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1 **Psychochemical Characteristics of Black Rice Powder Based on** 2 **Maltodextrin and Skimmed Milk Powder Ratio as Encapsulant**

7 **ABSTRACT**

9 *Anthocyanins are bioactive components in black rice. Black rice anthocyanins are potent*
10 *antioxidants, so they have the potential to be developed into functional food products. As*
11 *a bioactive component, anthocyanins in extracts have low stability to environmental*
12 *conditions such as light, temperature, and pH. Encapsulation process using the spray*
13 *drying technique is known to protect and increase the stability of anthocyanin bioactive*
14 *compounds. This study's general objective was to determine the effect of the ratio of*
15 *maltodextrin (MDE) and skim milk powder (SMP) as an encapsulant on black rice extract*
16 *powder's physical and chemical characteristics. The research method is an experimental*
17 *type using a monofactor Completely Randomized Design (CRD), which consists of 6*
18 *treatments, namely the MDE:SMP ratio (100:0, 90:10, 80:20, 70:30, 60:40, and 50:50).*
19 *The results showed that there was a very significant effect of the ratio of MDE and SMP*
20 *on the physical and chemical characteristics of black rice powder. MDE and SMP ratios*
21 *of 50:50 resulted in black rice powder's best physical and chemical characteristics.*

Commented [H1]: Please add briefly why MDE and SMP were chosen as treatments

Commented [H2]: Single factor

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23 **Keywords:** *black rice extract, encapsulation, coating material, physicochemical*
24 *characteristics.*

25 **ABSTRAK**

26
27
28 Antosianin merupakan komponen bioaktif pada beras hitam. Antosianin beras hitam
29 bersifat antiosidan kuat sehingga sangat potensial dikembangkan menjadi produk
30 pangan fungsional. Sebagai komponen bioaktif antosianin dalam bentuk ekstrak memiliki
31 stabilitas yang rendah terhadap kondisi lingkungan seperti cahaya, suhu dan pH. Proses
32 enkapsulasi menggunakan Teknik spray drying diketahui mampu melindungi dan
33 meningkatkan stabilitas senyawa bioaktif antosianin. Tujuan umum penelitian yaitu untuk
34 mengetahui pengaruh rasio maltodekstrin (MDE) dan susu bubuk skim (SMP) sebagai
35 enkapsulan terhadap karakteristik fisik dan kimia serbuk ekstrak beras hitam. Metode
36 penelitian berjenis eksperimen menggunakan Rancangan Acak Lengkap (RAL)
37 monofaktor, yang terdiri dari 6 perlakuan yaitu rasio MDE:SMP (100:0, 90:10, 80:20,
38 70:30, 60:40 dan 50:50). Hasil penelitian menunjukkan ada pengaruh yang sangat nyata
39 dari rasio MDE dan SMP terhadap karakteristik fisik dan kimia serbuk beras hitam. Rasio
40 MDE dan SMP 50:50 menghasilkan karakteristik fisik dan kimia serbuk beras hitam
41 terbaik.

42 **Kata Kunci:** ekstrak beras hitam, enkapsulasi, bahan penyalut, karakteristik fisikokimia.
43

1. INTRODUCTION

Black rice is pigmented rice with an endosperm covered with black aleurone. Black rice cultivation in Indonesia is increasing, but its utilization in the food sector is not optimal. However, black rice has anthocyanin bioactive components, potent antioxidants, so it can potentially be developed as an ingredient in functional food products (Hosoda et al., 2018).

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Anthocyanin pigments in black rice can be obtained by extraction using the maceration method. Generally, ethanol solvent is used in the maceration method. Ethanol has advantages over other solvents such as methanol and acetone, namely it is more economical and has low toxicity (Nour et al., 2013). Acidified ethanol has been confirmed to increase yield and anthocyanin content in black rice extract (Pedro et al., 2016). Extraction with heat-assisted maceration technique based on a recent study (Nurhidajah et al., 2022) was reported to be very effective in preparing black rice anthocyanin extract. Black rice anthocyanin extract is volatile to light, temperature and pH (Cassidy, 2018). Encapsulation techniques can overcome this weakness using a freeze dryer, spray dryer, and others (Bao et al., 2019; Fang & Bhandari, 2010; Howard et al., 2013).

Commented [H5]: Are light and pH influence the volatility of a substance? If so.. please add some more references. "susceptible" is more appropriate word instead of using "volatile"

Encapsulation using a spray dryer is an effective solution to protect and increase the stability of anthocyanin bioactive compounds (Kalušević et al., 2017). This technique has been widely used in the food industry. In addition to being applicable, the spray drying (SD) method for thermosensitive products is due to a short contact time with a heat source (Lourenço et al., 2020). The success of SD is influenced by several things, including inlet and outlet temperatures, air flow rate, feed flow rate, speed of the atomizer, to the type

1 and concentration of coating material used (Patil-Gadhe & Pokharkar, 2014). The coating
2 material is one of the factors that will affect the encapsulated powder.

3 Mahdavi et al. (2014) reported maltodextrin (MDE) as the most optimal coating
4 material in anthocyanin encapsulation using the SD technique. MDE is a coating material
5 with various advantages, including high solubility, low viscosity, and maintaining the
6 stability of bioactive compounds (Suhag & Nanda, 2016). Recent research by Kalušević
7 et al. (2017) reported that skimmed milk powder (SMP) coating material produced higher
8 anthocyanin levels than maltodextrin. As a coating material, SMP has non-stick
9 properties and high encapsulation efficiency. Apart from being a source of protein,
10 lactose, vitamins, and minerals, this ingredient provides additional nutritional value
11 (Bylaitė et al., 2001; Jing & Giusti, 2005; Shamaei et al., 2016). However, using SMP to
12 encapsulate anthocyanin extracts resulted in powders with lower phenolic/flavonoid
13 content (Kalušević et al., 2017).

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14 Until now, there has been no research report on the combination of MDE and SMP
15 as a coating material in the encapsulation of black rice extract. Based on this, it is
16 necessary to review further the effect of the MDE-SMP combination as a coating material
17 for black rice extract, the right combination is expected to produce black rice extract
18 powder with the best physical and chemical characteristics.

19
20

2. MATERIALS AND METHODS

21

2.1. Materials

22

23 The research material was black rice of the Jeliteng variety collected from organic rice
24 farmers in the Karanganyar region, Central Java Province, Indonesia, food-grade citric
25 acid from PT Gunacipta Multirasa Indonesia, and aquadest. Pro-analytical chemicals
26

1 from Merck include ethanol, potassium chloride, sodium acetate, folin-ciocalteu, sodium
2 carbonate, gallic acid (C₇H₆O₅), and 2,2-Diphenyl-1-picrylhydrazyl (DPPH).

3 **2.2. Black rice anthocyanin extraction (Nurhidajah et al., 2022)**

4
5 Black rice is ground into flour. 100 g of black rice flour was added with ethanol (56%
6 v/v) and citrate (4.5% w/v) solvent in a ratio of 1:10 (w/v). The extraction process was
7 carried out in a thermostatic water bath with a controlled temperature (50 C) for 120
8 minutes with constant stirring (500 rpm). The following process separates the liquid from
9 the residue using 400 mesh filter paper. The ethanol in the extract was then evaporated
10 using a rotary evaporator at 55 °C. The anthocyanin extract of black rice was stored in a
11 dark glass bottle at 4°C until used.

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Commented [H10]: Black rice powder sample (100 g). don't put number at the beginning of the sentence.

Commented [H11]: How was the separation process (easy to flow or hard?) because 400 mesh size is very dense

Commented [H12]: The final volume or viscosity of anthocyanin extract ?.. it should be the same for all samples

12 **2.3. Anthocyanin extract encapsulation (Nurhidajah et al., 2022)**

13
14 MD and SMP coating materials emulsion solutions with different ratios (100:0, 90:10,
15 80:20, 70:30, 60:40 and 50:50) were prepared at a concentration of 20% (w/v) with
16 deionized water at room temperature. Anthocyanin extract was mixed with each coating
17 material in a ratio of 1:1 (v/v). Each mixture was homogenized separately for 15 min at
18 3000 rpm. It was dried using a laboratory-scale spray dryer with an inlet air temperature
19 of 120 ± 1 C, an outlet air temperature of 80 ± 5 C, and a feed flow rate of 6.0 mL/minute
20 at a pressure of 1.5 bar. The obtained powder was collected and stored at -20°C until
21 analyzed.

Commented [H13]: Emulsion and solution are different in molecular size

Commented [H14]: Coated anthocyanin powder

22 **2.3. Anthocyanin content (Yamuangmorn et al., 2018)**

23
24 A differential pH method was used to determine black rice powder's anthocyanin content.
25 First, 1 g of powder was dissolved with 1 mL of ethanol in a test tube, then mixed
26 separately with two buffer solutions (1 mL of potassium chloride buffer pH 1.0 and 1 mL

Commented [H15]: Is it the original black rice powder or coated anthocyanin powder? D

1 sodium acetate buffer pH 4.5), and incubated at room temperature (± 25 °C) for 15
2 minutes. The absorbance was measured by spectrophotometer at 520 and 700 nm. The
3 absorbance value was obtained by subtracting difference in absorbance at a wavelength
4 of 520 nm and 700 nm at pH 1.0 with the difference in absorbance at pH 4.5. Anthocyanin
5 content was obtained by multiplying the absorbance value by the molecular weight of
6 cyanidin-3-glucoside (448.8 g/mol) and the amount of dilution, then divided by the
7 coefficient of molar absorptivity of cyanidin-3-glucoside (26900 l/mol cm) and the width
8 of the cuvette (1 cm). The anthocyanin content of black rice extract was expressed in
9 mg/100g.

Commented [H16]: Please give the formula for calculation in order readers can understand more easily

10 2.4. Total phenolics

11
12 The Folin-Ciocalteu method (Pedro et al., 2016), with slight modifications, was used to
13 analyze total phenolics content (TPC). 0.5 g of microcapsules were prepared in a dark
14 tube, then 5 mL of Folin-Ciocalteu 10% (v/v) reagent was added. The solution was
15 homogenized for 5 mins and added 4 mL of 7.5% Na_2CO_3 (w/v). The mixture was
16 incubated for 60 mins at room temperature ($25 \pm 1^\circ\text{C}$). Ethanol is used as a blank solution.
17 Gallic acid in ethanol with a concentration of 100-500 ppm is used as a standard solution.
18 Next, the absorbance was measured using a UV-Vis spectrophotometer with a wavelength
19 of 765 nm. TPC of black rice extract was expressed as mg GAE/100g.

Commented [H17]: Please used the standard spelling (Na_2CO_3)

20 2.5. Flavonoid content

21
22 Cai et al. (2016) adopted a slight modification method in determining the flavonoid
23 content. A sample of 0.5 ml black rice extract was prepared in a dark tube, to which 1.5
24 ml of ethanol, 0.1 ml of 10% AlCl_3 , 0.1 ml of 1 M CH_3COOK , and 2.8 ml of distilled
25 water were added. The solution was homogenized and incubated at room temperature (25

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1 ± 1 °C) for 30 minutes. The absorbance of the sample was measured using a UV-Vis
2 spectrophotometer with a wavelength of 415 nm, the blank solution using aquadest. The
3 standard curve uses a quercetin solution in aquadest with a concentration range of 20-100
4 ppm. Therefore, the flavonoid content of black rice extract was expressed as mg QE/100
5 g.

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6 2.6. Antioxidant activities

7
8 Antioxidant activity was determined according to Pedro et al. (2016), with slight
9 modifications. A total of 1.5 ml of 0.2 mM DPPH in ethanol was mixed with 0.2 ml of
10 the sample into a test tube, and ethanol was added to a final volume of 3.5 ml. The tubes
11 were tightly closed, homogenized, and incubated at room temperature (25 ± 1 °C) for 60
12 minutes. The absorbance was measured at a wavelength of 517 nm, and the ability of the
13 extract to scavenge DPPH was obtained by reducing the absorbance of the blank to the
14 sample, then compared with the absorbance of the blank and expressed in % of inhibition.

Commented [H22]: please write the formula for calculation ..
same as above comment.
, also why the IC 50 was not determined?

15 2.7. pH value

16
17 The pH value of black rice powder was measured with the instrument Hanna HI 2211
18 bench pH meters (Hanna instruments Ltd.), calibration solution using standard buffer.

Commented [H23]: please write the sample preparation for pH
measurement

19 2.8. Yield, moisture content and water activity (a_w)

20
21 Yield was obtained from the final weight percentage of black rice powder (g) with the
22 total solids of the sample (g). Determination of the moisture content of the powder using
23 a moisture analyzer (Shimadzu MOC63u, Japan), while the a_w of the powder was
24 determined using a water activity analyzer (Rotronic Hygropalm-HP23-Aw-A,
25 Switzerland) at 25 C.

2.9. Color measurement

Color characteristics of black rice powder were measured using a calibrated Minolta CR-310 Chromameter (Konica Minolta Business Solution Asia Pte Ltd). The hue angle, H° [$\tan^{-1}(b^*/a^*)$] and chroma, C° [$(a^{*2} + b^{*2})^{1/2}$] were also determined. The H° is used to identify colors (red, yellow, green, and blue), whereas C° distinguishes between bright and dull colors (Caparino et al., 2012).

2.10. Statistical analysis

All experiments were carried out in 4 repetitions. Results are presented in terms of mean \pm standard deviation. The effect of treatment factors was analyzed using one-way ANOVA, and Duncan's post hoc test was used to detect differences between treatments. The significance level used is $p < 0.05$. Statistical analysis was carried out with the help of SPSS 22.0 software.

3. RESULTS AND DISCUSSION

3.1. Anthocyanin and pH value of capsules

The mean anthocyanin content of black rice extract powder obtained was 51.22-84.59 mg/100g (Figure 1a). This amount is higher than the research results reported by Pramitasari and Angelica (2020), which only ranged from 2.00–12.00 mg/100g. The highest anthocyanin levels were obtained at a coating ratio of 50:50 (84.59 mg/100g). There was an increase in anthocyanin levels of 65% (51.22mg/100g) compared to a ratio of 100:0. SMP generally contains β -casein components and has been reported to interact with Cyanidin-3-glucoside compounds from black soybean extract through reversible hydrogen bonds supported by an optimum pH (Kalušević et al., 2017). Previous studies also reported that casein protein interacts with malvidin-3-glucoside from grape extract

Commented [H24]: coated powder?

1 through complex hydrogen bonds, which then form a hydrophobic reaction and
2 effectively increase thermal stability so that the rate of anthocyanin degradation during
3 drying can be minimized (He et al. 2016). The pH value of black rice powder ranged from
4 2.60-2.95 (Figure 1b). The pH value of the black rice extract powder increased with the
5 addition of SMP in the coating material ratio because SMP had a neutral pH value.
6 However, the increase in pH value is still in the anthocyanin stability range, at a value of
7 2-3 (Sipahli et al., 2017). Anthocyanins are more stable under acidic conditions in the
8 form of a flavylum cation structure (Khoo et al., 2017).

9 **3.2. Total phenolics, flavonoid, and antioxidant activities**

10
11 Total phenolic and flavonoid content of black rice powder is presented in Table 1. From
12 the data, it is shown that the total phenolic and flavonoid content increases with the
13 addition of SMP in the ratio of the coating material. B-casein in protein-derived
14 encapsulation forms a more substantial complex with polyphenols, and this is due to the
15 more hydrophobic nature of B-casein. (Hasni et al., 2011). The increased flavonoids are
16 associated with the structure and bonds formed from proteins with flavonoid compounds.
17 This result has previously been described by Bohin et al. (2012) on the flavonoid
18 encapsulation of tea drinks. This underlies the increase in flavonoids when the ratio of
19 SMP to coating material increases. In a previous study, Kalušević et al. (2017) reported
20 the results of high antioxidant activity in coating black rice extract using SMP. Therefore,
21 the antioxidant activity of black rice powder is thought to be derived from the
22 encapsulated anthocyanin, phenolic, and flavonoid components. In addition, the increase
23 in antioxidant activity is also potential from the **maillard** reaction, which produces
24 melanoidin compounds with antioxidant activity. This reaction occurs in the protein

Commented [H25]: Maillard

1 component of SMP due to the high temperature during the drying process (Wang et al.,
2 2011).

3 3.3. Aw, mouisture content, and yield of microcapsules

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4 All samples showed an Aw value lower than 0.6 (0.49-0.58), so the black rice extract
5 powder in this study had an excellent aw value (Goyal et al., 2015). A low aw value tends
6 to result in extended shelf life. The water content values ranged from 0.91 to 1.13% (Table
7 2). The product moisture content was low to prevent particle agglomeration, avoid the
8 caking process, and improve powder dispersion and flowability (Daza et al., 2016).
9 Therefore, the powder from spray drying obtained both the aw level and the moisture
10 content in this study met the requirements and was suitable for inclusion in the dry food
11 matrix.
12

13 The highest yield was obtained from powders with a 50:50 coating material, which
14 is in line with the results reported in a study comparing milk protein-based microparticles
15 with other carriers (Belščak-Cvitanovic et al., 2015). A combination of MDE and SMP
16 coating materials resulted in good physical characteristics so that the coating material can
17 be proposed as a coating material for the bioactive component of black rice extract.

