastritis is one of the 10 most common diseases in hospitals with 33,154 cases (4.9%).^{1,2}

Gastritis is caused by hypersecretion of hydrochloric acid and pepsin which erode the lining of the gastrointestinal mucosa.^{3,4} When the stomach is exposed to gastric mucosal destroying agents (acetosal) there will be a back diffusion of H^+ from the lumen into the mucosa and causing a reaction that can harm the stomach and release a large amount of pepsin. Many sodiums (Na^+) and plasma proteins enter the lumen and release histamine.^{5,6} These results of escalation secretion of hydrochloric acid by parietal cells increased capillary permeability and bleeding. In addition, it will stimulate local

parasympathetic due to the increase of hydrochloric acid secretion so that venous congestion gets worse and eventually causes bleeding. If this condition is admissible to continue, superficial erosion or ulceration may occur. The process of gastritis is due to an imbalance between mucosal defences and several aggressive factors. One of them is caused by long-term consumption of Non-Steroid Anti-Inflammatory Drugs (NSAIDs). The exogenous aggressive factors that cause deterioration to the gastric mucosa ~~is-are~~ in the form of inflammation, or if it is chronic cause bleeding and perforation are NSAIDs.^{7,8}

The popular ways to prevent the formation of peptic ulcers is by administering drugs that function as cytoprotective on the gastric mucosa. Hence, ~~the~~ prevention efforts that have minimal side effects are needed, including the use of tropical plants ~~inas~~ the development of phytopharmaceutical use, considering that Indonesia is rich in a variety of medicinal plants.⁹⁻¹¹ Flavonoid antioxidants, tannins, saponins, and polyphenols have benefits as anti-inflammatory and antioxidant in hyper lipidemic DM rabbits.^{12,13}

Further, banana peel extract contains flavonoids, tannins, saponins, and polyphenols. Tannins ~~has~~ function to minimize gastric acid secretion and have a cytoprotective effect.^{9,11} Tannins also promote tissue reconstruction, inhibit gastric acid production, antioxidants, and inhibit the activity of *Helicobacter pylori*. Saponins inhibit gastric acid production and lower the pH levels of gastric juices.¹⁴ Based on this, the banana peel has potential as a gastroprotector.^{15,16} This study investigated the extract of the Yellow Kepok Banana (*Musa paradisiaca* L. var Kepok) peel as a gastroprotective in acetosal-induced Wistar rats. Thus, the aim of this study was to prove the effect of Kepok banana peel extract on gastritis through markers of oxidative stress (Malondialdehyde, Nitric Oxide) and the number of ulcers.

Methods

The Preparation of Kepok Banana Peel Extract

The Kepok banana (*Musa paradisiaca* L. var Kepok) peel extract was prepared to refer to the banana peel extraction procedure (Copyright Document No. HKI S00201809745) initiated with the preparation of the raw materials.

Extract characteristics

Kepok banana is one of the banana varieties in Indonesia Indonesia. Kepok bananas consist of white kepok bananas and yellow kepok bananas. The part used to be extracted in this study is the peels. The chopped banana peels are dried in an oven at 40°C for 24 hours until they are completely dry, characterized by a texture that is easily broken by hand squeezing. The dose extract is divided into 3 doses, namely kepok banana peel extract at a dose of 80mg/200gBW with 0.3% NaCMC solvent, 160mg/200gBW yellow Kepok banana peel extract with 0.3% NaCMC solvent, and Kepok banana peel extract with 0.3% NaCMC.

Phytochemical screening extract procedures

The banana peel was washed, roughly sliced, and then processed to dry by aerating. The drying process was carried out using an oven, and further mashed. The extraction method used 76% ethanol maceration.

Experimental Animal Care

The experimental animals ~~that~~ used in this study were Wistar rats kept in group cages sized 20 x 33 cm in the Experimental Animal Laboratory of Universitas Muhammadiyah Semarang. The environmental conditions of the cage were arranged ~~in of~~ 24-26°C temperature, supported by

sufficient ventilation. The feed was provided in the form of pellets, while its drinking water was provided ad libitum in the cannula bottles.

Experimental Animal Model Preparation

The gastritis experimental animal model was prepared by induction using 5% acetosal at a dose of 1500 mg/kg BW in one-time administration ⁷.

Research Design

This research is a type of pre-clinical research, post-test only design. The Experimental animal samples consisted of 33 female Wistar rats aged 3-4 months and ~~bodies weighed~~body-weighted 100-250 g.

Research Data Collection

This research was conducted for 18 days. Data collection was in the form of blood serum and counting the number of ulcers was carried out on the 18th day (post-test). The study began with the rats being adapted for 14 days. On the 15th day, the rats were divided into 3 groups: Musa Paradica Var Kepok 1 (MPVK1) treatment group, Musa Paradica Var kapok 2 (MPVK2) treatment group, and control group (K). The groups were divided using a simple random sampling technique. Treatment group 1 (MPVK 1) rats were given the yellow Kepok banana peel extract at a dose of 80mg/200gBW with 0.3% NaCMC solvent and treatment group 2 (MPVK 2) rats were given 160mg/200gBW yellow Kepok banana peel extract with 0.3% NaCMC solvent. %. The control group (K) was given 0.3% NaCMC. On the 16th day, the rats were enforced to not eat for 24 hours while still being given water ad libitum. On the 17th day, the rats in group K were given 0.3% NaCMC, meanwhile MPVK 1 group was given the yellow Kepok banana peel extract at a dose of 80mg/200gBW with 0.3% NaCMC solvent and the MPVK 2 group was given a yellow Kepok banana peel extract at a dose of 160mg/200g BW with 0.3% NaCMC solvent. After one hour, all rats were induced with 5% acetosal

at a dose of 1500 mg/kg BW. On the 18th day, the rats were anesthetized and its blood was taken through the orbital sinus, to be terminated. The stomach organs were taken and washed with 0.9% NaCl and the number of ulcers was counted. The Blood serum preparations were examined for markers of oxidative stress (MDA, NO). The MDA examination by HPLC method, NO examination by ELISA method, and ~~the macroscopic~~ the macroscopic examination by counting the number of ulcers on the gastric mucosa.

Data processing

The data with normal distribution ~~were~~ was tested by One-way ANOVA and Kruskal Wallis for abnormal data. The limit of the degree of significance was set on $\alpha < 0.05$ with 80% research power and 95% confidence intervention.

Ethical Consideration

The study was successfully accepted by Gadjah Mada University review board with institutional review board (IRB) number 00017/LPPT/VI/2021.

Results

Sample characteristics

The samples consisted of 33 female Wistar rats aged 3-4 months and ~~bodies~~ body ~~weighed~~ weighted 100-250 grams. They were grouped into MPVK 1, MPVK 2 dan K groups, the minimum weight of rats in the MPVK1 group was 170, and the maximum weight was 250 while the average was 207. The minimum body weight for rats in the MPVK2 group was 180, the maximum was 250 and the average was 217. At the same time, the minimum body weight for the rats in the control group (K) was 178, and its maximum of 250 and an average of 210, as shown in Figure 1.

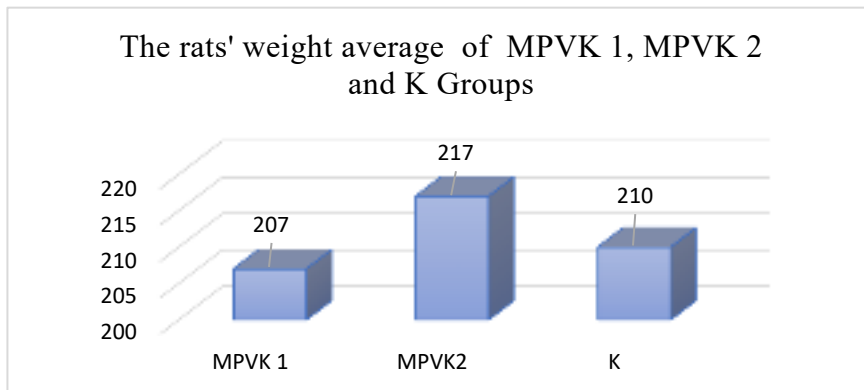


Figure 1. The average weight of 3 group rats used in this study; the MPVK 1, MPVK 2 dan K

The Average Levels of Malondialdehyde

The minimum level of Malondialdehyde (MDA) in the MPVK 1 group was 1.01, while the maximum level was 7.67, and its average was 2.60, respectively. The minimum level of Malondialdehyde (MDA) in the MPVK2 group was 1.5, while its maximum level was 2.10 and the average was 1.81. The minimum level of Malondialdehyde (MDA) in the control group (K) was 2.19, the maximum level was 3.93 and the average was 3.27. The lowest mean level of Malondialdehyde (MDA) was in the MPVK2 group, as shown in Figure 2.

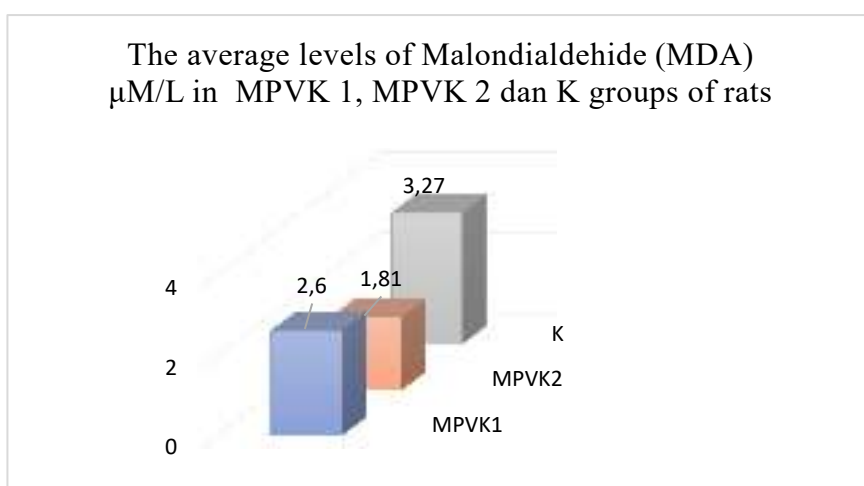


Figure 2. The Average levels of Malondialdehyde (MDA) of rats in MPVK 1, MPVK 2 and K Groups

The minimum level of Nitric Oxide (NO) in the MPVK1 group was 88, meanwhile the maximum level was 131.28 and the average was 112.73. The minimum level of Nitric Oxide (NO) in the MPVK2 group was 221.28, while the maximum level was 353.21 and the average was 286.17. The minimum level of Nitric Oxide (NO) in the control treatment group (K) was 101.95, meanwhile, the maximum level was 120.28 and the average was 111.74. The lowest average Nitric Oxide (NO) level was in group K, as shown in Figure 3.