18 3.4. Color characteristic

19 Color is one of the indicators in assessing the quality of food products. During the
20 production process, the color of black rice extract is expected to be maintained until a
21 powder is obtained from drying. Therefore, observing the color measurement of black
22 rice extract powder after the encapsulation process is necessary. The results of chromatic
23 color measurements are shown in Table 3. Based on the parameters a*, b*, and hue angle,
24 it can be concluded that the color of the MD-SMP-based powder is red-violet with varying
25

1 brightness levels. Among all SBHs, the coating with a 50:50 ratio had the lowest
2 brightness value (40.58 ± 0.83) and the highest values for a^* and C^* , indicating that this
3 ratio produced the darkest powder with the highest portion of red color. Many reports
4 have investigated the molecular level of the bond formed between casein-based coating
5 materials and black rice anthocyanins on their stability from a thermodynamic point of
6 view. The casein-anthocyanin bond is mainly stabilized through hydrophobic interactions
7 and hydrogen bonds involving the aglycone and glucosyl moieties of the encapsulated
8 molecule so that the color can be maintained during the drying process (Aprodu et al.,
9 2019).

10 4. CONCLUSIONS

11 ~~Based on the results of the research that has been carried out~~, it can be concluded that the
12 best combination of coating materials in black rice encapsulation was obtained at MDE:
13 SMP ratio of 50:50 with total anthocyanin levels obtained 84.59 mg/100g, with total
14 phenol 0.41 mg GAE/g, total flavonoid 0.21 mg QE/g and antioxidant activity 31.71%
15 RSA. Has an average value of L^* 40.5; a^* 43.64; b^* 12.75; Chroma 45.47; and Hue 16.31
16 with red-purple color types. Anthocyanin stability with a pH value of 2.95. Furthermore,
17 it has an aw of 0.49%, moisture content of 0.91%, and a yield of 51.29%. ~~The interaction~~
18 ~~between the combination of MDE and SMP~~ as a coating material in the encapsulation
19 process of black rice extract had a very significant effect on total anthocyanin levels,
20 phenolics, total flavonoids, antioxidant activity, pH, color, aw, water content, and yield.
21

Commented [H28]: please not to use 'interaction' because this experiment was designed as a single factor experiment.

5. ACKNOWLEDGMENTS

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6. REFERENCES/ DAFTAR PUSTAKA

- Aprodu I, Milea SA, Anghel RM, Enachi E, Barbu V, Crăciunescu O, Râpeanu G, Bahrim GE, Oancea A, Stănciuc N. 2019. New Functional Ingredients Based on Microencapsulation of Aqueous Anthocyanin-Rich Extracts Derived from Black Rice (*Oryza sativa* L.). *Molecules*, 24(18): 11-14. <https://doi.org/10.3390/molecules24183389>
- Bao C, Jiang P, Chai J, Jiang Y, Li D, Bao W, Liu B, Liu B, Norde W, Li Y. 2019. The delivery of sensitive food bioactive ingredients: Absorption mechanisms, influencing factors, encapsulation techniques and evaluation models. *Food Research International*, 120: 130–140. <https://doi.org/10.1016/j.foodres.2019.02.024>
- Belščak-Cvitanović A, Lević S, Kalušević A, Špoljarić I, Đorđević V, Komes D, Nedović V. 2015. Efficiency Assessment of Natural Biopolymers as Encapsulants of Green Tea (*Camellia sinensis* L.) Bioactive Compounds by Spray Drying. *Food and Bioprocess Technology*, 8(12): 2444–2460. <https://doi.org/10.1007/s11947-015-1592-y>
- Bohin MC, Vincken JP, van der Hijden HTWM, Gruppen H. 2012. Efficacy of food proteins as carriers for flavonoids. *J. Agric. Food Chem*, 60: 4136–4143. <https://doi.org/10.1021/jf205292r>
- Bylaitė E, Venskutonis PR, Maždpierienė R. 2001. Properties of caraway (*Carum carvi* L.) essential oil encapsulated into milk protein-based matrices. *European food research and technology*, 212(6): 661–670. <https://doi.org/10.1007/s002170100297>
- Cai Z, Qu Z, Lan Y, Zhao S, Ma X, Wan Q, Jing P, Li P. 2016. Conventional, ultrasound-assisted, and accelerated-solvent extractions of anthocyanins from purple sweet potatoes. *Food Chemistry*, 197: 266–272. <https://doi.org/10.1016/j.foodchem.2015.10.110>
- Caparino OA, Tang J, Nindo CI, Sablani SS, Powers JR, Fellman JK. 2012. Effect of drying methods on the physical properties and microstructures of mango (*Philippine 'Carabao' var.*) powder. *Journal of Food Engineering*, 111(1), 135-148. <https://doi.org/10.1016/j.jfoodeng.2012.01.010>
- Cassidy A. 2018. Berry anthocyanin intake and cardiovascular health. *Molecular Aspects of Medicine*, 61: 76–82. <https://doi.org/10.1016/j.mam.2017.05.002>

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- 1
2 Daza LD, Fujita A, Fávaro-Trindade CS, Rodrigues-Ract JN, Granato D, Genovese MI.
3 2016. Effect of spray conditions on the physical properties of cagaita (*Eugenia*
4 *dysenterica DC.*) fruit extracts. Food and Bioproducts Processing, 97: 20-29.
5 <https://doi.org/10.1016/j.fbp.2015.10.001>
6
- 7 Fang Z, Bhandari B. 2010. Encapsulation of polyphenols - A review. Trends in Food
8 Science and Technology, 21(10): 510–523.
9 <https://doi.org/10.1016/j.tifs.2010.08.003>
10
- 11 Goyal A, Sharma V, Sihag MK, Komar SK, Arora S, Sabikhi L, Singh AK. 2015.
12 Development and physico-chemical characterization of microencapsulated flaxseed
13 oil powder: A functional ingredient for omega-3 fortification. Powder Technology,
14 286: 527-537. <https://doi.org/10.1016/j.powtec.2015.08.050>
15
- 16 Hasni I, Bourassa P, Hamdani S, Samson G, Carpentier R, Tajmir-Riahi HA. 2011.
17 Interaction of milk α - and β -caseins with tea polyphenols. Food chemistry, 126(2):
18 630-639. <https://doi.org/10.1016/j.foodchem.2010.11.087>
19
- 20 He Z, Xu M, Zeng M, Qin F, Chen J. 2016. Interactions of milk α -and β -casein with
21 malvidin-3-O-glucoside and their effects on the stability of grape skin anthocyanin
22 extracts. Food chemistry, 199: 314-322.
23 <https://doi.org/10.1016/j.foodchem.2015.12.035>
24
- 25 Hosoda K, Sasahara H, Matsushita K, Tamura Y, Miyaji M, Matsuyama H. 2018.
26 Anthocyanin and proanthocyanidin contents, antioxidant activity, and in situ
27 degradability of black and red rice grains. Asian-Australasian Journal of Animal
28 Sciences, 31(8): 1213–1220. <https://doi.org/10.5713/ajas.17.0655>
29
- 30 Howard LR, Brownmiller C, Prior RL, Mauromoustakos A. 2013. Improved stability of
31 chokeberry juice anthocyanins by β -cyclodextrin addition and refrigeration. Journal
32 Agric Food Chem, 61(3): 693-9. <https://doi.org/10.1021/jf3038314>.
33
- 34 Jing P, Giusti MM. 2005. Characterization of anthocyanin-rich waste from purple corn cobs
35 (*Zea mays L.*) and its application to color milk. Journal of agricultural and food
36 chemistry, 53(22): 8775–8781. <https://doi.org/10.1021/jf051247o>
37
- 38 Kalušević AM, Lević SM, Čalija BR, Milić JR, Pavlović VB, Bugarski BM, Nedović VA.
39 2017. Effects of different carrier materials on physicochemical properties of
40 microencapsulated grape skin extract. Journal of Food Science and Technology,
41 54(11): 3411–3420. <https://doi.org/10.1007/s13197-017-2790-6>
42
- 43 Khoo HE, Azlan A, Tang ST, Lim SM. 2017. Anthocyanidins and Anthocyanins: Colored
44 Pigments as Food, Pharmaceutical Ingredients, and The Potential Health Benefits.
45 Food & Nutrition Research, 61: 1 – 21.
46 <https://doi.org/10.1080/16546628.2017.1361779>
47

- 1 Lourenço SC, Moldão-Martins M, Alves VD. 2020. Microencapsulation of Pineapple Peel
2 Extract by Spray Drying Using Maltodextrin, Inulin, and Arabic Gum as Wall
3 Matrices. *Foods*, 9(6): 718. <https://doi.org/10.3390/foods9060718>
4
- 5 Mahdavi SA, Jafari SM, Ghorbani M, Assadpoor E. 2014. Spray-Drying
6 Microencapsulation of Anthocyanins by Natural Biopolymers: A Review. *Drying*
7 *Technology*, 32(5): 509–518. <https://doi.org/10.1080/07373937.2013.839562>
8
- 9 Nour V, Stampar F, Veberic R, Jakopic J. 2013. Anthocyanins profile, total phenolics and
10 antioxidant activity of black currant ethanolic extracts as influenced by genotype and
11 ethanol concentration. *Food Chemistry*, 141(2): 961–966.
12 <https://doi.org/10.1016/j.foodchem.2013.03.105>
13
- 14 Nurhidajah, Rosidi A, Yonata D, and Pranata B. 2022. Optimizing extraction of functional
15 compounds from Indonesian black rice using response surface methodology. *Food*
16 *Research*, 6(4): 83-91. [https://doi.org/10.26656/fr.2017.6\(4\).732](https://doi.org/10.26656/fr.2017.6(4).732)
17
- 18 Patil-Gadhe A, Pokharkar V. 2014. Single step spray drying method to develop
19 proliposomes for inhalation: A systematic study based on quality by design approach.
20 *Pulmonary Pharmacology and Therapeutics*, 27(2): 197–207.
21 <https://doi.org/10.1016/j.pupt.2013.07.006>
22
- 23 Pedro AC, Granato D, Rosso ND. 2016. Extraction of anthocyanins and polyphenols from
24 black rice (*Oryza sativa L.*) by modeling and assessing their reversibility and
25 stability. *Food Chemistry*, 191: 12–20.
26 <https://doi.org/10.1016/j.foodchem.2015.02.045>
27
- 28 Pramitasari R, Angelica N. 2020. Ekstraksi, Pengeringan Semprot, dan Analisis Sifat
29 Fisikokimia Antosianin Beras Hitam (*Oryza sativa L.*). *Jurnal Aplikasi Teknologi*
30 *Pangan*, 9(2): 83–94. <https://doi.org/10.17728/jatp.5889>
31
- 32 Shamaei S, Seiiedlou SS, Aghbashlo M, Tsotsas E, Kharaghani A. 2017.
33 Microencapsulation of walnut oil by spray drying: Effects of wall material and drying
34 conditions on physicochemical properties of microcapsules. *Innovative food science*
35 *& emerging technologies*, 39: 101-112. <https://doi.org/10.1016/j.ifset.2016.11.011>
36
- 37 Sipahli S, Mohanlall V, Mellem JJ. 2017. Stability and degradation kinetics of crude
38 anthocyanin extracts from *H. sabdariffa*. *Food Science and Technology*, 37(2): 209–
39 215. <https://doi.org/10.1590/1678-457X.14216>
40
- 41 Suhag Y, Nanda V. 2016. Optimization for spray drying process parameters of nutritionally
42 rich honey powder using response surface methodology. *Cogent Food and*
43 *Agriculture*, 2(1): 1–12. <https://doi.org/10.1080/23311932.2016.1176631>
44
- 45 Wang HY, Qian H, Yao WE, 2011. Melanoidins produced by the Maillard reaction:
46 reaction: structure and biological activity. *Food chemistry*, 128(3): 573-584.
47 <https://doi.org/10.1016/j.foodchem.2011.03.075>
48

- 1 Yamuangmorn S, Dell B, Prom-u-thai C. 2018. Effects of cooking on anthocyanin
- 2 concentration and bioactive antioxidant capacity in glutinous and non-glutinous
- 3 purple rice. *Rice Science*, 25(5): 270–278. <https://doi.org/10.1016/j.rsci.2018.04.004>
- 4
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- 7

1 Table 1. Total phenolic, total flavonoid and antioxidant activity of black rice extract
2 powder based on MD and SMP ratio

MD:SMP Ratio	Total Phenolic (mg GAE/g)	Total Flavonoid (mg QE/g)	Antioxidant Activity (%)
100:0	0.27 ± 0.01 ^a	0.14 ± 0.02 ^a	25.07 ± 0.39 ^a
90:10	0.30 ± 0.01 ^b	0.16 ± 0.01 ^{ab}	26.52 ± 0.35 ^b
80:20	0.33 ± 0.01 ^c	0.17 ± 0.01 ^{bc}	29.23 ± 0.22 ^c
70:30	0.37 ± 0.01 ^d	0.18 ± 0.03 ^{bcd}	30.37 ± 0.38 ^d
60:40	0.38 ± 0.01 ^d	0.19 ± 0.02 ^{cd}	31.18 ± 0.48 ^e
50:50	0.41 ± 0.05 ^e	0.21 ± 0.02 ^d	31.71 ± 0.94 ^e

3 Information:

- 4 1. All values are mean ± standard deviation 4 times repetition
5 2. Different superscript values showed significant differences with a 95% confidence level
6 based on the ANOVA difference test and the LSD post hoc test.
7
8
9

10 Table 2. Physical properties of black rice powder based on MD and SMP ratio

MD:SMP Ratio	Yield (%)	Moisture content (%)	a _w (%)
100:0	33.51 ± 0.82 ^a	1.13 ± 0.03 ^e	0.58 ± 0.58 ^c
90:10	35.11 ± 0.75 ^a	1.08 ± 0.01 ^{de}	0.55 ± 0.55 ^{bc}
80:20	31.14 ± 0.68 ^a	1.04 ± 0.09 ^{cd}	0.54 ± 0.54 ^{bc}
70:30	34.06 ± 0.82 ^a	0.99 ± 0.01 ^{bc}	0.52 ± 0.52 ^{ab}
60:40	47.64 ± 0.73 ^b	0.96 ± 0.02 ^{cb}	0.51 ± 0.51 ^{ab}
50:50	51.29 ± 0.49 ^b	0.91 ± 0.05 ^a	0.49 ± 0.49 ^a

11 Information:

- 12 1. All values are mean ± standard deviation 4 times repetition
13 2. Different superscript values showed significant differences with a 95% confidence level
14 based on the ANOVA difference test and the LSD post hoc test.
15
16

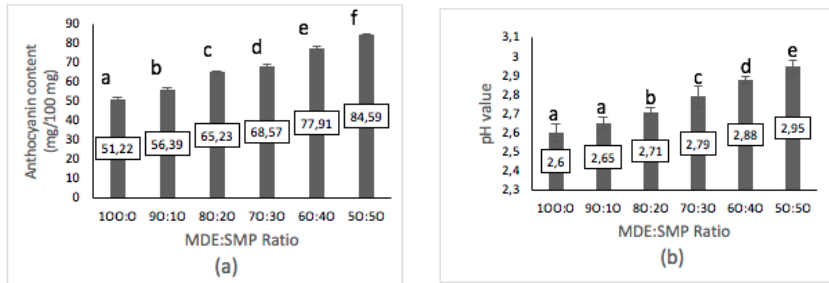
17 Table 3. Color parameters of black rice powder based on MD and SMP ratio

MD:SMP Ratio	L*	a*	b*	C*	H°	Color
100:0	56.06 ± 0.79 ^d	26.83 ± 0.66 ^a	4.43 ± 0.31 ^a	27.20 ± 0.62 ^a	9.39 ± 0.81 ^a	Red-violet
90:10	55.46 ± 1.46 ^c	28.48 ± 0.12 ^b	4.30 ± 0.51 ^a	28.80 ± 0.16 ^b	8.58 ± 0.99 ^a	Red-violet
80:20	52.91 ± 0.61 ^c	28.71 ± 0.39 ^b	5.34 ± 0.50 ^{ab}	29.20 ± 0.35 ^b	10.55 ± 1.06 ^a	Red-violet
70:30	49.07 ± 0.82 ^b	35.78 ± 1.45 ^c	6.36 ± 1.56 ^b	36.36 ± 1.65 ^c	10.03 ± 2.13 ^a	Red-violet
60:40	48.06 ± 0.54 ^a	39.90 ± 0.47 ^d	9.71 ± 0.39 ^c	41.06 ± 0.39 ^d	13.68 ± 0.65 ^b	Red-violet
50:50	40.58 ± 0.83 ^a	43.64 ± 1.43 ^e	12.75 ± 0.83 ^d	45.47 ± 1.16 ^e	16.31 ± 1.50 ^c	Red-violet

18 Information:

- 19 1. All values are mean ± standard deviation 4 times repetition
20 2. Different superscript values showed significant differences with a 95% confidence level
21 based on the ANOVA difference test and the LSD post hoc test.
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Figure 1. Anthocyanin and pH value black rice powder based on MDE:SMP Ratio

1 **Psychochemical Characteristics of Black Rice Powder Based on** 2 **Maltodextrin and Skimmed Milk Powder Ratio as Encapsulant**

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7 **ABSTRACT**

9 *Anthocyanins are bioactive components in black rice. Black rice anthocyanins are potent*
10 *antioxidants, so they have the potential to be developed into functional food products. As*
11 *a bioactive component, anthocyanins in extracts have low stability to environmental*
12 *conditions such as light, temperature, and pH. Encapsulation process using the spray*
13 *drying technique is known to protect and increase the stability of anthocyanin bioactive*
14 *compounds. This study's general objective was to determine the effect of the ratio of*
15 *maltodextrin (MDE) and skim milk powder (SMP) as an encapsulant on black rice extract*
16 *powder's physical and chemical characteristics. The research method is an experimental*
17 *type using a monofactor Completely Randomized Design (CRD), which consists of 6*
18 *treatments, namely the MDE:SMP ratio (100:0, 90:10, 80:20, 70:30, 60:40, and 50:50).*
19 *The results showed that there was a very significant effect of the ratio of MDE and SMP*
20 *on the physical and chemical characteristics of black rice powder. MDE and SMP ratios*
21 *of 50:50 resulted in black rice powder's best physical and chemical characteristics.*

23 **Keywords:** *black rice extract, encapsulation, coating material, physicochemical*
24 *characteristics.*

25 **ABSTRAK**

27
28 *Antosianin merupakan komponen bioaktif pada beras hitam. Antosianin beras hitam*
29 *bersifat antiosidan kuat sehingga sangat potensial dikembangkan menjadi produk*
30 *pangan fungsional. Sebagai komponen bioaktif antosianin dalam bentuk ekstrak memiliki*
31 *stabilitas yang rendah terhadap kondisi lingkungan seperti cahaya, suhu dan pH. Proses*
32 *enkapsulasi menggunakan Teknik spray drying diketahui mampu melindungi dan*
33 *meningkatkan stabilitas senyawa bioaktif antosianin. Tujuan umum penelitian yaitu untuk*
34 *mengetahui pengaruh rasio maltodekstrin (MDE) dan susu bubuk skim (SMP) sebagai*
35 *enkapsulan terhadap karakteristik fisik dan kimia serbuk ekstrak beras hitam. Metode*
36 *penelitian berjenis eksperimen menggunakan Rancangan Acak Lengkap (RAL)*
37 *monofaktor, yang terdiri dari 6 perlakuan yaitu rasio MDE:SMP (100:0, 90:10, 80:20,*
38 *70:30, 60:40 dan 50:50). Hasil penelitian menunjukkan ada pengaruh yang sangat nyata*
39 *dari rasio MDE dan SMP terhadap karakteristik fisik dan kimia serbuk beras hitam. Rasio*
40 *MDE dan SMP 50:50 menghasilkan karakteristik fisik dan kimia serbuk beras hitam*
41 *terbaik.*

42 **Kata Kunci:** *ekstrak beras hitam, enkapsulasi, bahan penyalut, karakteristik fisikokimia.*
43

1. INTRODUCTION

Black rice is pigmented rice with an endosperm covered with black aleurone. Black rice cultivation in Indonesia is increasing, but its utilization in the food sector is not optimal. However, black rice has anthocyanin bioactive components, potent antioxidants, so it can potentially be developed as an ingredient in functional food products (Hosoda et al., 2018).

Anthocyanin pigments in black rice can be obtained by extraction using the maceration method. Generally, ethanol solvent is used in the maceration method. Ethanol has advantages over other solvents such as methanol and acetone, namely it is more economical and has low toxicity (Nour et al., 2013). Acidified ethanol has been confirmed to increase yield and anthocyanin content in black rice extract (Pedro et al., 2016). Extraction with heat-assisted maceration technique based on a recent study (Nurhidajah et al., 2022) was reported to be very effective in preparing black rice anthocyanin extract.

Black rice anthocyanin extract is volatile to light, temperature and pH (Cassidy, 2018). Encapsulation techniques can overcome this weakness using a freeze dryer, spray dryer, and others (Bao et al., 2019; Fang & Bhandari, 2010; Howard et al., 2013).

Encapsulation using a spray dryer is an effective solution to protect and increase the stability of anthocyanin bioactive compounds (Kalušević et al., 2017). This technique has been widely used in the food industry. In addition to being applicable, the spray drying (SD) method for thermosensitive products is due to a short contact time with a heat source (Lourenço et al., 2020). The success of SD is influenced by several things, including inlet and outlet temperatures, air flow rate, feed flow rate, speed of the atomizer, to the type

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so the sentence is one of the weaknesses of black rice anthocyanin extract which is unstable and volatile to light, temperature and pH (Cassidy, 2018)

1 and concentration of coating material used (Patil-Gadhe & Pokharkar, 2014). The coating
2 material is one of the factors that will affect the encapsulated powder.

3 Mahdavi et al. (2014) reported maltodextrin (MDE) as the most optimal coating
4 material in anthocyanin encapsulation using the SD technique. MDE is a coating material
5 with various advantages, including high solubility, low viscosity, and maintaining the
6 stability of bioactive compounds (Suhag & Nanda, 2016). Recent research by Kalušević
7 et al. (2017) reported that skimmed milk powder (SMP) coating material produced higher
8 anthocyanin levels than maltodextrin. As a coating material, SMP has non-stick
9 properties and high encapsulation efficiency. Apart from being a source of protein,
10 lactose, vitamins, and minerals, this ingredient provides additional nutritional value
11 (Bylaitė et al., 2001; Jing & Giusti, 2005; Shamaei et al., 2016). However, using SMP to
12 encapsulate anthocyanin extracts resulted in powders with lower phenolic/flavonoid
13 content (Kalušević et al., 2017).

14 Until now, there has been no research report on the combination of MDE and SMP
15 as a coating material in the encapsulation of black rice extract. Based on this, it is
16 necessary to review further the effect of the MDE-SMP combination as a coating material
17 for black rice extract, the right combination is expected to produce black rice extract
18 powder with the best physical and chemical characteristics.

19 20 **2. MATERIALS AND METHODS**

21 **2.1. Materials**

22 The research material was black rice of the Jeliteng variety collected from organic rice
23 farmers in the Karanganyar region, Central Java Province, Indonesia, food-grade citric
24 acid from PT Gunacipta Multirasa Indonesia, and aquadest. Pro-analytical chemicals
25
26

1 from Merck include ethanol, potassium chloride, sodium acetate, folin-ciocalteu, sodium
2 carbonate, gallic acid (C₇H₆O₅), and 2,2-Diphenyl-1-picrylhydrazyl (DPPH).

3 **2.2. Black rice anthocyanin extraction (Nurhidajah et al., 2022)**

4
5 Black rice is ground into flour. 100 g of black rice flour was added with ethanol (56%
6 v/v) and citrate (4.5% w/v) solvent in a ratio of 1:10 (w/v). The extraction process was
7 carried out in a thermostatic water bath with a controlled temperature (50 C) for 120
8 minutes with constant stirring (500 rpm). The following process separates the liquid from
9 the residue using 400 mesh filter paper. The ethanol in the extract was then evaporated
10 using a rotary evaporator at 55 °C. The anthocyanin extract of black rice was stored in a
11 dark glass bottle at 4°C until used.

12 **2.3. Anthocyanin extract encapsulation (Nurhidajah et al., 2022)**

13
14 MD and SMP coating materials emulsion solutions with different ratios (100:0, 90:10,
15 80:20, 70:30, 60:40 and 50:50) were prepared at a concentration of 20% (w/v) with
16 deionized water at room temperature. Anthocyanin extract was mixed with each coating
17 material in a ratio of 1:1 (v/v). Each mixture was homogenized separately for 15 min at
18 3000 rpm. It was dried using a laboratory-scale spray dryer with an inlet air temperature
19 of 120 ± 1 C, an outlet air temperature of 80 ± 5 C, and a feed flow rate of 6.0 mL/minute
20 at a pressure of 1.5 bar. The obtained powder was collected and stored at -20°C until
21 analyzed.

22 **2.3. Anthocyanin content (Yamuangmorn et al., 2018)**

23
24 A differential pH method was used to determine black rice powder's anthocyanin content.
25 First, 1 g of powder was dissolved with 1 mL of ethanol in a test tube, then mixed
26 separately with two buffer solutions (1 mL of potassium chloride buffer pH 1.0 and 1 mL

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1 sodium acetate buffer pH 4.5), and incubated at room temperature (± 25 °C) for 15
2 minutes. The absorbance was measured by spectrophotometer at 520 and 700 nm. The
3 absorbance value was obtained by subtracting difference in absorbance at a wavelength
4 of 520 nm and 700 nm at pH 1.0 with the difference in absorbance at pH 4.5. Anthocyanin
5 content was obtained by multiplying the absorbance value by the molecular weight of
6 cyanidin-3-glucoside (448.8 g/mol) and the amount of dilution, then divided by the
7 coefficient of molar absorptivity of cyanidin-3-glucoside (26900 l/mol cm) and the width
8 of the cuvette (1 cm). The anthocyanin content of black rice extract was expressed in
9 mg/100g.

10 **2.4. Total phenolics**

11
12 The Folin-Ciocalteu method (Pedro et al., 2016), with slight modifications, was used to
13 analyze total phenolics content (TPC). 0.5 g of microcapsules were prepared in a dark
14 tube, then 5 mL of Folin-Ciocalteu 10% (v/v) reagent was added. The solution was
15 homogenized for 5 mins and added 4 mL of 7.5% Na₂CO₃ (w/v). The mixture was
16 incubated for 60 mins at room temperature (25 ± 1 °C). Ethanol is used as a blank solution.
17 Gallic acid in ethanol with a concentration of 100-500 ppm is used as a standard solution.
18 Next, the absorbance was measured using a UV-Vis spectrophotometer with a wavelength
19 of 765 nm. TPC of black rice extract was expressed as mg GAE/100g.

20 **2.5. Flavonoid content**

21
22 Cai et al. (2016) adopted a slight modification method in determining the flavonoid
23 content. A sample of 0.5 ml black rice extract was prepared in a dark tube, to which 1.5
24 ml of ethanol, 0.1 ml of 10% AlCl₃, 0.1 ml of 1 M CH₃COOK, and 2.8 ml of distilled
25 water were added. The solution was homogenized and incubated at room temperature (25

1 ± 1 °C) for 30 minutes. The absorbance of the sample was measured using a UV-Vis
2 spectrophotometer with a wavelength of 415 nm, the blank solution using aquadest. The
3 standard curve uses a quercetin solution in aquadest with a concentration range of 20-100
4 ppm. Therefore, the flavonoid content of black rice extract was expressed as mg QE/100
5 g.

6 **2.6. Antioxidant activities**

7
8 Antioxidant activity was determined according to Pedro et al. (2016), with slight
9 modifications. A total of 1.5 ml of 0.2 mM DPPH in ethanol was mixed with 0.2 ml of
10 the sample into a test tube, and ethanol was added to a final volume of 3.5 ml. The tubes
11 were tightly closed, homogenized, and incubated at room temperature (25 ± 1 °C) for 60
12 minutes. The absorbance was measured at a wavelength of 517 nm, and the ability of the
13 extract to scavenge DPPH was obtained by reducing the absorbance of the blank to the
14 sample, then compared with the absorbance of the blank and expressed in % of inhibition.

15 **2.7. pH value**

16
17 The pH value of black rice powder was measured with the instrument Hanna HI 2211
18 bench pH meters (Hanna instruments Ltd.), calibration solution using standard buffer.

19 **2.8. Yield, moisture content and water activity (a_w)**

20
21 Yield was obtained from the final weight percentage of black rice powder (g) with the
22 total solids of the sample (g). Determination of the moisture content of the powder using
23 a moisture analyzer (Shimadzu MOC63u, Japan), while the a_w of the powder was
24 determined using a water activity analyzer (Rotronic Hygropalm-HP23-Aw-A,
25 Switzerland) at 25 C.

2.9. Color measurement

Color characteristics of black rice powder were measured using a calibrated Minolta CR-310 Chromameter (Konica Minolta Business Solution Asia Pte Ltd). The hue angle, H° [$\tan^{-1}(b^*/a^*)$] and chroma, C° [$(a^{*2} + b^{*2})^{1/2}$] were also determined. The H° is used to identify colors (red, yellow, green, and blue), whereas C° distinguishes between bright and dull colors (Caparino et al., 2012).

2.10. Statistical analysis

All experiments were carried out in 4 repetitions. Results are presented in terms of mean \pm standard deviation. The effect of treatment factors was analyzed using one-way ANOVA, and Duncan's post hoc test was used to detect differences between treatments. The significance level used is $p < 0.05$. Statistical analysis was carried out with the help of SPSS 22.0 software.

3. RESULTS AND DISCUSSION

3.1. Anthocyanin and pH value of capsules

The mean anthocyanin content of black rice extract powder obtained was 51.22-84.59 mg/100g (Figure 1a). This amount is higher than the research results reported by Pramitasari and Angelica (2020), which only ranged from 2.00–12.00 mg/100g. The highest anthocyanin levels were obtained at a coating ratio of 50:50 (84.59 mg/100g). There was an increase in anthocyanin levels of 65% (51.22mg/100g) compared to a ratio of 100:0. SMP generally contains β -casein components and has been reported to interact with Cyanidin-3-glucoside compounds from black soybean extract through reversible hydrogen bonds supported by an optimum pH (Kalušević et al., 2017). Previous studies also reported that casein protein interacts with malvidin-3-glucoside from grape extract

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1 through complex hydrogen bonds, which then form a hydrophobic reaction and
2 effectively increase thermal stability so that the rate of anthocyanin degradation during
3 drying can be minimized (He et al. 2016). The pH value of black rice powder ranged from
4 2.60-2.95 (Figure 1b). The pH value of the black rice extract powder increased with the
5 addition of SMP in the coating material ratio because SMP had a neutral pH value.
6 However, the increase in pH value is still in the anthocyanin stability range, at a value of
7 2-3 (Sipahli et al., 2017). Anthocyanins are more stable under acidic conditions in the
8 form of a flavylum cation structure (Khoo et al., 2017).

9 **3.2. Total phenolics, flavonoid, and antioxidant activities**

10
11 Total phenolic and flavonoid content of black rice powder is presented in Table 1. From
12 the data, it is shown that the total phenolic and flavonoid content increases with the
13 addition of SMP in the ratio of the coating material. B-casein in protein-derived
14 encapsulation forms a more substantial complex with polyphenols, and this is due to the
15 more hydrophobic nature of B-casein. (Hasni et al., 2011). The increased flavonoids are
16 associated with the structure and bonds formed from proteins with flavonoid compounds.
17 This result has previously been described by Bohin et al. (2012) on the flavonoid
18 encapsulation of tea drinks. This underlies the increase in flavonoids when the ratio of
19 SMP to coating material increases. In a previous study, Kalušević et al. (2017) reported
20 the results of high antioxidant activity in coating black rice extract using SMP. Therefore,
21 the antioxidant activity of black rice powder is thought to be derived from the
22 encapsulated anthocyanin, phenolic, and flavonoid components. In addition, the increase
23 in antioxidant activity is also potential from the maillard reaction, which produces
24 melanoidin compounds with antioxidant activity. This reaction occurs in the protein

1 component of SMP due to the high temperature during the drying process (Wang et al.,
2 2011).

3 **3.3. Aw, moisture content, and yield of microcapsules**

4
5 All samples showed an Aw value lower than 0.6 (0.49-0.58), so the black rice extract
6 powder in this study had an excellent aw value (Goyal et al., 2015). A low aw value tends
7 to result in extended shelf life. The water content values ranged from 0.91 to 1.13% (Table
8 2). The product moisture content was low to prevent particle agglomeration, avoid the
9 caking process, and improve powder dispersion and flowability (Daza et al., 2016).
10 Therefore, the powder from spray drying obtained both the aw level and the moisture
11 content in this study met the requirements and was suitable for inclusion in the dry food
12 matrix.