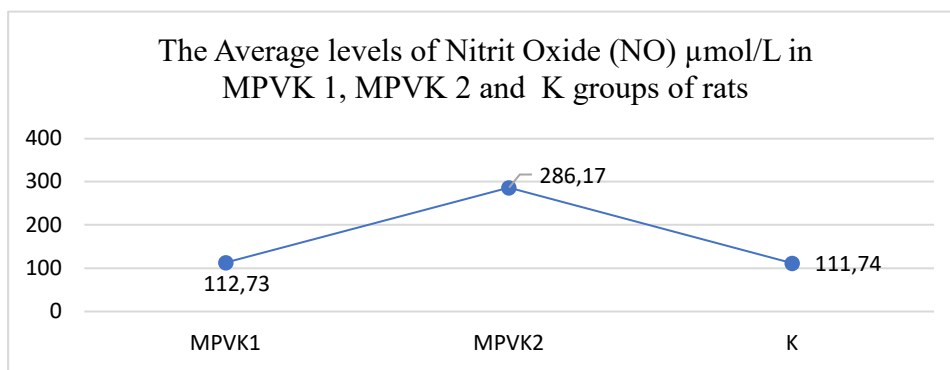


Figure 3. The Average levels of Nitric Oxide (NO) in MPVK 1, MPVK 2 and K Groups of Rats

The minimum number of ulcers in the MPVK1 group was 0, with its maximum was 3 and the average was 0.45. The number of ulcers in the MPVK2 group was 0, a maximum of 1 and its average was 0.04. The number of ulcers in the control group (K) was 0, a maximum of 7 and its average was 3.55. The lowest mean of number ulcers was in the MPVK 2 group, as seen in Figure 4

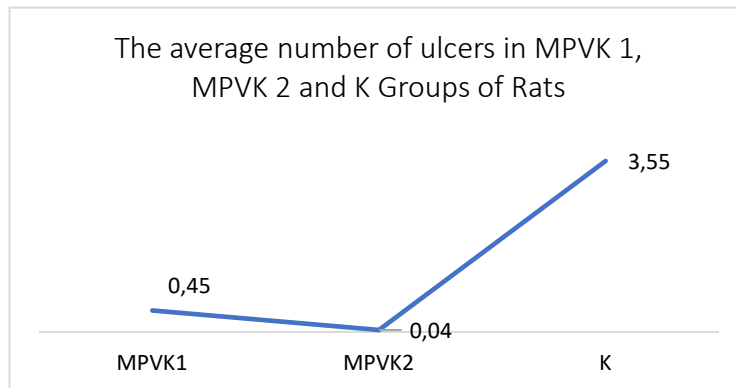


Figure 4. The average number of ulcers in the MPVK 1, MPVK 2 and K Groups of rats

Table 1. One-Way ANOVA test results has displayed the average values of MDA and NO, accompanied by the number of ulcers in the experimental groups of MPVK 1, MPVK 2 and K

	MPVK1	MPVK2	K	<i>p value</i>
MDA	2,6	1,81	3,27	0,013
NO	112,73	286,17	111,74	0,000
Number of	0,45	0,04	3,55	0,000

Table 1 shows the results of the One-Way ANOVA test. It revealed that there were significant distinctions in the average of MDA, NO and the number of ulcers in the 3 group of experimental rats; MPVK 1, MPVK 2 and K.

Discussion

An induction of free radicals and oxidative stress causes gastric mucosal deterioration.² Further, an imbalance of aggressive and defensive factors can cause gastric mucosal ulcers. The aggressive factor is more dominant than the defensive factor. The existence of free radicals is part of the aggressive factor.¹⁷ An example of free radicals is non-steroidal anti-inflammatory drugs (NSAIDs).

Acetylsalicylic acid or acetosal works by blocking certain natural substances in the body ~~is~~ to reduce pain and swelling. Acetosal is an irritant.^{4,5,18} Acetosal causes a defect in the mucosal barrier and back diffusion of H^+ ions will be occurred. Histamine is stimulated to secrete more gastric acid, resulting in dilation and increased permeability of capillaries, gastric mucosal damage, acute or chronic gastritis and gastric mucosal ulcers.¹⁹

The experimental animal model of gastric mucosal ulcers in this study was carried out by inducing Acetosal. Acetosal was given at a dose of 5% at a dose of 1500 mg/kg BW in all study groups. The results showed that the highest average number of ulcers was found in the control group. The minimum number of gastric ulcers in the control group (K) was 0, the maximum number of ulcers was 7 and the average ulcer was 3.55. A macroscopic picture of gastric mucosal deterioration is shown in Figure 5.

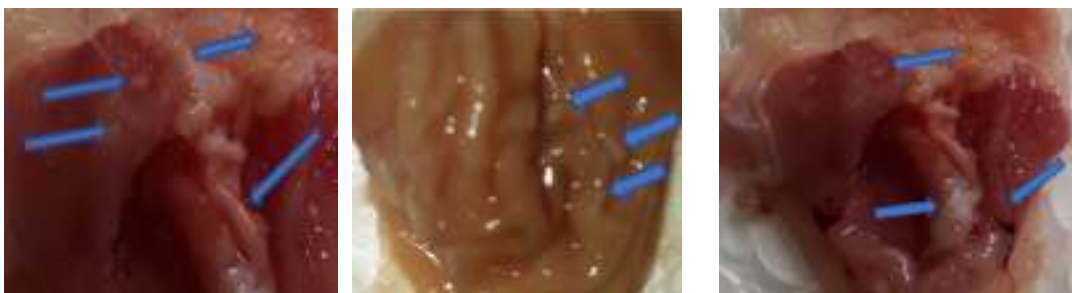


Figure 5. Gastric macroscopic characteristics that observed in the control group. The signs of hypoxemia, bleeding and gastric mucosal ulcers were displayed by the blue arrows.

The detrimental effect of free radicals that because biological damage is oxidative stress. Cells exposed to oxidative stress will activate defense mechanisms to survive.^{20,21} Lipid peroxidation itself is the result of the performance of free radicals which is very popular -and the most accessible to measure.^{2,11} Lipid peroxidase can damage membrane structures, leads changes in permeability, inhibits metabolic processes and transform ion transport as well.¹¹ The measurement of lipid

peroxidation level is carried out by measuring the final product, one of which is MDA. The accumulation of MDA is an early indicator of the mechanism of cell and tissue deterioration. Malondialdehyde (MDA) as final the product of the lipid peroxidation process is used as an indicator of cell deterioration in the stomach due to the oxidative stress.^{9,11,22}

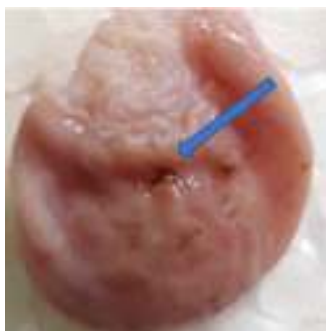
The results of this study ~~has~~have revealed that the minimum level of Malondialdehyde (MDA) in the control group (K) of -2.19 is the highest among those three ~~groups; MPVK~~groups; MPVK1 (01) and MPVK2 (1.5). The maximum level of Malondialdehyde (MDA) in the control group (K) was 3.93, that is the highest among MPVK1 (7.67) and MPVK2 (2.10). The average level of Malondialdehyde (MDA) in the control group (K) was 3.27, that is ~~the~~ highest among MPVK1 (2.60) and MPVK2 (1.81).

Kepok and Uli banana peel extracts had increased SOD activity and decreased MDA levels in hypercholesterolemic rats.²³ Kepok banana peel extracts decreased malondialdehyde levels in male mice (*mus musculus*) that ~~exposed~~exposure to cigarette smoke.¹⁴ Kepok banana peel contains flavonoid and phenolic antioxidants, the antioxidant content of flavonoids and phenolics is a hepatoprotection of aspirin-induced rats.²⁴ Other antioxidants component in Kepok banana peels are flavonoids, tannins, saponins and polyphenols. Tannins are useful in minimizing gastric acid secretion and have cytoprotective effect.⁵ Other benefits of tannins are the increasing tissue reconstruction, gastric acid production inhibition, and the activity of *Helicobacter pylori* inhibition. Further, saponins is also inhibit gastric acid production and minimize gastric fluid pH levels.^{8,16,25}

Kepok banana peel extract reduces oxidative stress in peptic ulcers through the antioxidant pathway.²⁴ Prevention and treatment of gastric mucosal ulcers by exploring natural products is something that is very ~~impressive~~impressing. The rats that ~~were given~~ were given -Kepok banana peel extract performed the increase of protection phenomena against acetosal induction by elevating the antioxidant level of Nitrit Oxide (NO). Acetosal converts hydroperoxyl to hydroxy fatty acids which is culminated from

lipid peroxidation of cell destruction. The release of damaging free radicals was likely occurred resulting in the death of tissue cells in the stomach. The ulceration effect of superficial epithelial cells on the gastric mucosa constructs the base of gastric ulceration.¹⁴ The gastroprotective effect of Kepok banana peel extract increases mucosal defence factors by increasing the body's antioxidant levels, Nitric Oxide (NO).^{14,19}

This study revealed that the results of the rats given Kepok banana peel extract performed the increase of the average level of Nitric Oxide (NO) antioxidants compared to the control group, namely MPVK1 (112.73) and MPVK2 (286.17). The lowest average Nitric Oxide (NO) level was found in the control group, that is 111.74. There is a significant distinction in the mean of Nitric Oxide (NO) in the three groups with (p-value) = 0.000. The results of gastric macroscopic examination in treatment of group 1 and 2 are shown in Figure 6.



Picture A. Minimal
bleeding for MPVK1
group



Picture B. Not perform
bleeding and ulcers for
MPVK 2 group



Picture C. Not perform
bleeding and ulcers for
MPVK 2 group

Figure 6. Gastric macroscopic characteristics was observed in the experimental groups; MPVK 1, MPVK 2 and K Groups of rats. A. Macroscopic characteristics of stomach in the MPVK1 group showing minimal bleeding (pointed by the blue arrow), with fewer ulcers than that in the control group B

and C, showing macroscopic characteristics of the stomach in the MPVK2 group which did not perform bleeding and ulcers.

The average number of ulcers in the treatment group was lower than in the control group. Hence, the mean number of ulcers in the MPVK1 group was 0.45. The average number of ulcers in the MPVK2 group was 0.04. The mean number of ulcers in control group (K) was 3.55. Meanwhile, the lowest mean number of ulcers was found in MPVK2 group. There was a significant distinction in the mean number of ulcers in the three groups with (p-value) = 0.000.

Conclusion

There is a significant distinction ~~of~~ in the mean value of Malondialdehyde (MDA), Nitric Oxide (NO) and the distinction of ulcers number among the three groups. Therefore, kepok banana peel extract has good efficacy in reducing the gastroprotective effect of markers Malondialdehyde (MDA), Nitric Oxide (NO), and the number of gastric ulcers in Wistar rats.

Acknowledgements

We would like to thank the ~~participants~~ Gadjah Mada University, Universitas Muhammadiyah Semarang and any parties who contribute in this study

Author contribution

AS conceptualized, designed, wrote the first draft and framework. SP, SNE conceptualized, interpreted the data and supervised. KK, ARS, ARV collected the data, evaluated the data. The published version of the manuscript has been read and approved by all authors.