13 The highest yield was obtained from powders with a 50:50 coating material, which
14 is in line with the results reported in a study comparing milk protein-based microparticles
15 with other carriers (Bejšćak-Cvitanovic et al., 2015). A combination of MDE and SMP
16 coating materials resulted in good physical characteristics so that the coating material can
17 be proposed as a coating material for the bioactive component of black rice extract.

18 **3.4. Color characteristic**

19
20 Color is one of the indicators in assessing the quality of food products. During the
21 production process, the color of black rice extract is expected to be maintained until a
22 powder is obtained from drying. Therefore, observing the color measurement of black
23 rice extract powder after the encapsulation process is necessary. The results of chromatic
24 color measurements are shown in Table 3. Based on the parameters a*, b*, and hue angle,
25 it can be concluded that the color of the MD-SMP-based powder is red-violet with varying

1 brightness levels. Among all SBHs, the coating with a 50:50 ratio had the lowest
2 brightness value (40.58 ± 0.83) and the highest values for a^* and C^* , indicating that this
3 ratio produced the darkest powder with the highest portion of red color. Many reports
4 have investigated the molecular level of the bond formed between casein-based coating
5 materials and black rice anthocyanins on their stability from a thermodynamic point of
6 view. The casein-anthocyanin bond is mainly stabilized through hydrophobic interactions
7 and hydrogen bonds involving the aglycone and glucosyl moieties of the encapsulated
8 molecule so that the color can be maintained during the drying process (Aprodu et al.,
9 2019).

10 4. CONCLUSIONS

11 Based on the results of the research that has been carried out, it can be concluded that the
12 best combination of coating materials in black rice encapsulation was obtained at MDE:
13 SMP ratio of 50:50 with total anthocyanin levels obtained 84.59 mg/100g, with total
14 phenol 0.41 mg GAE/g, total flavonoid 0.21 mg QE/g and antioxidant activity 31.71%
15 RSA. Has an average value of L^* 40.5; a^* 43.64; b^* 12.75; Chroma 45.47; and Hue 16.31
16 with red-purple color types. Anthocyanin stability with a pH value of 2.95. Furthermore,
17 it has an aw of 0.49%, moisture content of 0.91%, and a yield of 51.29%. The interaction
18 between the combination of MDE and SMP as a coating material in the encapsulation
19 process of black rice extract had a very significant effect on total anthocyanin levels,
20 phenolics, total flavonoids, antioxidant activity, pH, color, aw, water content, and yield.
21

5. ACKNOWLEDGMENTS

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6. REFERENCES/ DAFTAR PUSTAKA

- Aprodu I, Milea SA, Anghel RM, Enachi E, Barbu V, Crăciunescu O, Râpeanu G, Bahrim GE, Oancea A, Stănciuc N. 2019. New Functional Ingredients Based on Microencapsulation of Aqueous Anthocyanin-Rich Extracts Derived from Black Rice (*Oryza sativa* L.). *Molecules*, 24(18): 11-14. <https://doi.org/10.3390/molecules24183389>
- Bao C, Jiang P, Chai J, Jiang Y, Li D, Bao W, Liu B, Liu B, Norde W, Li Y. 2019. The delivery of sensitive food bioactive ingredients: Absorption mechanisms, influencing factors, encapsulation techniques and evaluation models. *Food Research International*, 120: 130–140. <https://doi.org/10.1016/j.foodres.2019.02.024>
- Belščak-Cvitanović A, Lević S, Kalušević A, Špoljarić I, Đorđević V, Komes D, Nedović V. 2015. Efficiency Assessment of Natural Biopolymers as Encapsulants of Green Tea (*Camellia sinensis* L.) Bioactive Compounds by Spray Drying. *Food and Bioprocess Technology*, 8(12): 2444–2460. <https://doi.org/10.1007/s11947-015-1592-y>
- Bohin MC, Vincken JP, van der Hijden HTWM, Gruppen H. 2012. Efficacy of food proteins as carriers for flavonoids. *J. Agric. Food Chem*, 60: 4136–4143. <https://doi.org/10.1021/jf205292r>
- Bylaitė E, Venskutonis PR, Maždpierienė R. 2001. Properties of caraway (*Carum carvi* L.) essential oil encapsulated into milk protein-based matrices. *European food research and technology*, 212(6): 661–670. <https://doi.org/10.1007/s002170100297>
- Cai Z, Qu Z, Lan Y, Zhao S, Ma X, Wan Q, Jing P, Li P. 2016. Conventional, ultrasound-assisted, and accelerated-solvent extractions of anthocyanins from purple sweet potatoes. *Food Chemistry*, 197: 266–272. <https://doi.org/10.1016/j.foodchem.2015.10.110>
- Caparino OA, Tang J, Nindo CI, Sablani SS, Powers JR, Fellman JK. 2012. Effect of drying methods on the physical properties and microstructures of mango (*Philippine 'Carabao' var.*) powder. *Journal of Food Engineering*, 111(1), 135-148. <https://doi.org/10.1016/j.jfoodeng.2012.01.010>
- Cassidy A. 2018. Berry anthocyanin intake and cardiovascular health. *Molecular Aspects of Medicine*, 61: 76–82. <https://doi.org/10.1016/j.mam.2017.05.002>

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- 1
2 Daza LD, Fujita A, Fávaro-Trindade CS, Rodrigues-Ract JN, Granato D, Genovese MI.
3 2016. Effect of spray conditions on the physical properties of cagaita (*Eugenia*
4 *dysenterica DC.*) fruit extracts. Food and Bioproducts Processing, 97: 20-29.
5 <https://doi.org/10.1016/j.fbp.2015.10.001>
6
- 7 Fang Z, Bhandari B. 2010. Encapsulation of polyphenols - A review. Trends in Food
8 Science and Technology, 21(10): 510–523.
9 <https://doi.org/10.1016/j.tifs.2010.08.003>
10
- 11 Goyal A, Sharma V, Sihag MK, Komar SK, Arora S, Sabikhi L, Singh AK. 2015.
12 Development and physico-chemical characterization of microencapsulated flaxseed
13 oil powder: A functional ingredient for omega-3 fortification. Powder Technology,
14 286: 527-537. <https://doi.org/10.1016/j.powtec.2015.08.050>
15
- 16 Hasni I, Bourassa P, Hamdani S, Samson G, Carpentier R, Tajmir-Riahi HA. 2011.
17 Interaction of milk α - and β -caseins with tea polyphenols. Food chemistry, 126(2):
18 630-639. <https://doi.org/10.1016/j.foodchem.2010.11.087>
19
- 20 He Z, Xu M, Zeng M, Qin F, Chen J. 2016. Interactions of milk α -and β -casein with
21 malvidin-3-O-glucoside and their effects on the stability of grape skin anthocyanin
22 extracts. Food chemistry, 199: 314-322.
23 <https://doi.org/10.1016/j.foodchem.2015.12.035>
24
- 25 Hosoda K, Sasahara H, Matsushita K, Tamura Y, Miyaji M, Matsuyama H. 2018.
26 Anthocyanin and proanthocyanidin contents, antioxidant activity, and in situ
27 degradability of black and red rice grains. Asian-Australasian Journal of Animal
28 Sciences, 31(8): 1213–1220. <https://doi.org/10.5713/ajas.17.0655>
29
- 30 Howard LR, Brownmiller C, Prior RL, Mauromoustakos A. 2013. Improved stability of
31 chokeberry juice anthocyanins by β -cyclodextrin addition and refrigeration. Journal
32 Agric Food Chem, 61(3): 693-9. <https://doi.org/10.1021/jf3038314>.
33
- 34 Jing P, Giusti MM. 2005. Characterization of anthocyanin-rich waste from purple corn cobs
35 (*Zea mays L.*) and its application to color milk. Journal of agricultural and food
36 chemistry, 53(22): 8775–8781. <https://doi.org/10.1021/jf051247o>
37
- 38 Kalušević AM, Lević SM, Čalija BR, Milić JR, Pavlović VB, Bugarski BM, Nedović VA.
39 2017. Effects of different carrier materials on physicochemical properties of
40 microencapsulated grape skin extract. Journal of Food Science and Technology,
41 54(11): 3411–3420. <https://doi.org/10.1007/s13197-017-2790-6>
42
- 43 Khoo HE, Azlan A, Tang ST, Lim SM. 2017. Anthocyanidins and Anthocyanins: Colored
44 Pigments as Food, Pharmaceutical Ingredients, and The Potential Health Benefits.
45 Food & Nutrition Research, 61: 1 – 21.
46 <https://doi.org/10.1080/16546628.2017.1361779>
47

- 1 Lourenço SC, Moldão-Martins M, Alves VD. 2020. Microencapsulation of Pineapple Peel
2 Extract by Spray Drying Using Maltodextrin, Inulin, and Arabic Gum as Wall
3 Matrices. *Foods*, 9(6): 718. <https://doi.org/10.3390/foods9060718>
4
- 5 Mahdavi SA, Jafari SM, Ghorbani M, Assadpoor E. 2014. Spray-Drying
6 Microencapsulation of Anthocyanins by Natural Biopolymers: A Review. *Drying*
7 *Technology*, 32(5): 509–518. <https://doi.org/10.1080/07373937.2013.839562>
8
- 9 Nour V, Stampar F, Veberic R, Jakopic J. 2013. Anthocyanins profile, total phenolics and
10 antioxidant activity of black currant ethanolic extracts as influenced by genotype and
11 ethanol concentration. *Food Chemistry*, 141(2): 961–966.
12 <https://doi.org/10.1016/j.foodchem.2013.03.105>
13
- 14 Nurhidajah, Rosidi A, Yonata D, and Pranata B. 2022. Optimizing extraction of functional
15 compounds from Indonesian black rice using response surface methodology. *Food*
16 *Research*, 6(4): 83-91. [https://doi.org/10.26656/fr.2017.6\(4\).732](https://doi.org/10.26656/fr.2017.6(4).732)
17
- 18 Patil-Gadhe A, Pokharkar V. 2014. Single step spray drying method to develop
19 proliposomes for inhalation: A systematic study based on quality by design approach.
20 *Pulmonary Pharmacology and Therapeutics*, 27(2): 197–207.
21 <https://doi.org/10.1016/j.pupt.2013.07.006>
22
- 23 Pedro AC, Granato D, Rosso ND. 2016. Extraction of anthocyanins and polyphenols from
24 black rice (*Oryza sativa L.*) by modeling and assessing their reversibility and
25 stability. *Food Chemistry*, 191: 12–20.
26 <https://doi.org/10.1016/j.foodchem.2015.02.045>
27
- 28 Pramitasari R, Angelica N. 2020. Ekstraksi, Pengeringan Semprot, dan Analisis Sifat
29 Fisikokimia Antosianin Beras Hitam (*Oryza sativa L.*). *Jurnal Aplikasi Teknologi*
30 *Pangan*, 9(2): 83–94. <https://doi.org/10.17728/jatp.5889>
31
- 32 Shamaei S, Seiielou SS, Aghbashlo M, Tsotsas E, Kharaghani A. 2017.
33 Microencapsulation of walnut oil by spray drying: Effects of wall material and drying
34 conditions on physicochemical properties of microcapsules. *Innovative food science*
35 *& emerging technologies*, 39: 101-112. <https://doi.org/10.1016/j.ifset.2016.11.011>
36
- 37 Sipahli S, Mohanlall V, Mellem JJ. 2017. Stability and degradation kinetics of crude
38 anthocyanin extracts from *H. sabdariffa*. *Food Science and Technology*, 37(2): 209–
39 215. <https://doi.org/10.1590/1678-457X.14216>
40
- 41 Suhag Y, Nanda V. 2016. Optimization for spray drying process parameters of nutritionally
42 rich honey powder using response surface methodology. *Cogent Food and*
43 *Agriculture*, 2(1): 1–12. <https://doi.org/10.1080/23311932.2016.1176631>
44
- 45 Wang HY, Qian H, Yao WE, 2011. Melanoidins produced by the Maillard reaction:
46 reaction: structure and biological activity. *Food chemistry*, 128(3): 573-584.
47 <https://doi.org/10.1016/j.foodchem.2011.03.075>
48

- 1 Yamuangmorn S, Dell B, Prom-u-thai C. 2018. Effects of cooking on anthocyanin
- 2 concentration and bioactive antioxidant capacity in glutinous and non-glutinous
- 3 purple rice. *Rice Science*, 25(5): 270–278. <https://doi.org/10.1016/j.rsci.2018.04.004>
- 4
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- 7

1 Table 1. Total phenolic, total flavonoid and antioxidant activity of black rice extract
2 powder based on MD and SMP ratio

MD:SMP Ratio	Total Phenolic (mg GAE/g)	Total Flavonoid (mg QE/g)	Antioxidant Activity (%)
100:0	0.27 ± 0.01 ^a	0.14 ± 0.02 ^a	25.07 ± 0.39 ^a
90:10	0.30 ± 0.01 ^b	0.16 ± 0.01 ^{ab}	26.52 ± 0.35 ^b
80:20	0.33 ± 0.01 ^c	0.17 ± 0.01 ^{bc}	29.23 ± 0.22 ^c
70:30	0.37 ± 0.01 ^d	0.18 ± 0.03 ^{bcd}	30.37 ± 0.38 ^d
60:40	0.38 ± 0.01 ^d	0.19 ± 0.02 ^{cd}	31.18 ± 0.48 ^e
50:50	0.41 ± 0.05 ^e	0.21 ± 0.02 ^d	31.71 ± 0.94 ^e

3 Information:

- 4 1. All values are mean ± standard deviation 4 times repetition
5 2. Different superscript values showed significant differences with a 95% confidence level
6 based on the ANOVA difference test and the LSD post hoc test.

7
8
9
10 Table 2. Physical properties of black rice powder based on MD and SMP ratio

MD:SMP Ratio	Yield (%)	Moisture content (%)	a _w (%)
100:0	33.51 ± 0.82 ^a	1.13 ± 0.03 ^e	0.58 ± 0.58 ^c
90:10	35.11 ± 0.75 ^a	1.08 ± 0.01 ^{de}	0.55 ± 0.55 ^{bc}
80:20	31.14 ± 0.68 ^a	1.04 ± 0.09 ^{cd}	0.54 ± 0.54 ^{bc}
70:30	34.06 ± 0.82 ^a	0.99 ± 0.01 ^{bc}	0.52 ± 0.52 ^{ab}
60:40	47.64 ± 0.73 ^b	0.96 ± 0.02 ^{cb}	0.51 ± 0.51 ^{ab}
50:50	51.29 ± 0.49 ^b	0.91 ± 0.05 ^a	0.49 ± 0.49 ^a

11 Information:

- 12 1. All values are mean ± standard deviation 4 times repetition
13 2. Different superscript values showed significant differences with a 95% confidence level
14 based on the ANOVA difference test and the LSD post hoc test.

15
16
17 Table 3. Color parameters of black rice powder based on MD and SMP ratio

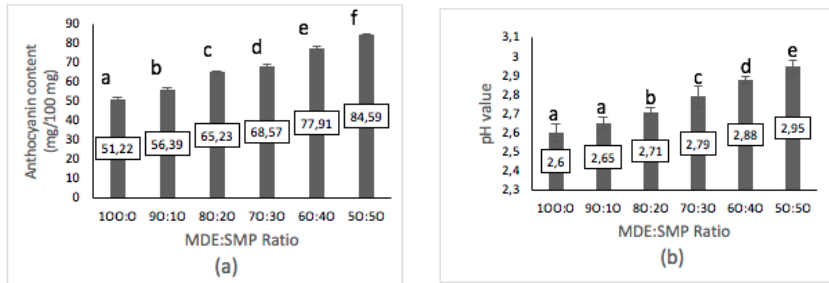
MD:SMP Ratio	L*	a*	b*	C*	H°	Color
100:0	56.06 ± 0.79 ^d	26.83 ± 0.66 ^a	4.43 ± 0.31 ^a	27.20 ± 0.62 ^a	9.39 ± 0.81 ^a	Red-violet
90:10	55.46 ± 1.46 ^c	28.48 ± 0.12 ^b	4.30 ± 0.51 ^a	28.80 ± 0.16 ^b	8.58 ± 0.99 ^a	Red-violet
80:20	52.91 ± 0.61 ^c	28.71 ± 0.39 ^b	5.34 ± 0.50 ^{ab}	29.20 ± 0.35 ^b	10.55 ± 1.06 ^a	Red-violet
70:30	49.07 ± 0.82 ^b	35.78 ± 1.45 ^c	6.36 ± 1.56 ^b	36.36 ± 1.65 ^c	10.03 ± 2.13 ^a	Red-violet
60:40	48.06 ± 0.54 ^a	39.90 ± 0.47 ^d	9.71 ± 0.39 ^c	41.06 ± 0.39 ^d	13.68 ± 0.65 ^b	Red-violet
50:50	40.58 ± 0.83 ^a	43.64 ± 1.43 ^e	12.75 ± 0.83 ^d	45.47 ± 1.16 ^e	16.31 ± 1.50 ^c	Red-violet

18 Information:

- 19 1. All values are mean ± standard deviation 4 times repetition
20 2. Different superscript values showed significant differences with a 95% confidence level
21 based on the ANOVA difference test and the LSD post hoc test.

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3
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Figure 1. Anthocyanin and pH value black rice powder based on MDE:SMP Ratio

3. Revised version received (14-11-2022)

1 dari 6 perlakuan yaitu rasio MDE:SMP (100:0, 90:10, 80:20, 70:30, 60:40 dan 50:50).
2 Hasil penelitian menunjukkan ada pengaruh yang sangat nyata dari rasio MDE dan SMP
3 terhadap karakteristik fisik dan kimia serbuk beras hitam. Rasio MDE dan SMP 50:50
4 menghasilkan karakteristik fisik dan kimia serbuk beras hitam terbaik.

5 **Kata Kunci:** ekstrak beras hitam, enkapsulasi, bahan penyalut, karakteristik fisikokimia.

6
7

8 1. INTRODUCTION

9

10
11 **Black rice (*Oryza sativa* L.) is a pigmented rice that has black bran covering the**
12 **endosperm. Consumption of pigmented rice, such as black rice is currently starting to**
13 **increase because people have begun to change their food consumption patterns towards**
14 **foods that are beneficial to health, namely functional foods. Black rice has potential as a**
15 **functional food because it contains bioactive components, namely polyphenolic**
16 **compounds, flavonoids, and anthocyanins that act as antioxidants, anti-inflammatory and**
17 **have other important health benefits (Kong et al., 2012; Hosoda et al., 2018).**

18 Anthocyanin pigments in black rice can be obtained by extraction using the
19 maceration method. Generally, ethanol solvent is used in the maceration method. Ethanol
20 has advantages over other solvents such as methanol and acetone, namely it is more
21 economical and has low toxicity (Nour et al., 2013). Acidified ethanol has been confirmed
22 to increase yield and anthocyanin content in black rice extract (Pedro et al., 2016).
23 Extraction with heat-assisted maceration technique based on a recent study (Nurhidajah
24 et al., 2022) was reported to be very effective in preparing black rice anthocyanin extract.
25 **Anthocyanin extracts can be degraded during the storage process. This is due to its low**
26 **chemical stability to environmental conditions including temperature, light intensity, and**
27 **pH (Cassidy, 2018; Fernandes, 2014). The anthocyanin decomposition process occurs**
28 **faster at high temperatures (Patras et al., 2010). Anthocyanins are unstable at high light**
29 **intensity. The release of the sugar group causes the anthocyanin aglycones that are formed**

1 to fade quickly when exposed to light (Janna et al., 2006). Encapsulation techniques can
2 overcome this weakness using a freeze dryer, spray dryer, and others (Bao et al., 2019;
3 Fang & Bhandari, 2010; Howard et al., 2013).

4 Encapsulation using a spray dryer is an effective solution to protect and increase
5 the stability of anthocyanin bioactive compounds (Kalušević et al., 2017). This technique
6 has been widely used in the food industry. In addition to being applicable, the spray drying
7 (SD) method for thermosensitive products is due to a short contact time with a heat source
8 (Lourenço et al., 2020). The success of SD is influenced by several things, including inlet
9 and outlet temperatures, air flow rate, feed flow rate, speed of the atomizer, to the type
10 and concentration of coating material used (Patil-Gadhe & Pokharkar, 2014). The coating
11 material is one of the factors that will affect the encapsulated powder.

12 Mahdavi et al. (2014) reported maltodextrin (MDE) is extensively used as a coating
13 material in anthocyanin encapsulation using the SD technique. MDE as a coating material
14 with various advantages, including high solubility, low viscosity, and maintaining the
15 stability of bioactive compounds (Suhag & Nanda, 2016). Recent research by Kalušević
16 et al. (2017) reported that skimmed milk powder (SMP) reported that SMP coating
17 material could encapsulate anthocyanin components at higher levels than maltodextrin.
18 As a coating material, SMP has non-stick properties and high encapsulation efficiency.
19 Apart from being a source of protein, lactose, vitamins, and minerals, this ingredient
20 provides additional nutritional value (Bylaitė et al., 2001; Jing & Giusti, 2005; Shamaei
21 et al., 2016). However, the use of SMP as a coating material in the encapsulation of black
22 rice extract still has a weakness, namely the coating efficiency of phenolic and flavonoid
23 components is still very low (Kalušević et al., 2017).

1 MD and SMP coating materials **emulsion** with different ratios (100:0, 90:10, 80:20,
 2 70:30, 60:40 and 50:50) were prepared at a concentration of 20% (w/v) with deionized
 3 water at room temperature. Anthocyanin extract was mixed with each coating material in
 4 a ratio of 1:1 (v/v). Each mixture was homogenized separately for 15 min at 3000 rpm. It
 5 was dried using a laboratory-scale spray dryer with an inlet air temperature of 120 ± 1 C,
 6 an outlet air temperature of 80 ± 5 C, and a feed flow rate of 6.0 mL/minute at a pressure
 7 of 1.5 bar. The obtained **coated anthocyanin powder** was collected and stored at -20°C
 8 until analyzed.

9 **2.4. Anthocyanin content (Yamuangmorn et al., 2018)**

10 A differential pH method was used to determine **coated anthocyanin powder**. First,
 11 **sample (1 g)** was dissolved with 1 mL of ethanol in a test tube, then mixed separately
 12 with two buffer solutions (1 mL of potassium chloride buffer pH 1.0 and 1 mL sodium
 13 acetate buffer pH 4.5), and incubated at room temperature (± 25 °C) for 15 minutes. The
 14 absorbance was read by spectrophotometer at 520 and 700 nm. The absorbance value was
 15 obtained by subtracting difference in absorbance at a wavelength of 520 nm and 700 nm
 16 at pH 1.0 with the difference in absorbance at pH 4.5. **The absorbance value of the extract**
 17 **sample was calculated using the equation:**

$$19 \quad A = [(A_{520} - A_{700})_{\text{pH1}} - (A_{520} - A_{700})_{\text{pH4.5}}]$$

20 **Then anthocyanin content of black rice extract was expressed in mg/100g:**

$$21 \quad \text{Anthocyanin content (mg/100 g)} = \frac{A \times MW \times DF \times V \times 100}{E \times 1 \times W}$$

22 **A** = absorbance

23 **MW** = molecular weight of cyanidin-3-glucoside (448.8 g/mol)

24 **DF** = dilution factor

1 V = volume of the sample mother liquor

2 E = coefficient of molar absorptivity of cyanidin-3-glucoside (26900 l/mol cm)

3 W = sample weight (g)

4 2.5. Total phenolics

5
6 Total phenolics content (TPC) used the Folin-Ciocalteu technique based on (Pedro et al.,
7 2016) with minor modifications. In a dark tube, coated anthocyanin powder (0.5 g) was
8 made, and 5 mL of Folin-Ciocalteu 10% (v/v) reagent was added. After homogenizing
9 the solution for 5 minutes, 4 mL of 7.5% Na₂CO₃ (w/v) was added. The mixture was
10 incubated at room temperature (25 °C) for 60 minutes. As a blank solution, ethanol is
11 employed. A standard solution of gallic acid in ethanol with a concentration of 100-500
12 ppm is utilized. The absorbance was read with a UV-Vis spectrophotometer set at 765
13 nm. TPC was calculated as mg GAE/100g of black rice extract.

14 2.6. Flavonoid content

15
16 Flavonoid content was determined according to Cai et al., (2016), with modifications. In
17 a dark tube, 0.5 ml of black rice extract was mixed with 1.5 ml of ethanol, 0.1 ml of 10%
18 AlCl₃, 0.1 ml of 1 M CH₃COOK, and 2.8 ml of distilled water. The solution was
19 homogenized and incubated for 30 minutes at room temperature (25 ± 1 °C). The
20 absorbance of the sample was read with a UV-Vis spectrophotometer at 415 nm, and the
21 blank solution was aquadest. A quercetin solution in aquadest with a concentration range
22 of 20-100 ppm is used in the standard curve. The flavonoid concentration of black rice
23 extract was therefore reported as mg QE/100 g.

24 2.7. Antioxidant activities

25

1 Antioxidant activity was determined according to Pedro et al. (2016), with slight
2 modifications. A total of 1.5 ml of 0.2 mM DPPH in ethanol was mixed with 0.2 ml of
3 the sample into a test tube, and ethanol was added to a final volume of 3.5 ml. The tubes
4 were tightly closed, homogenized, and incubated at room temperature (25 ± 1 °C) for 60
5 minutes. The absorbance was read at a wavelength of 517 nm and calculated using the
6 equation:

$$7 \quad \% \text{ RSA} = \frac{\text{Abs blanko} \times \text{Abs test}}{\text{Abs blanko}} \times 100\%$$

8 2.8. pH value

9
10 The pH value was measuring according to Park et al., (2016). Coated anthocyanin powder
11 sample (4g) was mixed with 16 mL of distilled water in a vortex mixer for 1 min. The pH
12 was recorded using a instrument Hanna HI 2211 bench pH meters (Hanna instruments
13 Ltd.), calibration solution using standard buffer.

14 2.9. Yield, moisture content and water activity (a_w)

15
16 Yield was obtained from the final weight percentage of coated anthocyanin powder (g)
17 with the total solids of the sample (g). Determination of the moisture content of the
18 powder using a moisture analyzer (Shimadzu MOC63u, Japan), while the a_w of the
19 powder was determined using a water activity analyzer (Rotronic Hygropalm-HP23-Aw-
20 A, Switzerland) at 25 C.

21 2.10. Color measurement

22
23 Color characteristics of coated anthocyanin powder were measured using a calibrated
24 Minolta CR-310 Chromameter (Konica Minolta Business Solution Asia Pte Ltd). The hue
25 angle, H° [$\tan^{-1}(b^*/a^*)$] and chroma, C° [$(a^{*2} + b^{*2})^{1/2}$] were also determined. The H°

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We will consider it for future research.

1 is used to identify colors (red, yellow, green, and blue), whereas C° distinguishes between
2 bright and dull colors (Caparino et al., 2012).

3 **2.11. Statistical analysis**

4
5 All experiments were carried out in 4 repetitions. Results are presented in terms of mean
6 ± standard deviation. The effect of treatment factors was analyzed using one-way
7 ANOVA, and Duncan's post hoc test was used to detect differences between treatments.
8 The significance level used is $p < 0.05$. Statistical analysis was carried out with the help of
9 SPSS 22.0 software.

10 **3. RESULTS AND DISCUSSION**

11 **3.1. Anthocyanin and pH value of capsules**

12
13
14 The mean anthocyanin content of black rice **coated anthocyanin powder** obtained was
15 51.22-84.59 mg/100g (Figure 1a). This amount is higher than the research results reported
16 by Pramitasari and Angelica (2020), which only ranged from 2.00–12.00 mg/100g. The
17 highest anthocyanin levels were obtained at a coating ratio of 50:50 (84.59 mg/100g).
18 There was an increase in anthocyanin levels of 65% (51.22mg/100g) compared to a ratio
19 of 100:0. **SMP 50% as a coating material was able to trap more anthocyanins, resulting**
20 **in a powder with the highest anthocyanin content, significantly different from all**
21 **treatments.** SMP generally contains β -casein components and has been reported to
22 interact with Cyanidin-3-glucoside compounds from black soybean extract through
23 reversible hydrogen bonds supported by an optimum pH (Kalušević et al., 2017). Previous
24 studies also reported that casein protein interacts with malvidin-3-glucoside from grape
25 extract through complex hydrogen bonds, which then form a hydrophobic reaction and
26 effectively increase thermal stability so that the rate of anthocyanin degradation during

1 drying can be minimized (He et al. 2016). The pH value of **coated anthocyanin powder**
2 ranged from 2.60-2.95 (Figure 1b). **The pH value increased** with the addition of SMP in
3 the coating material ratio because SMP had a neutral pH value. However, the increase in
4 pH value is still in the anthocyanin stability range, at a value of 2-3 (Sipahli et al., 2017).
5 Anthocyanins are more stable under acidic conditions in the form of a flavylium cation
6 structure (Khoo et al., 2017).

7 **3.2. Total phenolics, flavonoid, and antioxidant activities**

8
9 Total phenolic and flavonoid content of **coated anthocyanin powder** is presented in Table
10 1. **From the data, it is shown that the total phenolic and flavonoid content increases with**
11 **the addition of SMP 50% in the ratio of the coating material, and significantly different**
12 **with all treatments.** B-casein in protein-derived encapsulation forms a more substantial
13 complex with polyphenols, and this is due to the more hydrophobic nature of B-casein.
14 (Hasni et al., 2011). The increased flavonoids are associated with the structure and bonds
15 formed from proteins with flavonoid compounds. This result has previously been
16 described by Bohin et al. (2012) on the flavonoid encapsulation of tea drinks. This
17 underlies the increase in flavonoids when the ratio of SMP to coating material increases.
18 In a previous study, Kalušević et al. (2017) reported the results of high antioxidant activity
19 in coating black rice extract using SMP. Therefore, the antioxidant activity of **coated**
20 **anthocyanin powder** is thought to be derived from the encapsulated anthocyanin,
21 phenolic, and flavonoid components. In addition, the increase in antioxidant activity is
22 also potential from the **Maillard** reaction, which produces melanoidin compounds with
23 antioxidant activity. This reaction occurs in the protein component of SMP due to the
24 high temperature during the drying process (Wang et al., 2011).

3.3. Aw, moisture content, and yield of coated anthocyanin powder

All samples showed an Aw value lower than 0.6 (0.49-0.58), so the coated anthocyanin powder in this study had an excellent aw value (Goyal et al., 2015). A low aw value tends to result in extended shelf life. There is a positive tendency to increase the ratio of SMP as a coating material, SMP 50% is able to increase the aw of the product significantly. The water content values ranged from 0.91 to 1.13% (Table 2), the use of SMP 50% significantly made anthocyanin powder with the lowest water content. On the other hand, coatings with high MDE content produce high water content due to the nature of MDE, which quickly releases water (Kustyawati et al., 2022). The product moisture content was low to prevent particle agglomeration, avoid the caking process, and improve powder dispersion and flowability (Daza et al., 2016). Therefore, the powder from spray drying obtained both the aw level and the moisture content in this study met the requirements and was suitable for inclusion in the dry food matrix.

The highest yield was significantly obtained from powders with a 50:50 coating material, which is in line with the results reported in a study comparing milk protein-based microparticles with other carriers (Belščak-Cvitanovic et al., 2015). A combination of MDE and SMP coating materials resulted in good physical characteristics so that the coating material can be proposed as a coating material for the bioactive component of black rice extract.

3.4. Color characteristic

Color is one of the indicators in assessing the quality of food products. During the production process, the color of black rice extract is expected to be maintained until a powder is obtained from drying. Therefore, observing the color measurement of coated anthocyanin powder after the encapsulation process is necessary. The results of chromatic

1 color measurements are shown in Table 3. Based on the parameters a^* , b^* , and hue angle,
2 it can be concluded that the color of the MD-SMP-based powder is red-violet with varying
3 brightness levels. Among all coated anthocyanin powders, coating with a ratio of 50:50
4 differed significantly by having the lowest brightness value (40.58 ± 0.83) and the highest
5 value for a^* and C^* with significant differences with other treatments. Indicating that this
6 ratio produced the darkest powder with the highest portion of red color. Many reports
7 have investigated the molecular level of the bond formed between casein-based coating
8 materials and black rice anthocyanins on their stability from a thermodynamic point of
9 view. The casein-anthocyanin bond is mainly stabilized through hydrophobic interactions
10 and hydrogen bonds involving the aglycone and glucosyl moieties of the encapsulated
11 molecule so that the color can be maintained during the drying process (Aprodu et al.,
12 2019).