Conflict of interest

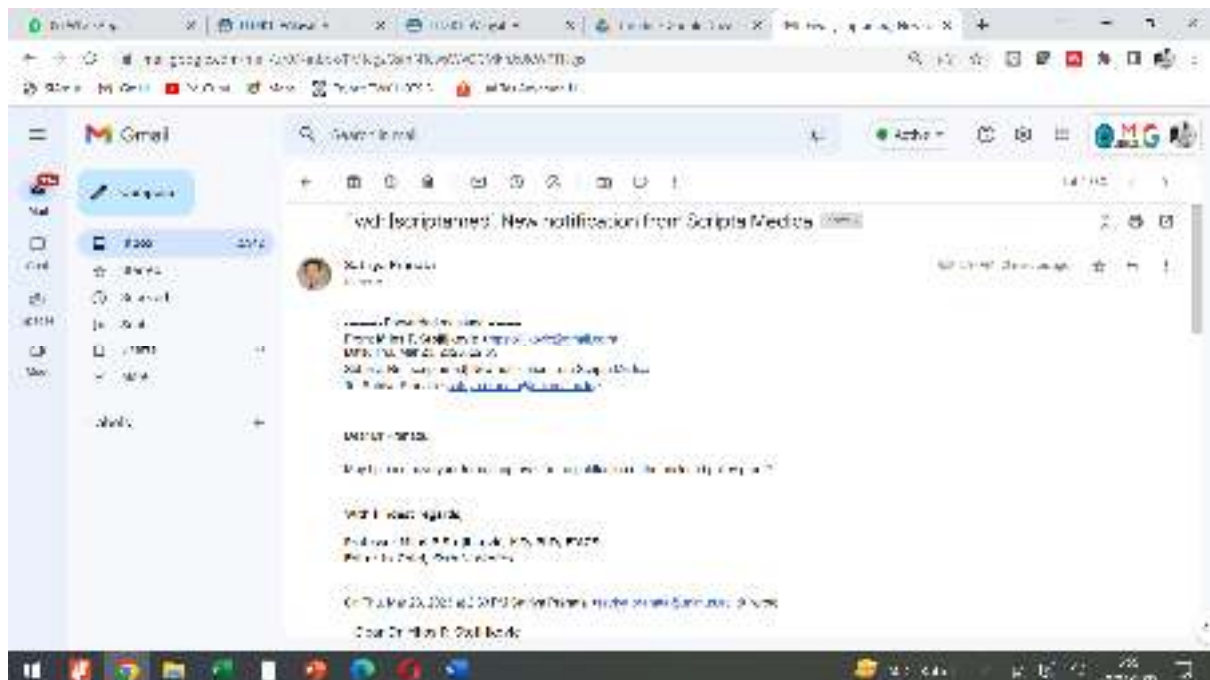
The authors declared no potential conflicts of interest in respect to the research, authorship, and publication of this article.

References

1. Tytgat GNJ. Etiopathogenetic Principles and Peptic Ulcer Disease Classification. Dig Dis [Internet]. 2011;29(5):454–8. Available from: <https://www.karger.com/DOI/10.1159/000331520>
2. Li K, Gesang L. Oxidative stress is associated with the gastric mucosa lesion of the Tibetans with high-altitude polycythemia. Int J Clin Exp Med. 2019;12(3):2503–10.
3. El-Meligy RM, Awaad AS, Soliman GA, Kenawy SA, Alqasoumi SI. Prophylactic and curative anti-ulcerogenic activity and the possible mechanisms of action of some desert plants. Saudi Pharm J SPJ Off Publ Saudi Pharm Soc [Internet]. 2016/10/19. 2017 Mar;25(3):387–96. Available from: <https://pubmed.ncbi.nlm.nih.gov/28344494>
4. Dionisio KL, Phillips K, Price PS, Grulke CM, Williams A, Biryol D, et al. The Chemical and Products Database, a resource for exposure-relevant data on chemicals in consumer products. Sci Data [Internet]. 2018;5(1):180125. Available from: <https://doi.org/10.1038/sdata.2018.125>
5. Zulaikhah ST. The Role of Antioxidant to Prevent Free Radicals in The Body. Sains Med. 2017;8(1):39.
6. Flower R. What are all the things that aspirin does? BMJ [Internet]. 2003 Sep 13;327(7415):572–3. Available from: <https://pubmed.ncbi.nlm.nih.gov/12969898>
7. Sharifi-Rad M, Anil Kumar N V., Zucca P, Varoni EM, Dini L, Panzarini E, et al. Lifestyle, Oxidative Stress, and Antioxidants: Back and Forth in the Pathophysiology of Chronic Diseases. Front Physiol. 2020;11(July):1–21.
8. Sharifi-Rad M, Fokou PVT, Sharopov F, Martorell M, Ademiluyi AO, Rajkovic J, et al. Antiulcer agents: From plant extracts to phytochemicals in healing promotion. Vol. 23, Molecules. 2018. 1–37 p.
9. Ayala A, Muñoz MF, Argüelles S. Lipid peroxidation: production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. Oxid Med Cell Longev [Internet]. 2014/05/08. 2014;2014:360438. Available from: <https://pubmed.ncbi.nlm.nih.gov/24999379>
10. Dhama K, Latheef SK, Dadar M, Samad HA, Munjal A, Khandia R, et al. Biomarkers in Stress Related Diseases/Disorders: Diagnostic, Prognostic, and Therapeutic Values. Front Mol Biosci [Internet]. 2019 Oct 18;6:91. Available from: <https://pubmed.ncbi.nlm.nih.gov/31750312>
11. Domínguez R, Pateiro M, Gagaoua M, Barba FJ, Zhang W, Lorenzo JM. A comprehensive review on lipid oxidation in meat and meat products. Antioxidants. 2019;8(10):1–31.
12. Tungmunnithum D, Thongboonyou A, Pholboon A, Yangsabai A. Flavonoids and Other Phenolic Compounds from Medicinal Plants for Pharmaceutical and Medical Aspects: An Overview. Med (Basel, Switzerland) [Internet]. 2018 Aug 25;5(3):93. Available from: <https://pubmed.ncbi.nlm.nih.gov/30149600>
13. Soesanto E, Pranata S, Rejeki S, Irham LM. The Role of Bamboo Shoot Gigantochloa Apus Extract in Decreasing the IL-17/IL-10 Ratio Level in the Atherosclerosis Process. Open Access Maced J Med Sci. 2021;9(A):817–21.
14. Amang AP, Mezui C, Siwe GT, Emakoua J, Mbah G, Nkwengoua EZ, et al. Healing and Antisecretory Effects of Aqueous Extract of Eremomastax speciosa (Acanthaceae) on Unhealed Gastric Ulcers. Biomed Res Int. 2017;2017.
15. Rao USM, Ahmad BA, Mohd KS, Zin T. Aktiviti antiulser dari ekstrak jantung dan kulit Musa paradisiaca (pisang) pada tikus albino teraruh ulser. Malaysian J Anal Sci. 2016;20(5):1203–16.

16. de Jesus NZT, de Souza Falcão H, Gomes IF, de Almeida Leite TJ, de Moraes Lima GR, Barbosa-Filho JM, et al. Tannins, peptic ulcers and related mechanisms. *Int J Mol Sci.* 2012;13(3):3203–28.
17. Tytgat G. Etiopathogenetic Principles and Peptic Ulcer Disease Classification. *Dig Dis.* 2011 Nov;29:454–8.
18. Flower R. What are all the things that aspirin does? *BMJ.* 2003 Sep;327(7415):572–3.
19. de Jesus NZT, de Souza Falcão H, Gomes IF, de Almeida Leite TJ, de Moraes Lima GR, Barbosa-Filho JM, et al. Tannins, peptic ulcers and related mechanisms. *Int J Mol Sci.* 2012;13(3):3203–28.
20. Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, Arcoraci V, et al. Oxidative Stress: Harms and Benefits for Human Health. *Oxid Med Cell Longev* [Internet]. 2017/07/27. 2017;2017:8416763. Available from: <https://pubmed.ncbi.nlm.nih.gov/28819546>
21. Sharifi-Rad M, Anil Kumar N V., Zucca P, Varoni EM, Dini L, Panzarini E, et al. Lifestyle, Oxidative Stress, and Antioxidants: Back and Forth in the Pathophysiology of Chronic Diseases. *Front Physiol.* 2020 Jul;0:694.
22. Dhama K, Latheef SK, Dadar M, Samad HA, Munjal A, Khandia R, et al. Biomarkers in Stress Related Diseases/Disorders: Diagnostic, Prognostic, and Therapeutic Values. *Front Mol Biosci.* 2019;0:91.
23. Ulfa A, Ekastuti DR, Wresdiyati T. Potensi Ekstrak Kulit Pisang Kepok (*Musa paradisiaca* forma *typica*) dan Uli (*Musa paradisiaca sapientum*) Menaikkan Aktivitas Superoksida Dismutase dan Menurunkan Kadar Malondialdehid Organ Hati Tikus Model Hiperkolesterolemia. *Acta Vet Indones.* 2020;8(1):40–6.
24. Yulis PAR, Sari Y. Aktivitas Antioksidan Kulit Pisang Muli (*Musa acuminata* linn) dan Kulit Pisang Kepok (*Musa paradisiaca formatypica*). *Al-Kimia.* 2020 Dec;8(2):189–200.
25. Mekonnen AN, Asrade Atnafie S, Wahab Atta MA. Evaluation of Antiulcer Activity of 80% Methanol Extract and Solvent Fractions of the Root of *Croton macrostachyus* Hocsht: Ex Del. (*Euphorbiaceae*) in Rodents. *Evidence-based Complement Altern Med.* 2020;2020.

3. Galey Proof





The Gastroprotective Role of Yellow Kepok Banana (*Musa x Paradisiaca* L. var. *Kepok*) Peel Extract and Influence on Markers of Oxidative Stress: Malondialdehyde and Nitric Oxide

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Abstract

Background/Aim: Flavonoids, tannins, saponins and polyphenols in yellow kepok banana (*Musa x paradisiaca* L. var. *kepok*) peel potentially could be a solution for peptic ulcer prevention. This study aimed to prove the efficacy of kepok banana peel extract as gastroprotective by analysing the number of gastric ulcers and markers of oxidative stress - malondialdehyde (MDA) and nitric oxide (NO).

Methods: The study was performed on 33 female Wistar rats aged 3-4 months, weighed 100-250 g. Rats were divided into 3 groups: Musa Paradisiaca Var Kepok 1 (MPVK1) treatment group, Musa Paradisiaca Var Kepok 2 (MPVK2) and control group (K). In MPVK1 kepok banana peel extract at a dose of 80 mg / 200 g body weight (BW) was given and the MPVK2 group dose was 160 mg / 200 g BW. The gastritis induction was performed by using 5 % acetylsalicylic acid at a dose of 1500 mg/kg BW. MDA examination by HPLC method, NO examination by ELISA method and macroscopic examination by counting the number of ulcers on the gastric mucosa was performed.

Results: The results showed that the lowest average MDA level, as well as the highest average NO level was in the MPVK2 group 3.27 and 286.17, respectively. The highest mean number of ulcers was in the control group 3.55. By analysing all the results it can be concluded that there is a significant difference in the average levels of MDA ($p = 0.013$), NO ($p < 0.001$) and the number of ulcers ($p < 0.001$) in the three groups.

Conclusion: Banana peel extract was proven to be effective as a gastroprotective through markers of MDA, NO and the number of ulcers in Wistar rats.

Key words: Banana peel; Malondialdehyde; Nitric oxide; Gastroprotection.