13 4. CONCLUSIONS

14 It can be concluded that the best combination of coating materials in black rice
15 encapsulation was obtained at MDE: SMP ratio of 50:50 with total anthocyanin levels
16 obtained 84.59 mg/100g, with total phenol 0.41 mg GAE/g, total flavonoid 0.21 mg QE/g
17 and antioxidant activity 31.71% RSA. Has an average value of L^* 40.5; a^* 43.64; b^*
18 12.75; Chroma 45.47; and Hue 16.31 with red-purple color types. Anthocyanin stability
19 with a pH value of 2.95. Furthermore, it has an a_w of 0.49%, moisture content of 0.91%,
20 and a yield of 51.29%. The combination of MDE and SMP as a coating material in the
21 encapsulation process of black rice extract had a very significant effect on total
22 anthocyanin levels, phenolics, total flavonoids, antioxidant activity, pH, color, a_w , water
23 content, and yield.
24

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6. REFERENCES

- Aprodu I, Milea SA, Anghel RM, Enachi E, Barbu V, Crăciunescu O, Râpeanu G, Bahrim GE, Oancea A, Stănciuc N. 2019. New Functional Ingredients Based on Microencapsulation of Aqueous Anthocyanin-Rich Extracts Derived from Black Rice (*Oryza sativa* L.). *Molecules*, 24(18): 11-14. <https://doi.org/10.3390/molecules24183389>
- Bao C, Jiang P, Chai J, Jiang Y, Li D, Bao W, Liu B, Liu B, Norde W, Li Y. 2019. The delivery of sensitive food bioactive ingredients: Absorption mechanisms, influencing factors, encapsulation techniques and evaluation models. *Food Research International*, 120: 130–140. <https://doi.org/10.1016/j.foodres.2019.02.024>
- Belščak-Cvitanović A, Lević S, Kalušević A, Špoljarić I, Đorđević V, Komes D, Nedović V. 2015. Efficiency Assessment of Natural Biopolymers as Encapsulants of Green Tea (*Camellia sinensis* L.) Bioactive Compounds by Spray Drying. *Food and Bioprocess Technology*, 8(12): 2444–2460. <https://doi.org/10.1007/s11947-015-1592-y>
- Bohin MC, Vincken JP, van der Hijden HTWM, Gruppen H. 2012. Efficacy of food proteins as carriers for flavonoids. *J. Agric. Food Chem*, 60: 4136–4143. <https://doi.org/10.1021/jf205292r>
- Bylaitė E, Venskutonis PR, Maždpierienė R. 2001. Properties of caraway (*Carum carvi* L.) essential oil encapsulated into milk protein-based matrices. *European food research and technology*, 212(6): 661–670. <https://doi.org/10.1007/s002170100297>
- Cai Z, Qu Z, Lan Y, Zhao S, Ma X, Wan Q, Jing P, Li P. 2016. Conventional, ultrasound-assisted, and accelerated-solvent extractions of anthocyanins from purple sweet potatoes. *Food Chemistry*, 197: 266–272. <https://doi.org/10.1016/j.foodchem.2015.10.110>
- Caparino OA, Tang J, Nindo CI, Sablani SS, Powers JR, Fellman JK. 2012. Effect of drying methods on the physical properties and microstructures of mango (*Philippine 'Carabao' var.*) powder. *Journal of Food Engineering*, 111(1), 135-148. <https://doi.org/10.1016/j.jfoodeng.2012.01.010>
- Cassidy A. 2018. Berry anthocyanin intake and cardiovascular health. *Molecular Aspects of Medicine*, 61: 76–82. <https://doi.org/10.1016/j.mam.2017.05.002>

- 1
2 Daza LD, Fujita A, Fávoro-Trindade CS, Rodrigues-Ract JN, Granato D, Genovese MI.
3 2016. Effect of spray conditions on the physical properties of cagaita (*Eugenia*
4 *dysenterica DC.*) fruit extracts. Food and Bioproducts Processing, 97: 20-29.
5 <https://doi.org/10.1016/j.fbp.2015.10.001>
6
- 7 Fang Z, Bhandari B. 2010. Encapsulation of polyphenols - A review. Trends in Food
8 Science and Technology, 21(10): 510–523.
9 <https://doi.org/10.1016/j.tifs.2010.08.003>
10
- 11 Fernandes I, Faria A, Calhau C, de Freitas V, Mateus N. 2014. Bioavailability of
12 anthocyanins and derivatives. Journal of Functional Foods, 7: 54-66.
13 <http://dx.doi.org/10.1016/j.jff.2013.05.010>
14
- 15 Goyal A, Sharma V, Sihag MK, Komar SK, Arora S, Sabikhi L, Singh AK. 2015.
16 Development and physico-chemical characterization of microencapsulated flaxseed
17 oil powder: A functional ingredient for omega-3 fortification. Powder Technology,
18 286: 527-537. <https://doi.org/10.1016/j.powtec.2015.08.050>
19
- 20 Hasni I, Bourassa P, Hamdani S, Samson G, Carpentier R, Tajmir-Riahi HA. 2011.
21 Interaction of milk α - and β -caseins with tea polyphenols. Food chemistry, 126(2):
22 630-639. <https://doi.org/10.1016/j.foodchem.2010.11.087>
23
- 24 He Z, Xu M, Zeng M, Qin F, Chen J. 2016. Interactions of milk α - and β -casein with
25 malvidin-3-O-glucoside and their effects on the stability of grape skin anthocyanin
26 extracts. Food chemistry, 199: 314-322.
27 <https://doi.org/10.1016/j.foodchem.2015.12.035>
28
- 29 Hosoda K, Sasahara H, Matsushita K, Tamura Y, Miyaji M, Matsuyama H. 2018.
30 Anthocyanin and proanthocyanidin contents, antioxidant activity, and in situ
31 degradability of black and red rice grains. Asian-Australasian Journal of Animal
32 Sciences, 31(8): 1213–1220. <https://doi.org/10.5713/ajas.17.0655>
33
- 34 Howard LR, Brownmiller C, Prior RL, Mauromoustakos A. 2013. Improved stability of
35 chokeberry juice anthocyanins by β -cyclodextrin addition and refrigeration. Journal
36 Agric Food Chem, 61(3): 693-9. <https://doi.org/10.1021/jf3038314>.
37
- 38 Janna O, Khairul A, Maizah M, Mohd MY. 2006. Flower Pigment Analysis of *Melastoma*
39 *malabatricum*, Journal of African Biotechnology 5 (2):170-174.
40
- 41 Jing P, Giusti MM. 2005. Characterization of anthocyanin-rich waste from purple corn cobs
42 (*Zea mays L.*) and its application to color milk. Journal of agricultural and food
43 chemistry, 53(22): 8775–8781. <https://doi.org/10.1021/jf051247o>
44
- 45 Kalušević AM, Lević SM, Čalija BR, Milić JR, Pavlović VB, Bugarski BM, Nedović VA.
46 2017. Effects of different carrier materials on physicochemical properties of
47 microencapsulated grape skin extract. Journal of Food Science and Technology,
48 54(11): 3411–3420. <https://doi.org/10.1007/s13197-017-2790-6>

- 1
2 Khoo HE, Azlan A, Tang ST, Lim SM. 2017. Anthocyanidins and Anthocyanins: Colored
3 Pigments as Food, Pharmaceutical Ingredients, and The Potential Health Benefits.
4 Food & Nutrition Research, 61: 1–21.
5 <https://doi.org/10.1080/16546628.2017.1361779>
6
- 7 Kong S, Kim DJ, Oh SK, Choi IS, Jeong HS, Lee J. 2012. Black Rice Bran as an Ingredient
8 in Noodles: Chemical and Functional Evaluation. Journal of Food Science, 77(3):
9 307-307. <https://doi.org/10.1111/j.1750-3841.2011.02590.x>
10
- 11 Kustyawati ME, Sugiharto R, Rini R. 2022. Microencapsulation of Green Capulaga
12 (*Elettaria cardamomum*) Essential Oil with Maltodextrin and Its Applications in
13 Coffee Drink. Jurnal Teknologi Pertanian Lampung, 11(3): 531-541.
14 <http://dx.doi.org/10.23960/jtep-l.v11i3.531-541>
15
- 16 Lourenço SC, Moldão-Martins M, Alves VD. 2020. Microencapsulation of Pineapple Peel
17 Extract by Spray Drying Using Maltodextrin, Inulin, and Arabic Gum as Wall
18 Matrices. Foods, 9(6): 718. <https://doi.org/10.3390/foods9060718>
19
- 20 Mahdavi SA, Jafari SM, Ghorbani M, Assadpoor E. 2014. Spray-Drying
21 Microencapsulation of Anthocyanins by Natural Biopolymers: A Review. Drying
22 Technology, 32(5): 509–518. <https://doi.org/10.1080/07373937.2013.839562>
23
- 24 Nour V, Stampar F, Veberic R, Jakopic J. 2013. Anthocyanins profile, total phenolics and
25 antioxidant activity of black currant ethanolic extracts as influenced by genotype and
26 ethanol concentration. Food Chemistry, 141(2): 961–966.
27 <https://doi.org/10.1016/j.foodchem.2013.03.105>
28
- 29 Nurhidajah, Rosidi A, Yonata D, and Pranata B. 2022. Optimizing extraction of functional
30 compounds from Indonesian black rice using response surface methodology. Food
31 Research, 6(4): 83-91. [https://doi.org/10.26656/fr.2017.6\(4\).732](https://doi.org/10.26656/fr.2017.6(4).732)
32
- 33 Park SY, Kim HY. 2016. Effect of Black Rice Powder Levels on Quality Properties of
34 Emulsion-type Sausage. Korean Journal for Food Science of Animal Resources,
35 36(6): 737–743.
36
- 37 Patil-Gadhe A, Pokharkar V. 2014. Single step spray drying method to develop
38 proliposomes for inhalation: A systematic study based on quality by design approach.
39 Pulmonary Pharmacology and Therapeutics, 27(2): 197–207.
40 <https://doi.org/10.1016/j.pupt.2013.07.006>
41
- 42 Patras A, Brunton NP, O'donnell C, Tiwari, BK. 2010. Effect of therma processing on
43 anthocyanin stability in foods; Mechanisms and kinetics of degradation a review.
44 Trends in Food Science & Technology, 21: 3-11.
45 <https://doi.org/10.1016/j.tifs.2009.07.004>
46
- 47 Pedro AC, Granato D, Rosso ND. 2016. Extraction of anthocyanins and polyphenols from
48 black rice (*Oryza sativa L.*) by modeling and assessing their reversibility and

- 1 stability. Food Chemistry, 191: 12–20.
2 <https://doi.org/10.1016/j.foodchem.2015.02.045>
3
- 4 Pramitasari R, Angelica N. 2020. Ekstraksi, Pengeringan Semprot, dan Analisis Sifat
5 Fisikokimia Antosianin Beras Hitam (*Oryza sativa L.*). Jurnal Aplikasi Teknologi
6 Pangan, 9(2): 83–94. <https://doi.org/10.17728/jatp.5889>
7
- 8 Shamaei S, Seiiedlou SS, Aghbashlo M, Tsotsas E, Kharaghani A. 2017.
9 Microencapsulation of walnut oil by spray drying: Effects of wall material and drying
10 conditions on physicochemical properties of microcapsules. Innovative food science
11 & emerging technologies, 39: 101-112. <https://doi.org/10.1016/j.ifset.2016.11.011>
12
- 13 Sipahli S, Mohanlall V, Mellem JJ. 2017. Stability and degradation kinetics of crude
14 anthocyanin extracts from *H. sabdariffa*. Food Science and Technology, 37(2): 209–
15 215. <https://doi.org/10.1590/1678-457X.14216>
16
- 17 Suhag Y, Nanda V. 2016. Optimization for spray drying process parameters of nutritionally
18 rich honey powder using response surface methodology. Cogent Food and
19 Agriculture, 2(1): 1–12. <https://doi.org/10.1080/23311932.2016.1176631>
20
- 21 Wang HY, Qian H, Yao WE, 2011. Melanoidins produced by the Maillard reaction:
22 reaction: structure and biological activity. Food chemistry, 128(3): 573-584.
23 <https://doi.org/10.1016/j.foodchem.2011.03.075>
24
- 25 Yamuangmorn S, Dell B, Prom-u-thai C. 2018. Effects of cooking on anthocyanin
26 concentration and bioactive antioxidant capacity in glutinous and non-glutinous
27 purple rice. Rice Science, 25(5): 270–278. <https://doi.org/10.1016/j.rsci.2018.04.004>
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1 Table 1. Total phenolic, total flavonoid and antioxidant activity of **coated anthocyanin**
 2 **powder** based on MD and SMP ratio

MD:SMP Ratio	Total Phenolic (mg GAE/g)	Total Flavonoid (mg QE/g)	Antioxidant Activity (%)
100:0	0.27 ± 0.01 ^a	0.14 ± 0.02 ^a	25.07 ± 0.39 ^a
90:10	0.30 ± 0.01 ^b	0.16 ± 0.01 ^{ab}	26.52 ± 0.35 ^b
80:20	0.33 ± 0.01 ^c	0.17 ± 0.01 ^{bc}	29.23 ± 0.22 ^c
70:30	0.37 ± 0.01 ^d	0.18 ± 0.03 ^{bcd}	30.37 ± 0.38 ^d
60:40	0.38 ± 0.01 ^d	0.19 ± 0.02 ^{cd}	31.18 ± 0.48 ^e
50:50	0.41 ± 0.05 ^e	0.21 ± 0.02 ^d	31.71 ± 0.94 ^e

3 Information:

- 4 1. All values are mean ± standard deviation 4 times repetition
 5 2. Different superscript values showed significant differences with a 95% confidence level
 6 based on the ANOVA difference test and the LSD post hoc test.

10 Table 2. Physical properties of **coated anthocyanin powder** based on MD and SMP ratio

MD:SMP Ratio	Yield (%)	Moisture content (%)	a _w (%)
100:0	33.51 ± 0.82 ^a	1.13 ± 0.03 ^e	0.58 ± 0.58 ^c
90:10	35.11 ± 0.75 ^a	1.08 ± 0.01 ^{de}	0.55 ± 0.55 ^{bc}
80:20	31.14 ± 0.68 ^a	1.04 ± 0.09 ^{cd}	0.54 ± 0.54 ^{bc}
70:30	34.06 ± 0.82 ^a	0.99 ± 0.01 ^{bc}	0.52 ± 0.52 ^{ab}
60:40	47.64 ± 0.73 ^b	0.96 ± 0.02 ^{cb}	0.51 ± 0.51 ^{ab}
50:50	51.29 ± 0.49 ^b	0.91 ± 0.05 ^a	0.49 ± 0.49 ^a

11 Information:

- 12 1. All values are mean ± standard deviation 4 times repetition
 13 2. Different superscript values showed significant differences with a 95% confidence level
 14 based on the ANOVA difference test and the LSD post hoc test.

17 Table 3. Color parameters of **coated anthocyanin powder** based on MD and SMP ratio

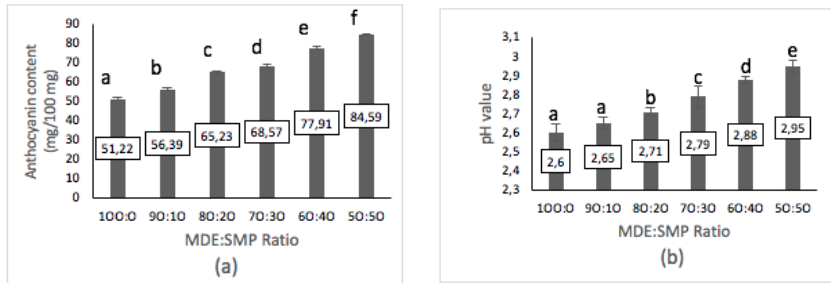
MD:SMP Ratio	L*	a*	b*	C*	H°	Color
100:0	56.06 ± 0.79 ^d	26.83 ± 0.66 ^a	4.43 ± 0.31 ^a	27.20 ± 0.62 ^a	9.39 ± 0.81 ^a	Red-violet
90:10	55.46 ± 1.46 ^c	28.48 ± 0.12 ^b	4.30 ± 0.51 ^a	28.80 ± 0.16 ^b	8.58 ± 0.99 ^a	Red-violet
80:20	52.91 ± 0.61 ^c	28.71 ± 0.39 ^b	5.34 ± 0.50 ^{ab}	29.20 ± 0.35 ^b	10.55 ± 1.06 ^a	Red-violet
70:30	49.07 ± 0.82 ^b	35.78 ± 1.45 ^c	6.36 ± 1.56 ^b	36.36 ± 1.65 ^c	10.03 ± 2.13 ^a	Red-violet
60:40	48.06 ± 0.54 ^b	39.90 ± 0.47 ^d	9.71 ± 0.39 ^c	41.06 ± 0.39 ^d	13.68 ± 0.65 ^b	Red-violet
50:50	40.58 ± 0.83 ^a	43.64 ± 1.43 ^e	12.75 ± 0.83 ^d	45.47 ± 1.16 ^e	16.31 ± 1.50 ^c	Red-violet

18 Information:

- 19 1. All values are mean ± standard deviation 4 times repetition
 20 2. Different superscript values showed significant differences with a 95% confidence level
 21 based on the ANOVA difference test and the LSD post hoc test.

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Figure 1. Anthocyanin and pH value coated anthocyanin powder based on MDE:SMP Ratio

4. Paper Accepted (15-11-2022)

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Berdasarkan revisi yang kami terima, dengan gembira kami sampaikan bahwa naskah Saudara yang berjudul: **Psychochemical Characteristics of Black Rice Powder Based on Maltodextrin and Skimmed Milk Powder Ratio as Encapsulant** dinyatakan **ACCEPTED** dan dapat dipublikasikan di JTEP.

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Physicochemical Characteristics of Anthocyanin Extract Powder from Black Rice Based on Maltodextrin and Skimmed Milk Powder Ratio as Encapsulant

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ABSTRACT

Anthocyanins are bioactive components in black rice. Black rice anthocyanins are potent antioxidants, so they have the potential to be developed into functional food products. As a bioactive component, anthocyanins in extracts have low stability to environmental conditions such as light, temperature, and pH. Encapsulation process using the spray drying technique is known to protect and increase the stability of anthocyanin bioactive compounds. Maltodextrin (MDE) is widely used as a coating material in the encapsulation of anthocyanins using the spray drying technique with many advantages. However, skim milk powder (SMP) was found to encapsulate black rice anthocyanins efficiently. This study's general objective was to determine the effect of the ratio of MDE and SMP as an encapsulant on black rice extract powder's physical and chemical characteristics. The research method is an experimental type using a single factor Completely Randomized Design (CRD), which consisted of 6 treatments, namely the MDE:SMP ratio (100:0, 90:10, 80:20, 70:30, 60:40, and 50:50). The results showed that there was a very significant effect of the ratio of MDE and SMP on the physical and chemical characteristics of coated anthocyanin powder. MDE and SMP ratios of 50:50 resulted in coated anthocyanin powder's best physical and chemical characteristics.

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1. INTRODUCTION

Black rice (*Oryza sativa* L.) is a pigmented rice that has black bran covering the endosperm. Consumption of pigmented rice, such as black rice is currently starting to increase because people have begun to change their food consumption patterns towards foods that are beneficial to health, namely functional foods. Black rice has potential as a functional food because it contains bioactive components, namely polyphenolic compounds, flavonoids, and anthocyanins that act as antioxidants, anti-inflammatory and have other important health benefits (Kong *et al.*, 2012; Hosoda *et al.*, 2018).

Anthocyanin pigments in black rice can be obtained by extraction using the maceration method. Generally, ethanol solvent is used in the maceration method. Ethanol has advantages over other solvents such as methanol and acetone, namely it is more economical and has low toxicity (Nour *et al.*, 2013). Acidified ethanol has been confirmed to increase yield and anthocyanin content in black rice extract (Pedro *et al.*, 2016). Extraction with heat-assisted maceration technique based on a recent study (Nurhidajah *et al.*, 2022) was reported to be very effective in preparing black rice anthocyanin extract. Anthocyanin extracts can be degraded during the storage process. This is due to its low chemical stability to environmental conditions including temperature, light intensity, and pH (Cassidy, 2018; Fernandes *et al.*, 2014). The anthocyanin decomposition process occurs faster at high temperatures (Patras *et al.*, 2010). Anthocyanins are unstable at high light intensity. The release of the sugar group causes the anthocyanin aglycones that are formed to fade quickly when exposed to light (Janna *et al.*, 2006). Encapsulation techniques can overcome this weakness using a freeze dryer, spray dryer, and others (Bao *et al.*, 2019; Fang & Bhandari, 2010; Howard *et al.*, 2013).

Encapsulation using a spray dryer is an effective solution to protect and increase the stability of anthocyanin bioactive compounds (Kalušević *et al.*, 2017). This technique has been widely used in the food industry. In addition to being applicable, the spray drying (SD) method for thermosensitive products is due to a short contact time with a heat source (Lourenço *et al.*, 2020). The success of SD is influenced by several factors, including temperatures of inlet and outlet, feed flow rate, speed of the atomizer, air flow rate, and type as well as concentration of coating material (Patil-Gadhe & Pokharkar, 2014). One of the elements that will influence the encapsulated powder is the coating substance.

Mahdavi *et al.* (2014) reported maltodextrin (MDE) is extensively used as a coating material in anthocyanin encapsulation using the SD technique. MDE as a coating material with various advantages, including high solubility, low viscosity, and maintaining the stability of bioactive compounds (Suhag & Nanda, 2016). Recent research by Kalušević *et al.* (2017) reported that skimmed milk powder (SMP) reported that SMP coating material could encapsulate anthocyanin components at higher levels than maltodextrin. As a coating material, SMP has excellent encapsulation effectiveness and non-stick characteristics. This ingredient adds extra nutritional value in addition to being a source of protein, lactose, vitamins, and minerals (Bylaitė *et al.*, 2001; Jing & Giusti, 2005; Shamaei *et al.*, 2016). However, the use of SMP as a coating material in the encapsulation of black rice extract still has a weakness, namely the coating efficiency of phenolic and flavonoid components is still very low (Kalušević *et al.*, 2017).

Until now, there has been no research report on the combination of MDE and SMP as a coating material in the encapsulation of black rice extract. Based on this, it is necessary to review further the effect of the MDE-SMP combination as a coating material for black rice extract, the right combination is expected to produce coated anthocyanin powder with the best physical and chemical characteristics.

2. MATERIALS AND METHODS

2.1. Materials

The main ingredient used is black rice of the jellyteng variety obtained from organic rice farmers in Central Java. While the chemicals used are citric acid, gallic acid (C₇H₆O₅), ethanol, 2,2-Diphenyl-1-picrylhydrazyl (DPPH), potassium chloride, folin-

ciocalteu, sodium acetate, and sodium carbonate are some of the pro-analytical substances used. manufactured by Merck.

2.2. Black Rice Anthocyanin Extraction (Nurhidajah *et al.*, 2022)

Flour was made from pulverized black rice. Black rice flour sample (100 g) was mixed in a 1:10 (w/v) ratio with the solvents ethanol (56% v/v) and citrate (4.5% w/v). The extraction was carried out for 120 minutes with constant agitation (500 rpm) in waterbath at 50 °C. Using 100 mesh filter paper, the following procedure separates the liquid from the residue. The extracted ethanol was then evaporated at 55 °C using a rotary evaporator. The evaporation procedure was repeated until the ethanol solvent ceased pouring, yielding a final volume of 44% black rice extract. The black rice anthocyanin extract was kept at 4 °C in a dark glass bottle.

2.3. Anthocyanin Extract Encapsulation (Nurhidajah *et al.*, 2022)

MD and SMP coating materials emulsion with different ratios (100:0, 90:10, 80:20, 70:30, 60:40 and 50:50) were prepared at a concentration of 20% (w/v) with deionized water at room temperature. Anthocyanin extract was mixed with each coating material in a ratio of 1:1 (v/v). Each mixture was homogenized separately for 15 min at 3000 rpm. It was dried using a lab-scale spray drier with 120 ± 1 °C inlet air temperatures, and 80 ± 5 °C outlet air temperatures, 6.0 mL/min feed flow rate, and 1.5 bar feed pressure. The obtained coated anthocyanin powder was gathered and kept at -20°C until analyzed.

2.4. Anthocyanin Content (Yamuangmorn *et al.*, 2018)

To measure covered anthocyanin powder, a differential pH technique was employed. First, the sample (1 g) was dissolved in 1 mL of ethanol in a test tube. Next, two buffer solutions 1 mL of potassium chloride buffer, pH 1.0, and 1 mL of sodium acetate buffer, pH 4.5 were combined independently, and the test tube was incubated at room temperature (± 25 °C) for 15 min. The absorbance was read by spectrophotometer at 520 and 700 nm. The following equation was used to determine the extract sample's absorption value (A):

$$A = [(A_{520} - A_{700})_{\text{pH}1} - (A_{520} - A_{700})_{\text{pH}4.5}] \quad (1)$$

Then anthocyanin content (AC) of black rice extract was calculated according to Equation (2) and was expressed in mg/100g:

$$AC \text{ (mg/100 g)} = \frac{A \times MW \times DF \times V \times 100}{E \times l \times W} \quad (2)$$

where MW = 448.8 g/mol is molecular weight of cyanidin-3-glucoside, DF is dilution factor, V is volume of the sample mother liquor, E = 26900 l/mol cm is the measure of molar absorptivity of cyanidin-3-glucoside, and W is sample weight (g).

2.5. Total Phenolics

The Folin-Ciocalteu method, which was modified slightly from Pedro *et al.* (2016), was used to measure the total phenolic content (TPC). Sample (0.5 g) was mixed with 5 ml of 10% Folin-Ciocalteu reagent. The addition of 7.5% Na₂CO₃ as much as 4 mL then homogenized. The solution was incubated for 60 minutes at 25 ± 1 °C. Reference solution used gallic acid at a strength of 100–500 ppm. The absorbance was measured using a UV-Vis spectrometer set at 765 nm. TPC was calculated as mg GAE/100g of black rice.

2.6. Flavonoid Content

Flavonoid content was determined according to [Cai et al., \(2016\)](#), with modifications. Black rice extract was combined with 2.8 milliliters of purified water, 0.1 milliliters of 10% AlCl₃, 1.5 milliliters of ethanol, and 0.1 milliliters of 1 M CH₃COOK in a dark tube. The mixture was homogenized and kept for 30 minutes at room temperature (25 ± 1 °C). The blank solution was aquadest, and the sample's absorbance was measured with a UV-Vis spectrometer at 415 nm. The reference curve uses a quercetin solution in aquadest with a concentration range of 20-100 ppm. Thus, mg QE/100 g was stated as the black rice extract's flavonoid content.

2.7. Antioxidant Activities

Antioxidant activity was determined according to [Pedro et al. \(2016\)](#), with slight modifications. The sample (0,2 ml) was mixed with 1.5 ml DPPH (0.2 mM), then diluted by adding 3.5 ml of ethanol. The solution was incubated at 25 ± 1 °C for 60 minutes, then read at 517 nm. The following equation was used to determine it:

$$\% \text{ RSA} = \frac{\text{Abs blanko} \times \text{Abs test}}{\text{Abs blanko}} \times 100\% \quad (3)$$

2.8. pH value

The pH value was measuring according to [Park & Kim \(2016\)](#). Coated anthocyanin powder sample (4g) was mixed with 16 mL of distilled water in a vortex mixer for 1 min. The pH was recorded using instrument Hanna HI 2211 bench pH meters (Hanna instruments Ltd.), calibration solution using standard buffer.

2.9. Yield, Moisture Content and Water Activity (a_w)

Yield was obtained from the final weight percentage of coated anthocyanin powder (g) with the total solids of the sample (g). Determination of water content and water activity using analysis tools, each of which is a moisture analyzer (Shimadzu MOC63u) and a water activity analyzer (Rotronic Hygropalm-HP23-Aw-A) at room temperature.

2.10. Color Measurement

An appropriately adjusted Minolta CR-310 Chromameter was used to test the coated anthocyanin powder's color characteristics (Konica Minolta Business Solution Asia Pte Ltd). Additionally, the chroma, C° = [(a*² + b*²)^{1/2}], and the hue angle, H° = [tan⁻¹(b*/a*)], were calculated. Red, yellow, green, and blue are the four primary hues, while the C° differentiates between bright and dull colors ([Caparino et al., 2012](#)).

2.11. Statistical Analysis

All experiments were performed in 4 repetitions. Results were presented in terms of average value ± standard deviation. The effect of treatment factors was analyzed using one-way ANOVA, and differences among treatments was detected using Duncan's post hoc test. The significance level used is $p < 0.05$. Statistical analysis was conducted with the help of SPSS 22.0 software.

3. RESULTS AND DISCUSSION

3.1. Anthocyanin and pH Value of Capsules

The mean anthocyanin content of black rice coated anthocyanin powder obtained was 51.22-84.59 mg/100g (Figure 1a). This amount is higher than the research results

reported by [Prमितasari & Angelica \(2020\)](#), which only ranged from 2.00–12.00 mg/100g. The highest anthocyanin levels were obtained at a coating ratio of 50:50 (84.59 mg/100g). There was an increase in anthocyanin levels of 65% (51.22mg/100g) compared to a ratio of 100:0. SMP 50% as a coating material was able to trap more anthocyanins, resulting in a powder with the highest anthocyanin content, significantly different from all treatments. SMP generally contains b-casein components and has been reported to interact with Cyanidin-3-glucoside compounds from black soybean extract through reversible hydrogen bonds supported by an optimum pH ([Kalušević et al., 2017](#)). Previous studies also reported that casein protein interacts with malvidin-3-glucoside from grape extract through complex hydrogen bonds, which then form a hydrophobic reaction and effectively increase thermal stability so that the rate of anthocyanin degradation during drying can be minimized ([He et al., 2016](#)). The pH value of coated anthocyanin powder ranged from 2.60-2.95 (Figure 1b). The pH value increased with the addition of SMP in the coating material ratio because SMP had a neutral pH value. However, the increase in pH value is still in the anthocyanin stability range, at a value of 2-3 ([Sipahli et al., 2017](#)). Anthocyanins are more stable under acidic conditions in the form of a flavylum cation structure ([Khoo et al., 2017](#)).

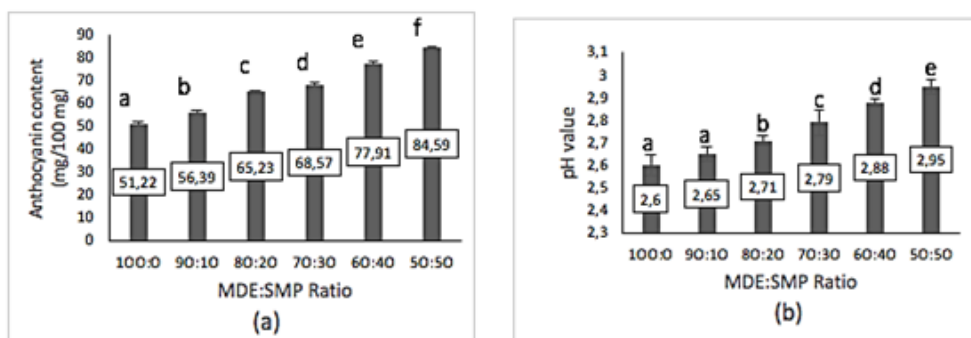


Figure 1. Anthocyanin and pH value coated anthocyanin powder based on MDE:SMP Ratio

3.2. Total Phenolics, Flavonoid, and Antioxidant Activities

Total phenolic and flavonoid content of coated anthocyanin powder is presented in Table 1. From the data, it is shown that the total phenolic and flavonoid content increases with the addition of SMP 50% in the ratio of the coating material, and significantly different with all treatments. B-casein in protein-derived encapsulation forms a more substantial complex with polyphenols, and this is due to the more hydrophobic nature of B-casein. ([Hasni et al., 2011](#)). The increased flavonoids are associated with the structure and bonds formed from proteins with flavonoid compounds. This result has previously been described by [Bohin et al. \(2012\)](#) on the flavonoid encapsulation of tea drinks. This underlies the increase in flavonoids when the ratio of SMP to coating material increases. In a previous study, [Kalušević et al. \(2017\)](#) reported the results of high antioxidant activity in coating black rice extract using SMP. Therefore, the antioxidant activity of coated anthocyanin powder is thought to be derived from the encapsulated anthocyanin, phenolic, and flavonoid components. In addition, the increase in antioxidant activity is also potential from the Maillard reaction, which produces melanoidin compounds with antioxidant activity. This reaction occurs in the protein component of SMP due to the high temperature during the drying process ([Wang et al., 2011](#)).

Table 1. Total phenolic, total flavonoid and antioxidant activity of coated anthocyanin powder based on MD and SMP ratio

MD:SMP Ratio	Total Phenolic (mg GAE/g)	Total Flavonoid (mg QE/g)	Antioxidant Activity (%)
100:0	0.27 ± 0.01 ^a	0.14 ± 0.02 ^a	25.07 ± 0.39 ^a
90:10	0.30 ± 0.01 ^b	0.16 ± 0.01 ^{ab}	26.52 ± 0.35 ^b
80:20	0.33 ± 0.01 ^c	0.17 ± 0.01 ^{bc}	29.23 ± 0.22 ^c
70:30	0.37 ± 0.01 ^d	0.18 ± 0.03 ^{bcd}	30.37 ± 0.38 ^d
60:40	0.38 ± 0.01 ^d	0.19 ± 0.02 ^{cd}	31.18 ± 0.48 ^e
50:50	0.41 ± 0.05 ^e	0.21 ± 0.02 ^d	31.71 ± 0.94 ^e

Note: All values are mean ± standard deviation 4 times repetition. Different superscript values showed significant differences with a 95% confidence level based on the ANOVA difference test and the LSD post hoc test.