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Introduction

According to World Health Organization (WHO), the occurrence of gastritis in Indonesia is 40.8 % with quite high incidence rate in several regions in Indonesia, with a prevalence of 274,396 cases out of 238,452,952 people. Gastritis is one of

the 10 most common diseases in hospitals with 33,154 cases (4.9 %).^{1,2}

Gastritis is caused by hypersecretion of hydrochloric acid and pepsin which erode the lining of

the gastrointestinal mucosa.^{3, 4} When the stomach is exposed to gastric mucosal destroying agents (acetylsalicylic acid) there will be a back diffusion of H⁺ from the lumen into the mucosa, causing a reaction that can harm the stomach and release a large amount of pepsin. Many sodium (Na⁺) and plasma proteins enter the lumen and release histamine.^{5, 6} This results escalation in secretion of hydrochloric acid by parietal cells, increased capillary permeability and bleeding. In addition, it stimulate local parasympathetic system due to the increase of hydrochloric acid secretion so that venous congestion gets worse and eventually causes bleeding. If this condition is admissible to continue, superficial erosion or ulceration may occur. The process of gastritis is due to an imbalance between mucosal defences and several aggressive factors. One of them is caused by long-term consumption of non-steroid anti-inflammatory drugs (NSAIDs). The exogenous aggressive factors that cause deterioration to the gastric mucosa are in the form of inflammation, or if it is chronic inflammation it can cause bleeding and perforation.^{7, 8}

The popular way to prevent the formation of peptic ulcers is by administering drugs that function as cytoprotective on the gastric mucosa. Hence, prevention efforts that have minimal side effects are needed, including the use of tropical plants in the development of phytopharmaceutical use, considering that Indonesia is rich in a variety of medicinal plants.⁹⁻¹¹ Flavonoid antioxidants, tannins, saponins and polyphenols have benefits as anti-inflammatory and antioxidant in hyper lipidemic DM rabbits.^{12, 13}

Further, banana peel extract contains flavonoids, tannins, saponins and polyphenols. Tannins minimise gastric acid secretion and have a cytoprotective effect.^{9, 11} Tannins also promote tissue reconstruction, inhibit gastric acid production, act as antioxidants and inhibit the activity of *Helicobacter pylori*. Saponins inhibit gastric acid production and lower the pH levels of gastric juices.¹⁴ Based on this, the banana peel has potential as a gastroprotector.^{15, 16} This study investigated the extract of the yellow kepok banana (*Musa paradisiaca* L. var *Kepok*) peel as a gastroprotective in acetylsalicylic acid-induced Wistar rats. Thus, the aim of this study was to prove the effect of kepok banana peel extract on gastritis through markers of oxidative stress (malondialdehyde (MDA), nitric oxide - NO) and the number of ulcers.

Methods

The preparation of kepok banana peel extract

The kepok banana (*Musa x paradisiaca* L. var. *kepok*) peel extract was prepared to refer to the banana peel extraction procedure (Copyright Document No HKI S00201809745) initiated with the preparation of the raw materials.

Extract characteristics

Kepok banana is one of the banana varieties in Indonesia. Kepok bananas consist of white kepok bananas and yellow kepok bananas. The part used to be extracted in this study is the peels. The chopped banana peels were dried in an oven at 40 °C for 24 h until they were completely dry, characterised by a texture that was easily broken by hand squeezing. Extract was provided in two doses, namely kepok banana peel extract at a dose of 80 mg / 200 g body weight (BW) with 0.3 % Sodium carboxymethyl cellulose (NaCMC) solvent, 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent and control with 0.3 % NaCMC.

Phytochemical screening extract procedures

The banana peel was washed, roughly sliced and then processed to dry by aerating. The drying process was carried out using an oven and further mashed. The extraction method used 76 % ethanol maceration.

Experimental animals

The experimental animals used in this study were Wistar rats kept in group cages sized 20 x 33 cm in the Experimental Animal Laboratory of Universitas Muhammadiyah Semarang, Indonesia. The environmental conditions of the cage were arranged at 24-26 °C, supported by sufficient ventilation. The food was provided in the form of pellets, while its drinking water was provided *ad libitum* in the cannula bottles.

Experimental model

The experimental animal samples consisted of 33 female Wistar rats aged 3-4 months weighed 100-250 g. The gastritis experimental animal model was prepared by induction using 5 % acetylsalicylic acid at a dose of 1500 mg/kg BW in one-time administration.⁷

This research was conducted for 18 days. Data collection were in the form of blood serum and counting the number of ulcers and it was carried out on the 18th day (post-test). The study began with the rats being adapted for 14 days. On the

15th day, the rats were divided into 3 groups: Musa Paradica Var Kepok 1 (MPVK1) treatment group, Musa Paradica Var Kapok 2 (MPVK2) treatment group and control group (K). The groups were divided using a random sampling technique. Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC. On the 16th day, the rats were enforced to not eat for 24 h while still being given water *ad libitum*. On the 17th day, the rats in group K were given 0.3 % NaCMC, meanwhile MPVK1 group was given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and the MPVK2 group was given a yellow kepok banana peel extract at a dose of 160 mg / 200 g BW with 0.3 % NaCMC solvent. After one hour, all rats were induced with 5 % acetylsalicylic acid at a dose of 1500 mg/kg BW. On the 18th day, the rats were anesthetised and blood was taken through the orbital sinus, to be terminated. The stomach organs were taken and washed with 0.9 % NaCl and the number of ulcers was counted. The blood serum preparations were examined for markers of oxidative stress (MDA, NO). The MDA examination by high-performance liquid chromatography (HPLC) method, NO examination by enzyme-linked immunosorbent assay (ELISA) method and the macroscopic examination by counting the number of ulcers on the gastric mucosa were performed.

Data processing

The normality of data distribution was tested and appropriate tests were used: One-way ANOVA for data with normal distribution and Kruskal-Wallis for abnormal data. The limit of the degree of significance was set at $p < 0.05$ with 80 % research power and 95 % confidence interval.

Ethical Consideration

The study was successfully accepted by Gadjah Mada University, Indonesia review board with institutional review board (IRB) decision No 00017/LPPT/VI/2021.

Results

Sample characteristics

The samples consisted of 33 female Wistar rats aged 3-4 months weighed 100-250 g. They were

grouped into MPVK1, MPVK2 and K group, the minimum weight of rats in the MPVK1 group was 170, the maximum weight was 250 while the average was 207. The minimum body weight for rats in the MPVK2 group was 180, the maximum was 250 and the average was 217. At the same time, the minimum body weight for the rats in the control group (K) was 178, the maximum was 250 and the average was 210, as shown in Figure 1.

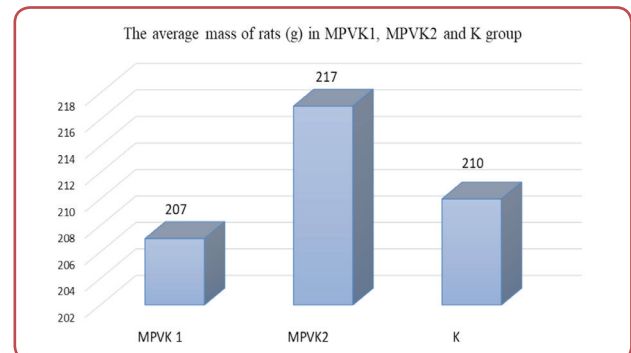


Figure 1: The average weight of Wistar rats in the MPVK 1, MPVK 2 and K group

Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

The minimum level of MDA in the MPVK 1 group was 1.01, while the maximum level was 7.67 and its average was 2.60, respectively. The minimum level of MDA in the MPVK2 group was 1.5, while its maximum level was 2.10 and the average was 1.81. The minimum level of MDA in the control group (K) was 2.19, the maximum level was 3.93 and the average was 3.27. The lowest mean level of MDA was in the MPVK2 group, as shown in Figure 2.

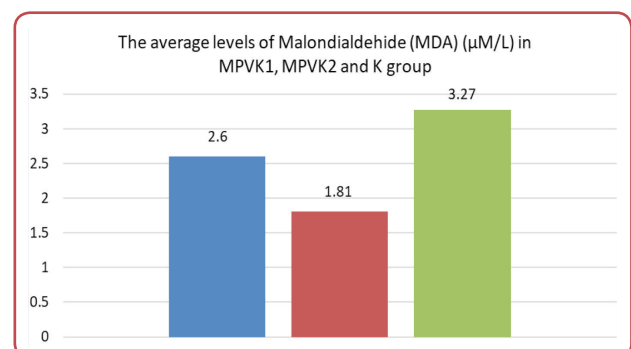


Figure 2: The average levels of malondialdehyde (MDA) of rats in MPVK1, MPVK2 and K group

Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

The average levels of nitric oxide (NO)

The minimum level of NO in the MPVK1 group

was 88, meanwhile the maximum level was 131.28 and the average was 112.73. The minimum level of NO in the MPVK2 group was 221.28, while the maximum level was 353.21 and the average was 286.17. The minimum level of NO in the K group was 101.95, meanwhile, the maximum level was 120.28 and the average was 111.74. The lowest average NO level was in group K, as shown in Figure 3.

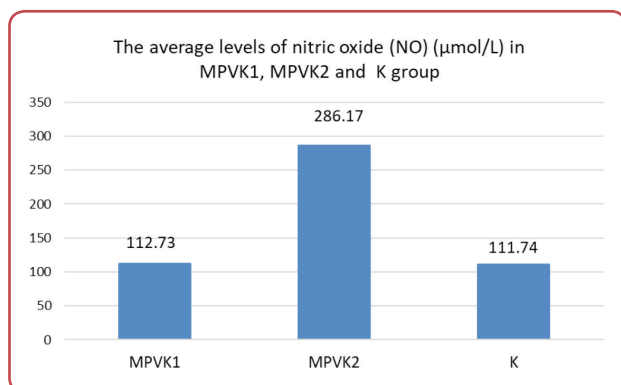


Figure 3: The average levels of nitric oxide (NO) in MPVK 1, MPVK 2 and K group

Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

The average number of ulcers

The minimum number of ulcers in the MPVK1 group was 0, with its maximum was 3 and the average was 0.45. The number of ulcers in the MPVK2 group was 0, a maximum of 1 and its average was 0.04. The number of ulcers in the control group (K) was 0, a maximum of 7 and its average was 3.55. The lowest mean of number ulcers was in the MPVK 2 group, as seen in Figure 4.

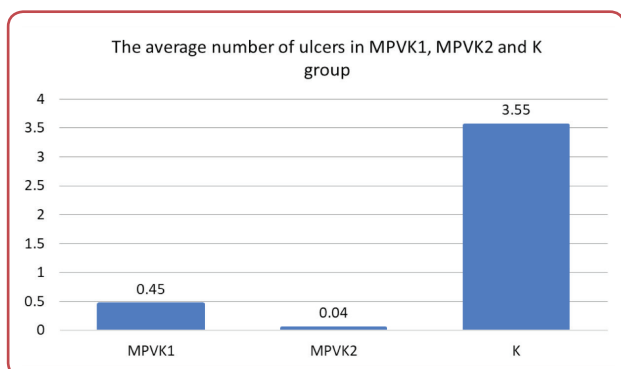


Figure 4: The average number of ulcers in the MPVK1, MPVK2 and K group

Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

A macroscopic picture of gastric mucosal deterioration is shown in Figure 5. The results showed that the highest average number of ulcers was found in the control group.

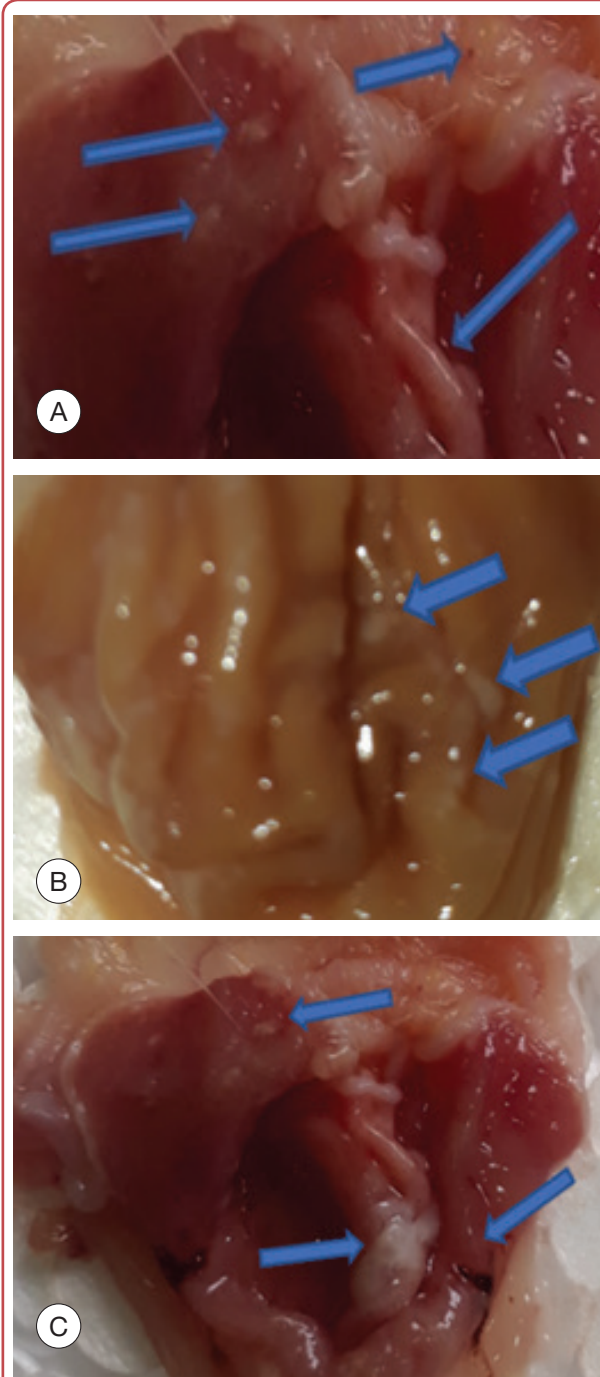


Figure 5 (A, B, C): Gastric macroscopic characteristics observed in the control group. The signs of hypoxaemia, bleeding and gastric mucosal ulcers were displayed by the blue arrows

The control group rats were given 0.3 % NaCMC.

The results of gastric macroscopic examination in treatment of group 1 and 2 are shown in Figure 6.

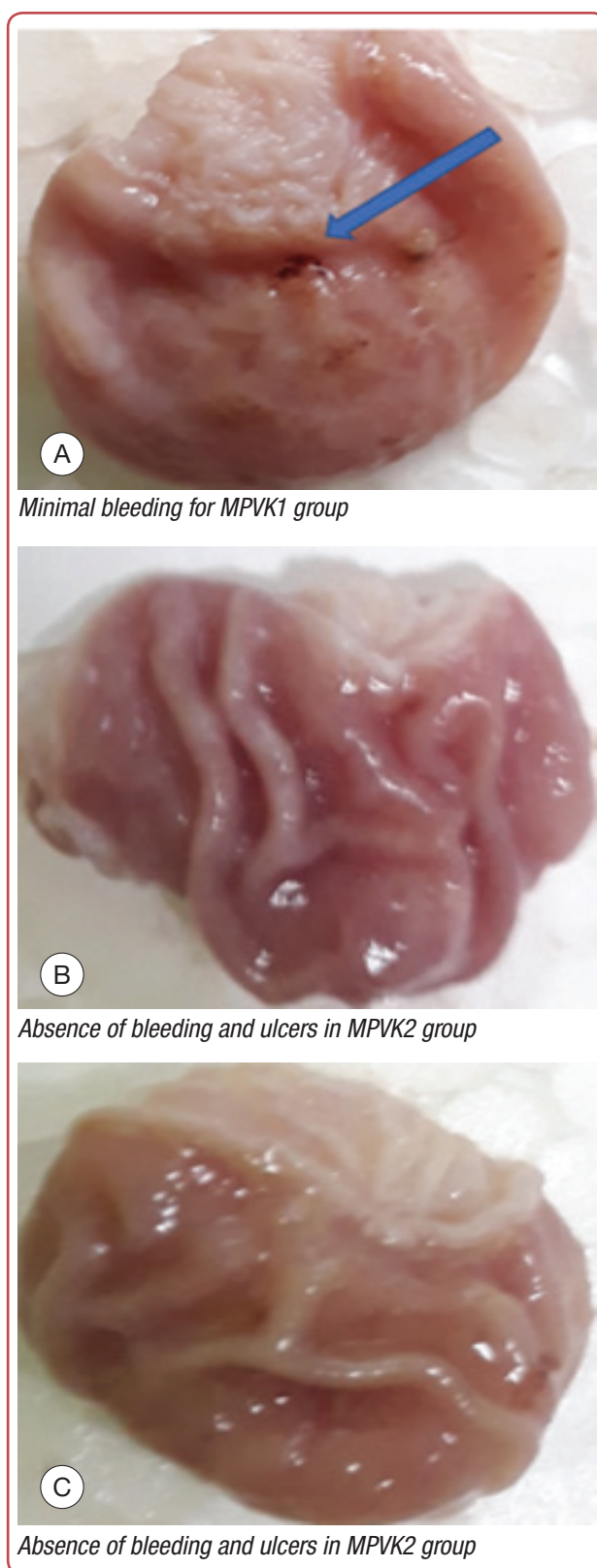


Figure 6: Gastric macroscopic characteristics observed in the treatment groups

A. Macroscopic characteristics of stomach in the MPVK1 group showing minimal bleeding (pointed by the blue arrow), with fewer ulcers than that in the control group; B and C, showing macroscopic characteristics of the stomach in the MPVK2 group which did not perform bleeding and ulcers. Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given the yellow kepok banana peel extract at a dose of 160 mg / 200 g BW with 0.3 % NaCMC solvent.

Table 1 shows the results of the One-Way ANOVA test. It revealed that there were significant distinctions in the average level of MDA and NO and the number of ulcers in the 3 group of experimental rats; MPVK 1, MPVK 2 and K.

Table 1: One-Way ANOVA test results has displayed the average values of MDA and NO, accompanied by the number of ulcers in the experimental groups MPVK1 and MPVK 2 and control group K

Parameter	MPVK1	MPVK2	K	p-value
MDA	2.60	1.81	3.27	0.013
NO	112.73	286.17	111.74	< 0.001
Number of ulcers	0.45	0.04	3.55	< 0.001

MDA: malondialdehyde; NO: nitric oxide; Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

Discussion

An induction of free radicals and oxidative stress causes gastric mucosal deterioration.² Further, an imbalance of aggressive and defensive factors can cause gastric mucosal ulcers. The aggressive factor is more dominant than the defensive factor. The existence of free radicals is part of the aggressive factor.¹ An example of free radicals is NSAIDs. Acetylsalicylic acid works by blocking certain natural substances in the body to reduce pain and swelling. Acetylsalicylic acid is an irritant.⁴⁻⁶ Acetylsalicylic acid causes a defect in the mucosal barrier and back diffusion of H⁺ ions occurs. Histamine is stimulated to secrete more gastric acid, resulting in dilation and increased permeability of capillaries, gastric mucosal damage, acute or chronic gastritis and gastric mucosal ulcers.¹⁶

The experimental animal model of gastric mucosal ulcers in this study was carried out by inducing acetylsalicylic acid. 5% acetylsalicylic acid was given at a dose of 1500 mg/kg BW in all study groups. The results showed that the highest average number of ulcers was found in the control group. The minimum number of gastric ulcers in the control group (K) was 0, the maximum number of ulcers was 7 and the average ulcer was 3.55.

The detrimental effect of free radicals that cause biological damage is oxidative stress. Cells exposed to oxidative stress will activate defence mechanisms to survive.^{7, 17} Lipid peroxidation itself is the result of the performance of free radicals and this parameter is the most accessible to measure.^{2, 11} Lipid peroxidase can damage membrane structures, it leads changes in permeability, inhibits metabolic processes and transform ion transport as well.¹¹ The measurement of lipid peroxidation level is carried out by measuring the final product, one of which is MDA. The accumulation of MDA is an early indicator of the mechanism of cell and tissue deterioration. MDA as final product of the lipid peroxidation process is used as an indicator of cell deterioration in the stomach due to the oxidative stress.⁹⁻¹¹

The results of this study have revealed that the minimum level of MDA in the control group (K) of 2.19 is the highest among those three groups, as well as the maximum and the average level of MDA.

Kepok and Uli banana peel extracts had increased superoxide dismutase (SOD) activity and decreased MDA levels in hypercholesterolemic rats.¹⁸ Kepok banana peel extracts decreased MDA levels in male mice (*mus musculus*) that was exposed to cigarette smoke.¹⁴ Kepok banana peel contains flavonoid and phenolic antioxidants, the antioxidant content of flavonoids and phenolics showed hepatoprotection in acetylsalicylic acid-induced rats.¹⁹ Other antioxidants component in kepok banana peels are flavonoids, tannins, saponins and polyphenols. Tannins are useful in minimising gastric acid secretion and have cytoprotective effect.⁵ Other benefits of tannins are the increasing tissue reconstruction, gastric acid production inhibition and the inhibition of *Helicobacter pylori* activity. Further, saponins also inhibit gastric acid production and minimise gastric fluid pH levels.^{8, 16, 20}

Kepok banana peel extract reduces oxidative stress in peptic ulcers through the antioxidant pathway.¹⁹ Prevention and treatment of gastric mucosal ulcers by exploring natural products is something that is very impressive. The rats that were given kepok banana peel extract performed the increase of protection phenomena against acetylsalicylic acid induction by elevating the antioxidant level of NO. Acetylsalicylic acid converts hydroperoxyl to hydroxy fatty acids which is culminated from lipid peroxidation of cell

destruction. The release of damaging free radicals likely occurs resulting in the death of tissue cells in the stomach. The ulceration effect of superficial epithelial cells on the gastric mucosa constructs the base of gastric ulceration.¹⁴ The kepok banana peel extract exhibits its gastroprotection by increasing mucosal defence factors by increasing the body's antioxidant levels, NO.^{14, 16}

This study revealed that the rats given kepok banana peel extract had increased average level of NO antioxidants compared to the control group, namely MPVK1 (112.73) and MPVK2 (286.17). The lowest average NO level was found in the control group, that is 111.74. There is a significant distinction in the mean of NO in the three groups with $p < 0.001$. The average number of ulcers in the treatment group was lower than in the control group. Hence, the mean number of ulcers in the MPVK1 group was 0.45. The average number of ulcers in the MPVK2 group was 0.04. The mean number of ulcers in control group (K) was 3.55. Meanwhile, the lowest mean number of ulcers was found in MPVK2 group. There was a significant distinction in the mean number of ulcers in the three groups with $p < 0.001$.

Conclusion

There is a significant distinction in the mean value of MDA, NO as well as the distinction of number of ulcers among the three groups. Therefore, kepok banana peel extract has good efficacy in reducing the markers of gastric damage in Wistar rats.

Acknowledgements

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Conflict of interest

None.