3.3. AW, Moisture Content, and Yield of Coated Anthocyanin Powder

All samples showed an Aw value lower than 0.6 (0.49-0.58), so the coated anthocyanin powder in this study had an excellent aw value (Goyal *et al.*, 2015). A low aw value tends to result in extended shelf life. There is a positive tendency to increase the ratio of SMP as a coating material, SMP 50% is able to increase the aw of the product significantly. The water content values ranged from 0.91 to 1.13% (Table 2), the use of SMP 50% significantly made anthocyanin powder with the lowest water content. On the other hand, coatings with high MDE content produce high water content due to the nature of MDE, which quickly releases water (Kustyawati *et al.*, 2022). The product moisture content was low to prevent particle agglomeration, avoid the caking process, and improve powder dispersion and flowability (Daza *et al.*, 2016). Therefore, the powder from spray drying obtained both the aw level and the moisture content in this study met the requirements and was suitable for inclusion in the dry food matrix.

Table 2. Physical properties of coated anthocyanin powder based on MD and SMP ratio

MD:SMP Ratio	Yield (%)	Moisture content (%)	a _w (%)
100:0	33.51 ± 0.82 ^a	1.13 ± 0.03 ^e	0.58 ± 0.58 ^c
90:10	35.11 ± 0.75 ^a	1.08 ± 0.01 ^{de}	0.55 ± 0.55 ^{bc}
80:20	31.14 ± 0.68 ^a	1.04 ± 0.09 ^{cd}	0.54 ± 0.54 ^{bc}
70:30	34.06 ± 0.82 ^a	0.99 ± 0.01 ^{bc}	0.52 ± 0.52 ^{ab}
60:40	47.64 ± 0.73 ^b	0.96 ± 0.02 ^{cb}	0.51 ± 0.51 ^{ab}
50:50	51.29 ± 0.49 ^b	0.91 ± 0.05 ^a	0.49 ± 0.49 ^a

Note: All values are mean ± standard deviation 4 times repetition. Different superscript values showed significant differences with a 95% confidence level based on the ANOVA difference test and the LSD post hoc test.

The highest yield was significantly obtained from powders with a 50:50 coating material, which is in line with the results reported in a study comparing milk protein-based microparticles with other carriers (Belščak-Cvitanovic *et al.*, 2015). A combination of MDE and SMP coating materials resulted in good physical characteristics so that the coating material can be proposed as a coating material for the bioactive component of black rice extract.

3.4. Color Characteristic

Color is one of the indicators in assessing the quality of food products. During the production process, the color of black rice extract is expected to be maintained until a powder is obtained from drying. Therefore, observing the color measurement of coated anthocyanin powder after the encapsulation process is necessary. The results of chromatic color measurements are shown in Table 3. Based on the parameters a^* , b^* , and hue angle, it can be concluded that the color of the MD-SMP-based powder is red-violet with varying brightness levels. Among all coated anthocyanin powders, coating with a ratio of 50:50 differed significantly by having the lowest brightness value (40.58 ± 0.83) and the highest value for a^* and C^* with significant differences with other treatments. Indicating that this ratio produced the darkest powder with the highest portion of red color. Many reports have investigated the molecular level of the bond formed between casein-based coating materials and black rice anthocyanins on their stability from a thermodynamic point of view. The casein-anthocyanin bond is mainly stabilized through hydrogen bonds and hydrophobic interactions involving the aglycone and glucosyl moieties of the encapsulated molecule so that the color can be maintained during the drying process (Aprodu *et al.*, 2019).

Table 3. Color parameters of coated anthocyanin powder based on MD and SMP ratio

MD:SMP Ratio	L^*	a^*	b^*	C^*	H°	Color [#]
100:0	56.06 ± 0.79^d	26.83 ± 0.66^a	4.43 ± 0.31^a	27.20 ± 0.62^a	9.39 ± 0.81^a	R-V
90:10	55.46 ± 1.46^c	28.48 ± 0.12^b	4.30 ± 0.51^a	28.80 ± 0.16^b	8.58 ± 0.99^a	R-V
80:20	52.91 ± 0.61^c	28.71 ± 0.39^b	5.34 ± 0.50^{ab}	29.20 ± 0.35^b	10.55 ± 1.06^a	R-V
70:30	49.07 ± 0.82^b	35.78 ± 1.45^c	6.36 ± 1.56^b	36.36 ± 1.65^c	10.03 ± 2.13^a	R-V
60:40	48.06 ± 0.54^b	39.90 ± 0.47^d	9.71 ± 0.39^c	41.06 ± 0.39^d	13.68 ± 0.65^b	R-V
50:50	40.58 ± 0.83^a	43.64 ± 1.43^e	12.75 ± 0.83^d	45.47 ± 1.16^e	16.31 ± 1.50^c	R-V

Note: #) R-V = red-violet.

All values are mean \pm standard deviation 4 times repetition. Different superscript values showed significant differences with a 95% confidence level based on the ANOVA difference test and the LSD post hoc test.

4. CONCLUSIONS

It can be concluded that the best combination of coating materials in black rice encapsulation was obtained at MDE: SMP ratio of 50:50 with total anthocyanin levels obtained 84.59 mg/100g, with total phenol 0.41 mg GAE/g, total flavonoid 0.21 mg QE/g and antioxidant activity 31.71% RSA. The encapsulated product has an average value of L^* 40.5; a^* 43.64; b^* 12.75; Chroma 45.47; and Hue 16.31 with red-purple color types. Anthocyanin stability with a pH value of 2.95. Furthermore, it has an a_w of 0.49%, moisture content of 0.91%, and a yield of 51.29%. The combination of MDE and SMP as a coating material in the encapsulation process of black rice extract had a very significant effect on total anthocyanin levels, phenolics, total flavonoids, antioxidant activity, pH, color, a_w , water content, and yield.

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REFERENCES

- Aprodu, I., Milea, Ș.A., Anghel, R.M., Enachi, E., Barbu, V., Crăciunescu, O., Râpeanu, G., Bahrim, G.E., Oancea, A., & Stănciuc, N. (2019). New functional ingredients based on microencapsulation of aqueous anthocyanin-rich extracts derived from black rice (*Oryza sativa* L.). *Molecules*, **24**(18), 3389. <https://doi.org/10.3390/molecules24183389>
- Bao, C., Jiang, P., Chai, J., Jiang, Y., Li, D., Bao, W., Liu, B., Liu, B., Norde, W., & Li, Y. (2019). The delivery of sensitive food bioactive ingredients: Absorption mechanisms, influencing factors, encapsulation techniques and evaluation models. *Food Research International*, **120**, 130–140. <https://doi.org/10.1016/j.foodres.2019.02.024>
- Belščak-Cvitanović, A., Lević, S., Kalušević, A., Špoljarić, I., Đorđević, V., Komes, D., & Nedović, V. (2015). Efficiency assessment of natural biopolymers as encapsulants of green tea (*Camellia sinensis* L.) bioactive compounds by spray drying. *Food and Bioprocess Technology*, **8**(12), 2444–2460. <https://doi.org/10.1007/s11947-015-1592-y>
- Bohin, M.C., Vincken, J.P., van der Hijden, H.T.W.M., & Gruppen, H. (2012). Efficacy of food proteins as carriers for flavonoids. *J. Agric. Food Chem*, **60**, 4136–4143. <https://doi.org/10.1021/jf205292r>
- Bylaitė, E., Venskutonis, P.R., & Maždžprienė, R. (2001). Properties of caraway (*Carum carvi* L.) essential oil encapsulated into milk protein-based matrices. *European food research and technology*, **212**(6), 661–670. <https://doi.org/10.1007/s002170100297>
- Cai, Z., Qu, Z., Lan, Y., Zhao, S., Ma, X., Wan, Q., Jing, P., & Li, P. (2016). Conventional, ultrasound-assisted, and accelerated-solvent extractions of anthocyanins from purple sweet potatoes. *Food Chemistry*, **197**, 266–272. <https://doi.org/10.1016/j.foodchem.2015.10.110>
- Caparino, O.A., Tang, J., Nindo, C.I., Sablani, S.S., Powers, J.R., & Fellman, J.K. (2012). Effect of drying methods on the physical properties and microstructures of mango (*Philippine 'Carabao' var.*) powder. *Journal of Food Engineering*, **111**(1), 135-148. <https://doi.org/10.1016/j.jfoodeng.2012.01.010>
- Cassidy, A. (2018). Berry anthocyanin intake and cardiovascular health. *Molecular Aspects of Medicine*, **61**, 76–82. <https://doi.org/10.1016/j.mam.2017.05.002>
- Daza, L.D., Fujita, A., Fávoro-Trindade, C.S., Rodrigues-Ract, J.N., Granato, D., & Genovese, M.I. (2016). Effect of spray conditions on the physical properties of cagaita (*Eugenia dysenterica* DC.) fruit extracts. *Food and Bioprocess Technology*, **97**, 20-29. <https://doi.org/10.1016/j.fbp.2015.10.001>
- Fang, Z., & Bhandari, B. (2010). Encapsulation of polyphenols - A review. *Trends in Food Science and Technology*, **21**(10), 510–523. <https://doi.org/10.1016/j.tifs.2010.08.003>
- Fernandes, I., Faria, A., Calhau, C., de Freitas, V., & Mateus, N. (2014). Bioavailability of anthocyanins and derivatives. *Journal of Functional Foods*, **7**, 54-66. <http://dx.doi.org/10.1016/j.jff.2013.05.010>
- Goyal, A., Sharma, V., Sihag, M.K., Komar, S.K., Arora, S., Sabikhi, L., & Singh, A.K. (2015). Development and physico-chemical characterization of

- microencapsulated flaxseed oil powder: A functional ingredient for omega-3 fortification. *Powder Technology*, **286**, 527-537. <https://doi.org/10.1016/j.powtec.2015.08.050>
- Hasni, I., Bourassa, P., Hamdani, S., Samson, G., Carpentier, R., & Tajmir-Riahi, H.A. (2011). Interaction of milk α - and β -caseins with tea polyphenols. *Food chemistry*, **126**(2), 630-639. <https://doi.org/10.1016/j.foodchem.2010.11.087>
- He, Z., Xu, M., Zeng, M., Qin, F., & Chen, J. (2016). Interactions of milk α - and β -casein with malvidin-3-O-glucoside and their effects on the stability of grape skin anthocyanin extracts. *Food chemistry*, **199**, 314-322. <https://doi.org/10.1016/j.foodchem.2015.12.035>
- Hosoda, K., Sasahara, H., Matsushita, K., Tamura, Y., Miyaji, M., & Matsuyama, H. (2018). Anthocyanin and proanthocyanidin contents, antioxidant activity, and in situ degradability of black and red rice grains. *Asian-Australasian Journal of Animal Sciences*, **31**(8), 1213–1220. <https://doi.org/10.5713/ajas.17.0655>
- Howard, L.R., Brownmiller, C., Prior, R.L., & Mauromoustakos, A. (2013). Improved stability of chokeberry juice anthocyanins by β -cyclodextrin addition and refrigeration. *J. Agric. Food Chem*, **61**(3), 693-9. <https://doi.org/10.1021/jf3038314>.
- Janna, O., Khairul, A., Maizah, M., & Mohd, M.Y. (2006). Flower pigment analysis of *Melastoma malabatricum*, *Journal of African Biotechnology*, **5**(2), 170-174.
- Jing, P., & Giusti, M.M. (2005). Characterization of anthocyanin-rich waste from purple corncobs (*Zea mays L.*) and its application to color milk. *J. Agric. Food Chem*, **53** (22), 8775–8781. <https://doi.org/10.1021/jf051247o>
- Kalušević, A.M., Lević, S.M., Čalića, B.R., Milić, J.R., Pavlović, V.B., Bugarski, B.M., & Nedović, V.A. (2017). Effects of different carrier materials on physicochemical properties of microencapsulated grape skin extract. *Journal of Food Science and Technology*, **54**(11), 3411–3420. <https://doi.org/10.1007/s13197-017-2790-6>
- Khoo, H.E., Azlan, A., Tang, S.T., & Lim, S.M. (2017). Anthocyanidins and anthocyanins: Colored pigments as food, pharmaceutical ingredients, and the potential health benefits. *Food & Nutrition Research*, **61**, 1–21. <https://doi.org/10.1080/16546628.2017.1361779>
- Kong, S., Kim, D.J., Oh, S.K., Choi, I.S., Jeong, H.S., & Lee, J. (2012). Black rice bran as an ingredient in noodles: Chemical and functional evaluation. *Journal of Food Science*, **77**(3), 307-307. <https://doi.org/10.1111/j.1750-3841.2011.02590.x>
- Kustyawati, M.E., Sugiharto, R., & Rini, R. (2022). Microencapsulation of green capulaga (*Elettaria cardamomum*) essential oil with maltodextrin and its applications in coffee drink. *Jurnal Teknologi Pertanian Lampung*, **11**(3), 531-541. <http://dx.doi.org/10.23960/jtep-l.v11i3.531-541>
- Lourenço, S.C., Moldão-Martins, M., & Alves, V.D. (2020). Microencapsulation of pineapple peel extract by spray drying using maltodextrin, inulin, and arabic gum as wall matrices. *Foods*, **9**(6), 718. <https://doi.org/10.3390/foods9060718>
- Mahdavi, S.A., Jafari, S.M., Ghorbani, M., & Assadpoor, E. (2014). Spray-drying microencapsulation of anthocyanins by natural biopolymers: A review. *Drying Technology*, **32**(5), 509–518. <https://doi.org/10.1080/07373937.2013.839562>
- Nour, V., Stampar, F., Veberic, R., & Jakopic, J. (2013). Anthocyanins profile, total phenolics and antioxidant activity of black currant ethanolic extracts as influenced by genotype and ethanol concentration. *Food Chemistry*, **141**(2), 961–966. <https://doi.org/10.1016/j.foodchem.2013.03.105>

- Nurhidajah, Rosidi, A., Yonata, D., & Pranata, B. (2022). Optimizing extraction of functional compounds from Indonesian black rice using response surface methodology. *Food Research*, **6**(4), 83-91. [https://doi.org/10.26656/fr.2017.6\(4\).732](https://doi.org/10.26656/fr.2017.6(4).732)
- Park, S.Y., & Kim, H.Y. (2016). Effect of black rice powder levels on quality properties of emulsion-type sausage. *Korean Journal for Food Science of Animal Resources*, **36**(6), 737–743.
- Patil-Gadhe, A., & Pokharkar V. (2014). Single step spray drying method to develop proliposomes for inhalation: A systematic study based on quality by design approach. *Pulmonary Pharmacology and Therapeutics*, **27**(2), 197–207. <https://doi.org/10.1016/j.pupt.2013.07.006>
- Patras, A., Brunton, N.P., O'donnell, C., & Tiwari, B.K. (2010). Effect of thermal processing on anthocyanin stability in foods; Mechanisms and kinetics of degradation a review. *Trends in Food Science & Technology*, **21**, 3-11. <https://doi.org/10.1016/j.tifs.2009.07.004>
- Pedro, A.C., Granato, D., & Rosso, N.D. (2016). Extraction of anthocyanins and polyphenols from black rice (*Oryza sativa* L.) by modeling and assessing their reversibility and stability. *Food Chemistry*, **191**, 12–20. <https://doi.org/10.1016/j.foodchem.2015.02.045>
- Pramitasari, R., & Angelica, N. (2020). Ekstraksi, pengeringan semprot, dan analisis sifat fisikokimia antosianin beras hitam (*Oryza sativa* L.). *Jurnal Aplikasi Teknologi Pangan*, **9**(2), 83–94. <https://doi.org/10.17728/jatp.5889>
- Shamaei, S., Seiedlou, S.S., Aghbashlo, M., Tsotsas, E., & Kharaghani, A. (2017). Microencapsulation of walnut oil by spray drying: Effects of wall material and drying conditions on physicochemical properties of microcapsules. *Innovative food science & emerging technologies*, **39**, 101-112. <https://doi.org/10.1016/j.ifset.2016.11.011>
- Sipahli, S., Mohanlall, V., & Mellem, J.J. (2017). Stability and degradation kinetics of crude anthocyanin extracts from *H. sabdariffa*. *Food Science and Technology*, **37**(2), 209–215. <https://doi.org/10.1590/1678-457X.14216>
- Suhag, Y., & Nanda, V. (2016). Optimization for spray drying process parameters of nutritionally rich honey powder using response surface methodology. *Cogent Food and Agriculture*, **2**(1), 1–12. <https://doi.org/10.1080/23311932.2016.1176631>
- Wang, H.Y., Qian, H., Yao, W.E. (2011). Melanoidins produced by the Maillard reaction: reaction: structure and biological activity. *Food Chemistry*, **128**(3), 573-584. <https://doi.org/10.1016/j.foodchem.2011.03.075>
- Yamuangmorn, S., Dell, B., & Prom-u-thai, C. (2018). Effects of cooking on anthocyanin concentration and bioactive antioxidant capacity in glutinous and non-glutinous purple rice. *Rice Science*, **25**(5), 270–278. <https://doi.org/10.1016/j.rsci.2018.04.004>