References

1. Tytgat GN. Etiopathogenetic principles and peptic ulcer disease classification. *Dig Dis* 2011;29(5):454-8.
2. Li K, Gesang L. Oxidative stress is associated with the gastric mucosa lesion of the Tibetans with high-altitude polycythemia. *Int J Clin Exp Med* 2019;12(3):2503-10.
3. El-Meligy RM, Awaad AS, Soliman GA, Kenawy SA, Alqasoumi SI. Prophylactic and curative anti-ulcerogenic activity and the possible mechanisms of action of some desert plants. *Saudi Pharm J* 2017 Mar;25(3):387-96.
4. Dionisio KL, Phillips K, Price PS, Grulke CM, Williams A, Biryol D, et al. The Chemical and Products Database, a resource for exposure-relevant data on chemicals in consumer products. *Sci Data* 2018 Jul 10;5:180125. doi: 10.1038/sdata.2018.125.
5. Zulaikhah ST. The role of antioxidant to prevent free radicals in the body. *Sains Med* 2017;8(1):39. doi: 10.30659/sainsmed.v8i1.1012.
6. Flower R. What are all the things that aspirin does? *BMJ* 2003 Sep 13;327(7415):572-3.
7. Sharifi-Rad M, Anil Kumar NV, Zucca P, Varoni EM, Dini L, Panzarini E, et al. Lifestyle, oxidative stress, and antioxidants: back and forth in the pathophysiology of chronic diseases. *Front Physiol* 2020 Jul 2;11:694. doi: 10.3389/fphys.2020.00694.
8. Sharifi-Rad M, Fokou PVT, Sharopov F, Martorell M, Ademiluyi AO, Rajkovic J, et al. Antiulcer agents: From plant extracts to phytochemicals in healing promotion. *Molecules* 2018 Jul 17;23(7):1751. doi: 10.3390/molecules23071751.
9. Ayala A, Muñoz MF, Argüelles S. Lipid peroxidation: production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. *Oxid Med Cell Longev* 2014;2014:360438. doi: 10.1155/2014/360438.
10. Dhama K, Latheef SK, Dadar M, Samad HA, Munjal A, Khandia R, et al. Biomarkers in stress related diseases/disorders: diagnostic, prognostic, and therapeutic values. *Front Mol Biosci* 2019 Oct 18;6:91. doi: 10.3389/fmolb.2019.00091.
11. Domínguez R, Pateiro M, Gagaoua M, Barba FJ, Zhang W, Lorenzo JM. A comprehensive review on lipid oxidation in meat and meat products. *Antioxidants* 2019;8(10):1-31.
12. Tungmunthum D, Thongboonyou A, Pholboon A, Yangsabai A. Flavonoids and other phenolic compounds from medicinal plants for pharmaceutical and medical aspects: an overview. *Medicines (Basel)* 2018 Aug 25;5(3):93. doi: 10.3390/medicines5030093.
13. Soesanto E, Pranata S, Rejeki S, Irham LM. The role of bamboo shoot *Gigantochloa Apus* extract in decreasing the IL-17/IL-10 ratio level in the atherosclerosis process. *Open Access Maced J Med Sci* 2021;9(A):817-21.
14. Amang AP, Mezui C, Siwe GT, Emakoua J, Mbah G, Nkwengoua EZ, et al. Healing and antisecretory effects of aqueous extract of *Eremomastax speciosa* (Acanthaceae) on unhealed gastric ulcers. *Biomed Res Int* 2017;2017:1924320. doi: 10.1155/2017/1924320.
15. Rao USM, Ahmad BA, Mohd KS, Zin T. Antiulcer activity of *Musa paradisiaca* (banana) tepal and skin extracts in ulcer induced albino mice. *Malaysian J Anal Sci* 2016;20(5):1203-16.
16. de Jesus NZT, de Souza Falcão H, Gomes IF, de Almeida Leite TJ, de Moraes Lima GR, Barbosa-Filho JM, et al. Tannins, peptic ulcers and related mechanisms. *Int J Mol Sci* 2012;13(3):3203-28.
17. Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, Arcoraci V, et al. Oxidative stress: harms and benefits for human health. *Oxid Med Cell Longev* 2017;2017:8416763. doi: 10.1155/2017/8416763.
18. Ulfa A, Ekastuti DR, Wresdiyati T. [The Potency of Kepok Banana peel Extract (*Musa paradisiaca* forma typica) and Uli (*Musa paradisiaca sapientum*) in increasing superoxide dismutase activity and reducing malondialdehyde levels in the liver of hypercholesterolemic rats mode]. *Acta Vet Indones* 2020;8(1):40-6. Indonesian.
19. Yulis PAR, Sari Y. [Antioxidant activity of muli banana peel (*Musa acuminata* linn) and kepok banana peel (*Musa paradisiaca* formatypica)]. *Al-Kimia* 2020 Dec;8(2):189-200. Indonesian.
20. Mekonnen AN, Asrade Atnafie S, Wahab Atta MA. Evaluation of antiulcer activity of 80% methanol extract and solvent fractions of the root of *Croton macrostachyus* Hocsht: Ex Del. (Euphorbiaceae) in rodents. *Evid Based Complement Alternat Med* 2020 Apr 9;2020:2809270. doi: 10.1155/2020/2809270.



The Gastroprotective Role of Yellow Kepok Banana (*Musa x Paradisiaca* L. var. *Kepok*) Peel Extract and Influence on Markers of Oxidative Stress: Malondialdehyde and Nitric Oxide

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Abstract

Background/Aim: Flavonoids, tannins, saponins and polyphenols in yellow kepok banana (*Musa x paradisiaca* L. var. *kepok*) peel potentially could be a solution for peptic ulcer prevention. This study aimed to prove the efficacy of kepok banana peel extract as gastroprotective by analysing the number of gastric ulcers and markers of oxidative stress - malondialdehyde (MDA) and nitric oxide (NO).

Methods: The study was performed on 33 female Wistar rats aged 3-4 months, weighed 100-250 g. Rats were divided into 3 groups: Musa Paradisiaca Var Kepok 1 (MPVK1) treatment group, Musa Paradisiaca Var Kepok 2 (MPVK2) and control group (K). In MPVK1 kepok banana peel extract at a dose of 80 mg / 200 g body weight (BW) was given and the MPVK2 group dose was 160 mg / 200 g BW. The gastritis induction was performed by using 5 % acetylsalicylic acid at a dose of 1500 mg/kg BW. MDA examination by HPLC method, NO examination by ELISA method and macroscopic examination by counting the number of ulcers on the gastric mucosa was performed.

Results: The results showed that the lowest average MDA level, as well as the highest average NO level was in the MPVK2 group 3.27 and 286.17, respectively. The highest mean number of ulcers was in the control group 3.55. By analysing all the results it can be concluded that there is a significant difference in the average levels of MDA ($p = 0.013$), NO ($p < 0.001$) and the number of ulcers ($p < 0.001$) in the three groups.

Conclusion: Banana peel extract was proven to be effective as a gastroprotective through markers of MDA, NO and the number of ulcers in Wistar rats.

Key words: Banana peel; Malondialdehyde; Nitric oxide; Gastroprotection.

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Introduction

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the 10 most common diseases in hospitals with 33,154 cases (4.9 %).^{1,2}

Gastritis is caused by hypersecretion of hydrochloric acid and pepsin which erode the lining of

the gastrointestinal mucosa.^{3, 4} When the stomach is exposed to gastric mucosal destroying agents (acetylsalicylic acid) there will be a back diffusion of H⁺ from the lumen into the mucosa, causing a reaction that can harm the stomach and release a large amount of pepsin. Many sodium (Na⁺) and plasma proteins enter the lumen and release histamine.^{5, 6} This results escalation in secretion of hydrochloric acid by parietal cells, increased capillary permeability and bleeding. In addition, it stimulate local parasympathetic system due to the increase of hydrochloric acid secretion so that venous congestion gets worse and eventually causes bleeding. If this condition is admissible to continue, superficial erosion or ulceration may occur. The process of gastritis is due to an imbalance between mucosal defences and several aggressive factors. One of them is caused by long-term consumption of non-steroid anti-inflammatory drugs (NSAIDs). The exogenous aggressive factors that cause deterioration to the gastric mucosa are in the form of inflammation, or if it is chronic inflammation it can cause bleeding and perforation.^{7, 8}

The popular way to prevent the formation of peptic ulcers is by administering drugs that function as cytoprotective on the gastric mucosa. Hence, prevention efforts that have minimal side effects are needed, including the use of tropical plants in the development of phytopharmaceutical use, considering that Indonesia is rich in a variety of medicinal plants.⁹⁻¹¹ Flavonoid antioxidants, tannins, saponins and polyphenols have benefits as anti-inflammatory and antioxidant in hyper lipidemic DM rabbits.^{12, 13}

Further, banana peel extract contains flavonoids, tannins, saponins and polyphenols. Tannins minimise gastric acid secretion and have a cytoprotective effect.^{9, 11} Tannins also promote tissue reconstruction, inhibit gastric acid production, act as antioxidants and inhibit the activity of *Helicobacter pylori*. Saponins inhibit gastric acid production and lower the pH levels of gastric juices.¹⁴ Based on this, the banana peel has potential as a gastroprotector.^{15, 16} This study investigated the extract of the yellow kepok banana (*Musa paradisiaca* L. var *Kepok*) peel as a gastroprotective in acetylsalicylic acid-induced Wistar rats. Thus, the aim of this study was to prove the effect of kepok banana peel extract on gastritis through markers of oxidative stress (malondialdehyde (MDA), nitric oxide - NO) and the number of ulcers.

Methods

The preparation of kepok banana peel extract

The kepok banana (*Musa x paradisiaca* L. var. *kepok*) peel extract was prepared to refer to the banana peel extraction procedure (Copyright Document No HKI S00201809745) initiated with the preparation of the raw materials.

Extract characteristics

Kepok banana is one of the banana varieties in Indonesia. Kepok bananas consist of white kepok bananas and yellow kepok bananas. The part used to be extracted in this study is the peels. The chopped banana peels were dried in an oven at 40 °C for 24 h until they were completely dry, characterised by a texture that was easily broken by hand squeezing. Extract was provided in two doses, namely kepok banana peel extract at a dose of 80 mg / 200 g body weight (BW) with 0.3 % Sodium carboxymethyl cellulose (NaCMC) solvent, 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent and control with 0.3 % NaCMC.

Phytochemical screening extract procedures

The banana peel was washed, roughly sliced and then processed to dry by aerating. The drying process was carried out using an oven and further mashed. The extraction method used 76 % ethanol maceration.

Experimental animals

The experimental animals used in this study were Wistar rats kept in group cages sized 20 x 33 cm in the Experimental Animal Laboratory of Universitas Muhammadiyah Semarang, Indonesia. The environmental conditions of the cage were arranged at 24-26 °C, supported by sufficient ventilation. The food was provided in the form of pellets, while its drinking water was provided *ad libitum* in the cannula bottles.

Experimental model

The experimental animal samples consisted of 33 female Wistar rats aged 3-4 months weighed 100-250 g. The gastritis experimental animal model was prepared by induction using 5 % acetylsalicylic acid at a dose of 1500 mg/kg BW in one-time administration.⁷

This research was conducted for 18 days. Data collection were in the form of blood serum and counting the number of ulcers and it was carried out on the 18th day (post-test). The study began with the rats being adapted for 14 days. On the

15th day, the rats were divided into 3 groups: Musa Paradica Var Kepok 1 (MPVK1) treatment group, Musa Paradica Var Kapok 2 (MPVK2) treatment group and control group (K). The groups were divided using a random sampling technique. Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC. On the 16th day, the rats were enforced to not eat for 24 h while still being given water *ad libitum*. On the 17th day, the rats in group K were given 0.3 % NaCMC, meanwhile MPVK1 group was given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and the MPVK2 group was given a yellow kepok banana peel extract at a dose of 160 mg / 200 g BW with 0.3 % NaCMC solvent. After one hour, all rats were induced with 5 % acetylsalicylic acid at a dose of 1500 mg/kg BW. On the 18th day, the rats were anesthetised and blood was taken through the orbital sinus, to be terminated. The stomach organs were taken and washed with 0.9 % NaCl and the number of ulcers was counted. The blood serum preparations were examined for markers of oxidative stress (MDA, NO). The MDA examination by high-performance liquid chromatography (HPLC) method, NO examination by enzyme-linked immunosorbent assay (ELISA) method and the macroscopic examination by counting the number of ulcers on the gastric mucosa were performed.

Data processing

The normality of data distribution was tested and appropriate tests were used: One-way ANOVA for data with normal distribution and Kruskal-Wallis for abnormal data. The limit of the degree of significance was set at $p < 0.05$ with 80 % research power and 95 % confidence interval.

Ethical Consideration

The study was successfully accepted by Gadjah Mada University, Indonesia review board with institutional review board (IRB) decision No 00017/LPPT/VI/2021.

Results

Sample characteristics

The samples consisted of 33 female Wistar rats aged 3-4 months weighed 100-250 g. They were

grouped into MPVK1, MPVK2 and K group, the minimum weight of rats in the MPVK1 group was 170, the maximum weight was 250 while the average was 207. The minimum body weight for rats in the MPVK2 group was 180, the maximum was 250 and the average was 217. At the same time, the minimum body weight for the rats in the control group (K) was 178, the maximum was 250 and the average was 210, as shown in Figure 1.

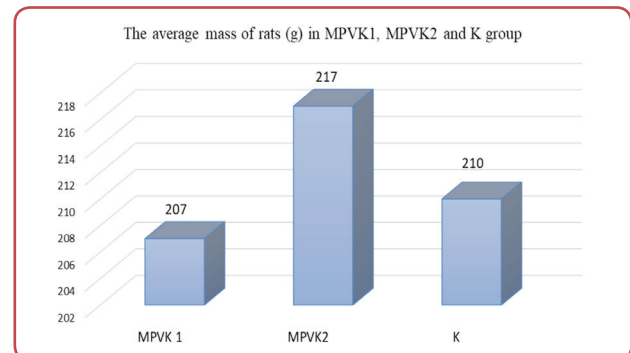


Figure 1: The average weight of Wistar rats in the MPVK 1, MPVK 2 and K group

Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

The minimum level of MDA in the MPVK 1 group was 1.01, while the maximum level was 7.67 and its average was 2.60, respectively. The minimum level of MDA in the MPVK2 group was 1.5, while its maximum level was 2.10 and the average was 1.81. The minimum level of MDA in the control group (K) was 2.19, the maximum level was 3.93 and the average was 3.27. The lowest mean level of MDA was in the MPVK2 group, as shown in Figure 2.

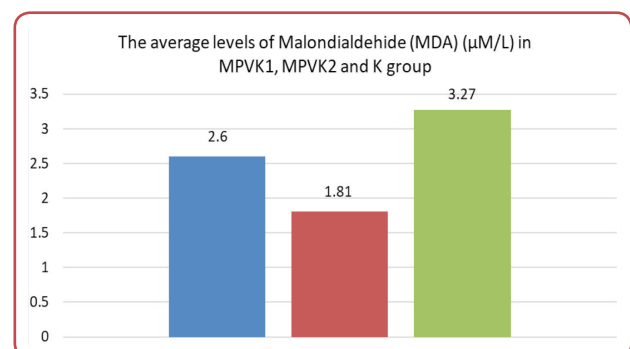


Figure 2: The average levels of malondialdehyde (MDA) of rats in MPVK1, MPVK2 and K group

Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

The average levels of nitric oxide (NO)

The minimum level of NO in the MPVK1 group

was 88, meanwhile the maximum level was 131.28 and the average was 112.73. The minimum level of NO in the MPVK2 group was 221.28, while the maximum level was 353.21 and the average was 286.17. The minimum level of NO in the K group was 101.95, meanwhile, the maximum level was 120.28 and the average was 111.74. The lowest average NO level was in group K, as shown in Figure 3.

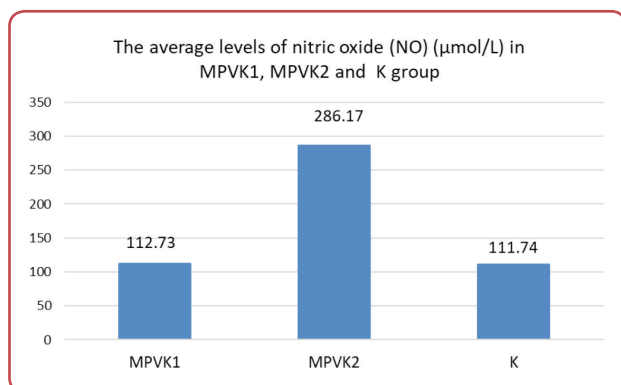


Figure 3: The average levels of nitric oxide (NO) in MPVK 1, MPVK 2 and K group

Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

The average number of ulcers

The minimum number of ulcers in the MPVK1 group was 0, with its maximum was 3 and the average was 0.45. The number of ulcers in the MPVK2 group was 0, a maximum of 1 and its average was 0.04. The number of ulcers in the control group (K) was 0, a maximum of 7 and its average was 3.55. The lowest mean of number ulcers was in the MPVK 2 group, as seen in Figure 4.

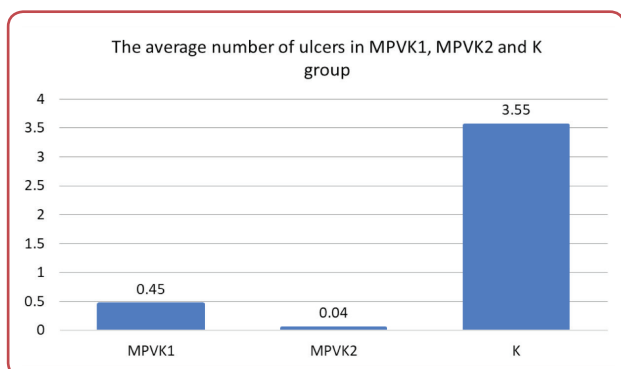


Figure 4: The average number of ulcers in the MPVK1, MPVK2 and K group

Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

A macroscopic picture of gastric mucosal deterioration is shown in Figure 5. The results showed that the highest average number of ulcers was found in the control group.

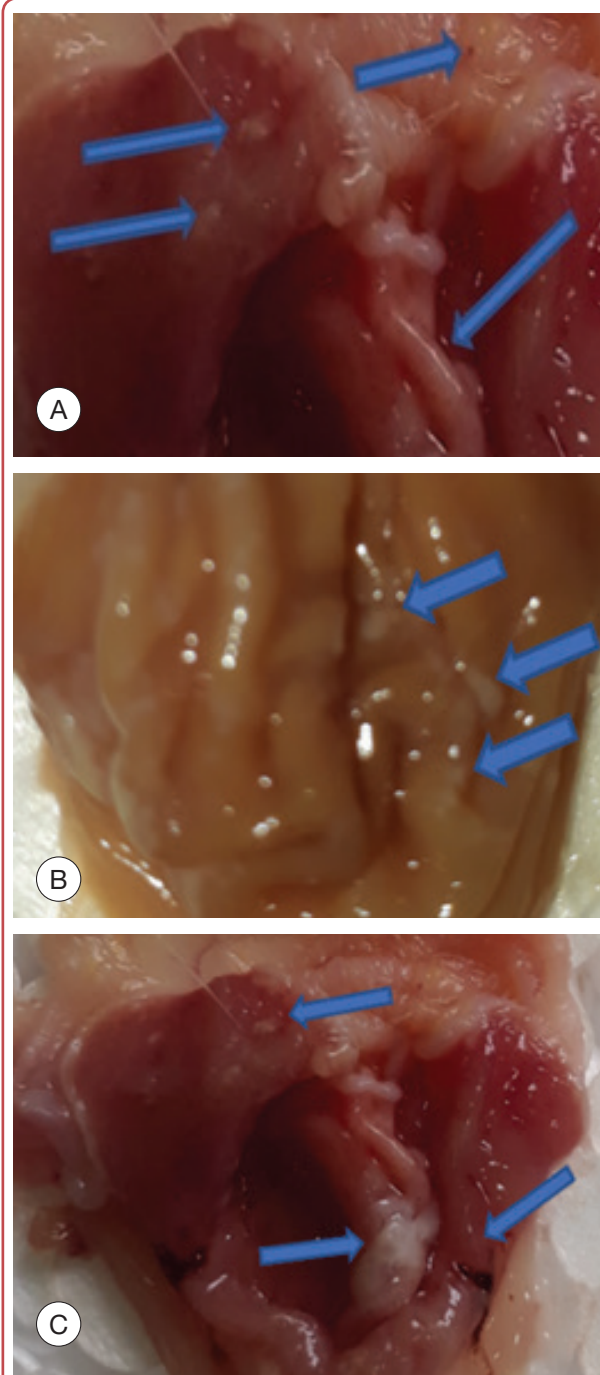


Figure 5 (A, B, C): Gastric macroscopic characteristics observed in the control group. The signs of hypoxaemia, bleeding and gastric mucosal ulcers were displayed by the blue arrows

The control group rats were given 0.3 % NaCMC solvent.

The results of gastric macroscopic examination in treatment of group 1 and 2 are shown in Figure 6.

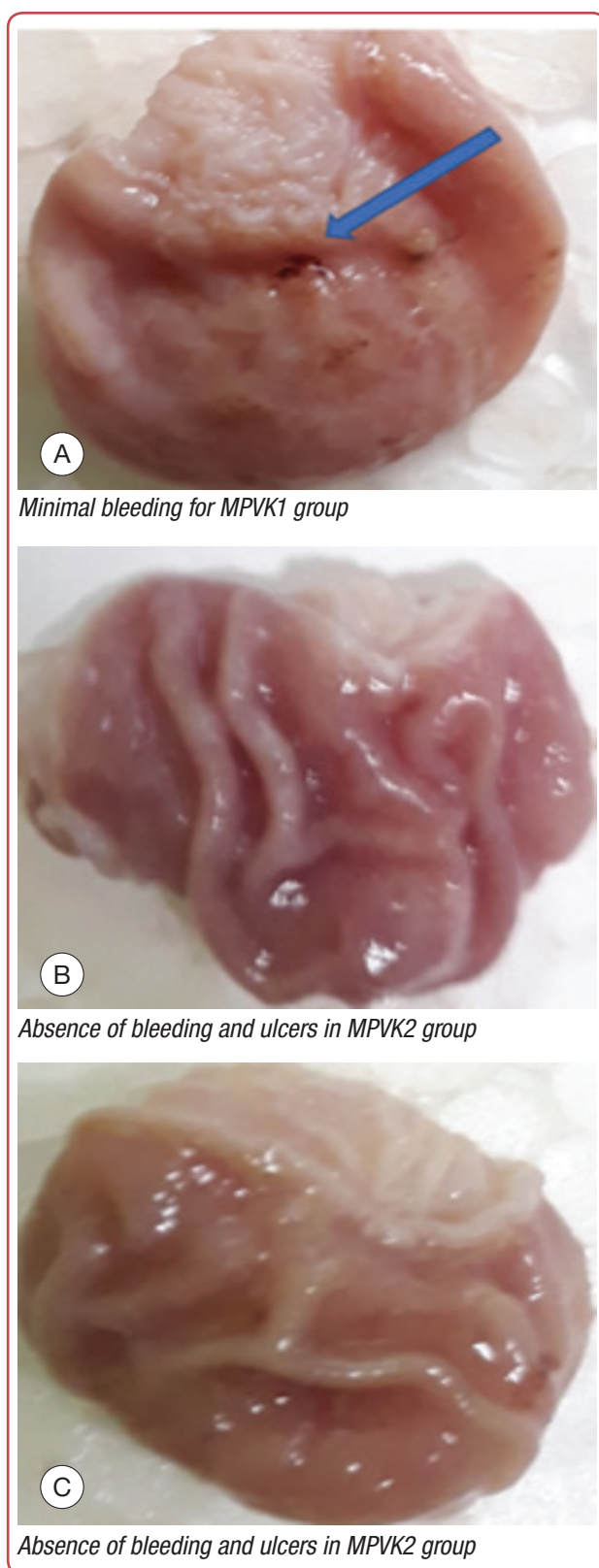


Figure 6: Gastric macroscopic characteristics observed in the treatment groups

A. Macroscopic characteristics of stomach in the MPVK1 group showing minimal bleeding (pointed by the blue arrow), with fewer ulcers than that in the control group; B and C, showing macroscopic characteristics of the stomach in the MPVK2 group which did not perform bleeding and ulcers. Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given the yellow kepok banana peel extract at a dose of 160 mg / 200 g BW with 0.3 % NaCMC solvent.

Table 1 shows the results of the One-Way ANOVA test. It revealed that there were significant distinctions in the average level of MDA and NO and the number of ulcers in the 3 group of experimental rats; MPVK 1, MPVK 2 and K.

Table 1: One-Way ANOVA test results has displayed the average values of MDA and NO, accompanied by the number of ulcers in the experimental groups MPVK1 and MPVK 2 and control group K

Parameter	MPVK1	MPVK2	K	p-value
MDA	2.60	1.81	3.27	0.013
NO	112.73	286.17	111.74	< 0.001
Number of ulcers	0.45	0.04	3.55	< 0.001

MDA: malondialdehyde; NO: nitric oxide; Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

Treatment group 1 (MPVK1) rats were given the yellow kepok banana peel extract at a dose of 80 mg / 200 g BW with 0.3 % NaCMC solvent and treatment group 2 (MPVK2) rats were given 160 mg / 200 g BW yellow kepok banana peel extract with 0.3 % NaCMC solvent. The control group (K) was given 0.3 % NaCMC.

Discussion

An induction of free radicals and oxidative stress causes gastric mucosal deterioration.² Further, an imbalance of aggressive and defensive factors can cause gastric mucosal ulcers. The aggressive factor is more dominant than the defensive factor. The existence of free radicals is part of the aggressive factor.¹ An example of free radicals is NSAIDs. Acetylsalicylic acid works by blocking certain natural substances in the body to reduce pain and swelling. Acetylsalicylic acid is an irritant.⁴⁻⁶ Acetylsalicylic acid causes a defect in the mucosal barrier and back diffusion of H⁺ ions occurs. Histamine is stimulated to secrete more gastric acid, resulting in dilation and increased permeability of capillaries, gastric mucosal damage, acute or chronic gastritis and gastric mucosal ulcers.¹⁶

The experimental animal model of gastric mucosal ulcers in this study was carried out by inducing acetylsalicylic acid. 5% acetylsalicylic acid was given at a dose of 1500 mg/kg BW in all study groups. The results showed that the highest average number of ulcers was found in the control group. The minimum number of gastric ulcers in the control group (K) was 0, the maximum number of ulcers was 7 and the average ulcer was 3.55.

The detrimental effect of free radicals that cause biological damage is oxidative stress. Cells exposed to oxidative stress will activate defence mechanisms to survive.^{7, 17} Lipid peroxidation itself is the result of the performance of free radicals and this parameter is the most accessible to measure.^{2, 11} Lipid peroxidase can damage membrane structures, it leads changes in permeability, inhibits metabolic processes and transform ion transport as well.¹¹ The measurement of lipid peroxidation level is carried out by measuring the final product, one of which is MDA. The accumulation of MDA is an early indicator of the mechanism of cell and tissue deterioration. MDA as final product of the lipid peroxidation process is used as an indicator of cell deterioration in the stomach due to the oxidative stress.⁹⁻¹¹

The results of this study have revealed that the minimum level of MDA in the control group (K) of 2.19 is the highest among those three groups, as well as the maximum and the average level of MDA.

Kepok and Uli banana peel extracts had increased superoxide dismutase (SOD) activity and decreased MDA levels in hypercholesterolemic rats.¹⁸ Kepok banana peel extracts decreased MDA levels in male mice (*mus musculus*) that was exposed to cigarette smoke.¹⁴ Kepok banana peel contains flavonoid and phenolic antioxidants, the antioxidant content of flavonoids and phenolics showed hepatoprotection in acetylsalicylic acid-induced rats.¹⁹ Other antioxidants component in kepok banana peels are flavonoids, tannins, saponins and polyphenols. Tannins are useful in minimising gastric acid secretion and have cytoprotective effect.⁵ Other benefits of tannins are the increasing tissue reconstruction, gastric acid production inhibition and the inhibition of *Helicobacter pylori* activity. Further, saponins also inhibit gastric acid production and minimise gastric fluid pH levels.^{8, 16, 20}

Kepok banana peel extract reduces oxidative stress in peptic ulcers through the antioxidant pathway.¹⁹ Prevention and treatment of gastric mucosal ulcers by exploring natural products is something that is very impressive. The rats that were given kepok banana peel extract performed the increase of protection phenomena against acetylsalicylic acid induction by elevating the antioxidant level of NO. Acetylsalicylic acid converts hydroperoxyl to hydroxy fatty acids which is culminated from lipid peroxidation of cell

destruction. The release of damaging free radicals likely occurs resulting in the death of tissue cells in the stomach. The ulceration effect of superficial epithelial cells on the gastric mucosa constructs the base of gastric ulceration.¹⁴ The kepok banana peel extract exhibits its gastroprotection by increasing mucosal defence factors by increasing the body's antioxidant levels, NO.^{14, 16}

This study revealed that the rats given kepok banana peel extract had increased average level of NO antioxidants compared to the control group, namely MPVK1 (112.73) and MPVK2 (286.17). The lowest average NO level was found in the control group, that is 111.74. There is a significant distinction in the mean of NO in the three groups with $p < 0.001$. The average number of ulcers in the treatment group was lower than in the control group. Hence, the mean number of ulcers in the MPVK1 group was 0.45. The average number of ulcers in the MPVK2 group was 0.04. The mean number of ulcers in control group (K) was 3.55. Meanwhile, the lowest mean number of ulcers was found in MPVK2 group. There was a significant distinction in the mean number of ulcers in the three groups with $p < 0.001$.

Conclusion

There is a significant distinction in the mean value of MDA, NO as well as the distinction of number of ulcers among the three groups. Therefore, kepok banana peel extract has good efficacy in reducing the markers of gastric damage in Wistar rats.

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Conflict of interest

None.

References

1. Tytgat GN. Etiopathogenetic principles and peptic ulcer disease classification. *Dig Dis* 2011;29(5):454-8.
2. Li K, Gesang L. Oxidative stress is associated with the gastric mucosa lesion of the Tibetans with high-altitude polycythemia. *Int J Clin Exp Med* 2019;12(3):2503-10.
3. El-Meligy RM, Awaad AS, Soliman GA, Kenawy SA, Alqasoumi SI. Prophylactic and curative anti-ulcerogenic activity and the possible mechanisms of action of some desert plants. *Saudi Pharm J* 2017 Mar;25(3):387-96.
4. Dionisio KL, Phillips K, Price PS, Grulke CM, Williams A, Biryol D, et al. The Chemical and Products Database, a resource for exposure-relevant data on chemicals in consumer products. *Sci Data* 2018 Jul 10;5:180125. doi: 10.1038/sdata.2018.125.
5. Zulaikhah ST. The role of antioxidant to prevent free radicals in the body. *Sains Med* 2017;8(1):39. doi: 10.30659/sainsmed.v8i1.1012.
6. Flower R. What are all the things that aspirin does? *BMJ* 2003 Sep 13;327(7415):572-3.
7. Sharifi-Rad M, Anil Kumar NV, Zucca P, Varoni EM, Dini L, Panzarini E, et al. Lifestyle, oxidative stress, and antioxidants: back and forth in the pathophysiology of chronic diseases. *Front Physiol* 2020 Jul 2;11:694. doi: 10.3389/fphys.2020.00694.
8. Sharifi-Rad M, Fokou PVT, Sharopov F, Martorell M, Ademiluyi AO, Rajkovic J, et al. Antiulcer agents: From plant extracts to phytochemicals in healing promotion. *Molecules* 2018 Jul 17;23(7):1751. doi: 10.3390/molecules23071751.
9. Ayala A, Muñoz MF, Argüelles S. Lipid peroxidation: production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. *Oxid Med Cell Longev* 2014;2014:360438. doi: 10.1155/2014/360438.
10. Dhama K, Latheef SK, Dadar M, Samad HA, Munjal A, Khandia R, et al. Biomarkers in stress related diseases/disorders: diagnostic, prognostic, and therapeutic values. *Front Mol Biosci* 2019 Oct 18;6:91. doi: 10.3389/fmolb.2019.00091.
11. Domínguez R, Pateiro M, Gagaoua M, Barba FJ, Zhang W, Lorenzo JM. A comprehensive review on lipid oxidation in meat and meat products. *Antioxidants* 2019;8(10):1-31.
12. Tungmunthum D, Thongboonyou A, Pholboon A, Yangsabai A. Flavonoids and other phenolic compounds from medicinal plants for pharmaceutical and medical aspects: an overview. *Medicines (Basel)* 2018 Aug 25;5(3):93. doi: 10.3390/medicines5030093.
13. Soesanto E, Pranata S, Rejeki S, Irham LM. The role of bamboo shoot *Gigantochloa Apus* extract in decreasing the IL-17/IL-10 ratio level in the atherosclerosis process. *Open Access Maced J Med Sci* 2021;9(A):817-21.
14. Amang AP, Mezui C, Siwe GT, Emakoua J, Mbah G, Nkwengoua EZ, et al. Healing and antiseptic effects of aqueous extract of *Eremomastax speciosa* (Acanthaceae) on unhealed gastric ulcers. *Biomed Res Int* 2017;2017:1924320. doi: 10.1155/2017/1924320.
15. Rao USM, Ahmad BA, Mohd KS, Zin T. Antiulcer activity of *Musa paradisiaca* (banana) tepal and skin extracts in ulcer induced albino mice. *Malaysian J Anal Sci* 2016;20(5):1203-16.
16. de Jesus NZT, de Souza Falcão H, Gomes IF, de Almeida Leite TJ, de Moraes Lima GR, Barbosa-Filho JM, et al. Tannins, peptic ulcers and related mechanisms. *Int J Mol Sci* 2012;13(3):3203-28.
17. Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, Arcoraci V, et al. Oxidative stress: harms and benefits for human health. *Oxid Med Cell Longev* 2017;2017:8416763. doi: 10.1155/2017/8416763.
18. Ulfa A, Ekastuti DR, Wresdiyati T. [The Potency of Kepok Banana peel Extract (*Musa paradisiaca* forma typica) and Uli (*Musa paradisiaca sapientum*) in increasing superoxide dismutase activity and reducing malondialdehyde levels in the liver of hypercholesterolemic rats mode]. *Acta Vet Indones* 2020;8(1):40-6. Indonesian.
19. Yulis PAR, Sari Y. [Antioxidant activity of muli banana peel (*Musa acuminata* linn) and kepok banana peel (*Musa paradisiaca* formatypica)]. *Al-Kimia* 2020 Dec;8(2):189-200. Indonesian.
20. Mekonnen AN, Asrade Atnafie S, Wahab Atta MA. Evaluation of antiulcer activity of 80% methanol extract and solvent fractions of the root of *Croton macrostachyus* Hocsht: Ex Del. (Euphorbiaceae) in rodents. *Evid Based Complement Alternat Med* 2020 Apr 9;2020:2809270. doi: 10.1155/2020/2809270.